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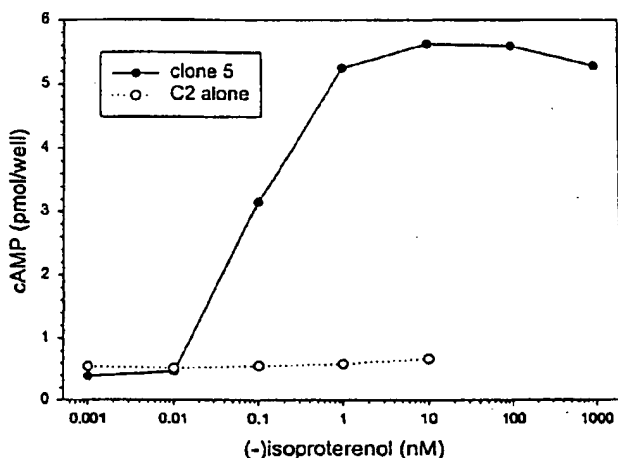
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(54) Title: IMPROVED SYSTEMS FOR SENSITIVE DETECTION OF G-PROTEIN COUPLED RECEPTOR AND ORPHAN RECEPTOR FUNCTION USING REPORTER ENZYME MUTANT COMPLEMENTATION

Agonist Stimulated cAMP Response in C2 Cells Expressing $\beta 2AR$ - $\beta gal\Delta\alpha$ 

(57) Abstract: Methods for detecting G-protein coupled receptor (GPCR) activity; methods for assaying GPCR activity; and methods for screening for GPCR ligands, G-protein-coupled receptor kinase (GRK) activity, and compounds that interact with components of the GPCR regulatory process are described. Included are methods for expanding ICASST technologies for assaying GPCR activity with applications for ligand fishing, and agonist or antagonist screening. These methods include: engineering serine/threonine phosphorylation sites into known or orphan GPCR open reading frames in order to increase the affinity of arrestin for the activated form of the GPCR or to increase the residence time of arrestin on the activated GPCR; engineering mutant arrestin proteins

that bind to activated GPCRs in the absence of G-protein coupled receptor kinases which may be limiting; and engineering mutant super arrestin proteins that have an increased affinity for activated GPCRs with or without phosphorylation. These methods are intended to increase the robustness of the GPCR/ICASST technology in situations in which G-protein coupled receptor kinases are absent or limiting, or in which the GPCR is not efficiently down-regulated or is rapidly resensitized (thus having a labile interaction with arrestin). Included are also more specific methods for using ICASST complementary enzyme fragments to monitor GPCR homo- and hetero- dimerization with applications for drug lead discovery and ligand and function discovery for orphan GPCRs.



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TITLE OF THE INVENTION**IMPROVED SYSTEMS FOR SENSITIVE DETECTION OF G-PROTEIN
COUPLED RECEPTOR AND ORPHAN RECEPTOR FUNCTION
USING REPORTER ENZYME MUTANT COMPLEMENTATION****BACKGROUND OF THE INVENTION**

This application is a continuation-in-part of U.S. Application Serial No.
09/654,499, filed September 1, 2000, which claims the benefit from Provisional
Application Serial No. 60/180,669, filed February 7, 2000. The entirety of U.S.
5 Application Serial No. 09/654,499 and Provisional Application Serial No.
60/180,669 are incorporated herein by reference.

Field of the Invention

The present invention relates to methods of detecting G-protein-coupled
10 receptor (GPCR) activity, and provides methods of assaying GPCR activity,
methods for screening for GPCR ligands, agonists and/or antagonists, methods for
screening natural and surrogate ligands for orphan GPCRs, and methods for
screening compounds that interact with components of the GPCR regulatory
process.

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Background of the Technology

The actions of many extracellular signals are mediated by the interaction of
G-protein- coupled receptors (GPCRs) and guanine nucleotide-binding regulatory
proteins (G-proteins). G-protein-mediated signaling systems have been identified in
20 many divergent organisms, such as mammals and yeast. The GPCRs represent a

large super family of proteins which have divergent amino acid sequences, but share common structural features, in particular, the presence of seven transmembrane helical domains. GPCRs respond to, among other extracellular signals, neurotransmitters, hormones, odorants and light. Individual GPCR types
5 activate a particular signal transduction pathway; at least ten different signal transduction pathways are known to be activated via GPCRs. For example, the beta 2-adrenergic receptor ($\beta 2AR$) is a prototype mammalian GPCR. In response to agonist binding, $\beta 2AR$ receptors activate a G-protein (G_s) which in turn stimulates adenylate cyclase activity and results in increased cyclic adenosine
10 monophosphate (cAMP) production in the cell.

The signaling pathway and final cellular response that result from GPCR stimulation depends on the specific class of G-protein with which the particular receptor is coupled (Hamm, "The Many Faces of G-Protein Signaling." J. Biol. Chem., 273:669-672 (1998)). For instance, coupling to the G_s class of G-proteins
15 stimulates cAMP production and activation of the Protein Kinase A and C pathways, whereas coupling to the G_i class of G-proteins down regulates cAMP. Other second messenger systems such as calcium, phospholipase C, and phosphatidylinositol 3 may also be utilized. As a consequence, GPCR signaling events have predominantly been measured via quantification of these second
20 messenger products.

The decrease of a response to a persistent stimulus is a widespread biological phenomenon. Signaling by diverse GPCRs is believed to be terminated by a uniform two-step mechanism. Activated receptor is first phosphorylated by a

GPCR kinase (GRK). An arrestin protein binds to the activated and phosphorylated receptor, thus blocking G-protein interaction. This process is commonly referred to as desensitization, a general mechanism that has been demonstrated in a variety of functionally diverse GPCRs. Arrestin also plays a part in regulating GPCR internalization and resensitization, processes that are heterogenous among different GPCRs (Oakley, et al., J. Biol. Chem., 274:32248-32257 (1999)). The interaction between an arrestin and GPCR in processes of internalization and resensitization is dictated by the specific sequence motif in the carboxyl terminus of a given GPCR. Only a subset of GPCRs, which possess clusters of three serine or threonine residues at the carboxyl termini, were found to co-traffick with the arrestins into the endocytic vesicles after ligand stimulation. The number of receptor kinases and arrestins involved in desensitization of GPCRs is rather limited.

A common feature of GPCR physiology is desensitization and recycling of the receptor through the processes of receptor phosphorylation, endocytosis and dephosphorylation (Ferguson, et al., "G-protein-coupled receptor regulation: role of G-protein-coupled receptor kinases and arrestins." Can. J. Physiol. Pharmacol., 74:1095-1110 (1996)). Ligand-occupied GPCRs can be phosphorylated by two families of serine/threonine kinases, the G-protein-coupled receptor kinases (GRKs) and the second messenger-dependent protein kinases such as protein kinase A and protein kinase C. Phosphorylation by either class of kinases serves to down-regulate the receptor by uncoupling it from its corresponding G-protein. GRK-phosphorylation also serves to down-regulate the receptor by recruitment of a

class of proteins known as the arrestins that bind the cytoplasmic domain of the receptor and promote clustering of the receptor into endocytic vesicles. Once the receptor is endocytosed, it will either be degraded in lysosomes or dephosphorylated and recycled back to the plasma membrane as a fully-functional receptor.

Binding of an arrestin protein to an activated receptor has been documented as a common phenomenon of a variety of GPCRs ranging from rhodopsin to β 2AR to the neurotensin receptor (Barak, et al., "A β -arrestin/Green Fluorescent Fusion Protein Biosensor for Detecting G-Protein-Coupled Receptor Activation," J. Biol. Chem., 272:27497-500 (1997)). Consequently, monitoring arrestin interaction with a specific GPCR can be utilized as a generic tool for measuring GPCR activation. Similarly, a single G-protein and GRK also partner with a variety of receptors (Hamm, et al. (1998) and Pitcher et al., "G-Protein-Coupled Receptor Kinases," Annu. Rev. Biochem., 67:653-92 (1998)), such that these protein/protein interactions may also be monitored to determine receptor activity.

Many therapeutic drugs in use today target GPCRs, as they regulate vital physiological responses, including vasodilation, heart rate, bronchodilation, endocrine secretion and gut peristalsis. See, e.g., Lefkowitz et al., Annu. Rev. Biochem., 52:159 (1983). Some of these drugs mimic the ligand for this receptor. Other drugs act to antagonize the receptor in cases when disease arises from spontaneous activity of the receptor.

Efforts such as the Human Genome Project are identifying new GPCRs ("orphan" receptors) whose physiological roles and ligands are unknown. It is estimated that several thousand GPCRs exist in the human genome.

Various approaches have been used to monitor intracellular activity in response to a stimulant, e.g., enzyme-linked immunosorbent assay (ELISA);
5 Fluorescence Imaging Plate Reader assay (FLIPR™, Molecular Devices Corp., Sunnyvale, CA); EVOscreen™, EVOTEC™, Evotec Biosystems GmbH, Hamburg, Germany; and techniques developed by CELLOMICS™, Cellomics, Inc., Pittsburgh, PA.

10 Germino et al., "Screening for in vivo protein-protein interactions." Proc. Natl. Acad. Sci., 90(3):933-937 (1993), discloses an *in vivo* approach for the isolation of proteins interacting with a protein of interest.

Phizicky et al., "Protein-protein interactions: methods for detection and analysis." Microbiol. Rev., 59(1): 94-123 (1995), discloses a review of
15 biochemical, molecular biological and genetic methods used to study protein-protein interactions.

Offermanns et al., "Gα₁₅ and Gα₁₆ Couple a Wide Variety of Receptors to Phospholipase C." J. Biol. Chem., 270(25):15175-15180 (1995), discloses that Gα₁₅ and Gα₁₆ can be activated by a wide variety of G-protein-coupled receptors.
20 The selective coupling of an activated receptor to a distinct pattern of G-proteins is regarded as an important requirement to achieve accurate signal transduction. Id.

Barak et al., "A β-arrestin/Green Fluorescent Protein Biosensor for Detecting G Protein-coupled Receptor Activation." J. Biol. Chem., 272(44):27497-

27500 (1997) and U.S. Patents Nos. 5,891,646 and 6,110,693 disclose the use of a β -arrestin/green fluorescent fusion protein (GFP) for imaging protein translocation upon stimulation of GPCR with optical devices.

Each of the references described above has drawbacks. For example,

- 5 • The prior art methodologies require over-expression of the proteins, which could cause artifact and tip the balance of cellular regulatory machineries.
- The prior art visualization or imaging assays are low throughput and lack thorough quantification. Therefore, they are not suitable for
- 10 high throughput pharmacological and kinetic assays.

In addition, many of the prior art assays require isolation of the GPCR rather than observation of the GPCR in a cell. There thus exists a need for improved methods for monitoring GPCR function.

15

SUMMARY OF THE INVENTION

The present invention provides modifications to the disclosure in U.S. Application Serial No. 09/654,499. In particular, the present invention is directed to modifications of the below aspects of the invention to further enhance assay sensitivity. The modifications include the use of genetically modified arrestins that

20 exhibit enhanced binding to activated GPCR regardless of whether the GPCR is phosphorylated or non-phosphorylated; the use of a serine/threonine cluster strategy to facilitate screening assays for orphan receptors that do not possess this

structural motif on their own; and the use of a combination of the above modifications to achieve even more enhanced detection.

A first aspect of the present invention is a method that monitors GPCR function proximally at the site of receptor activation, thus providing more information for drug discovery purposes due to fewer competing mechanisms. Activation of the GPCR is measured by a read-out for interaction of the receptor with a regulatory component such as arrestin, G-protein, GRK or other kinases, the binding of which to the receptor is dependent upon agonist occupation of the receptor. The present invention involves the detection of protein/protein interaction by complementation of mutant reporter enzymes.

Binding of arrestin to activated GPCR is a common process in the first step of desensitization that has been demonstrated for most, if not all, GPCRs studied so far. Measurement of GPCR interaction with arrestin via mutant enzyme complementation (i.e., ICAST) provides a more generic assay technology applicable for a wide variety of GPCRs and orphan receptors.

A further aspect of the present invention is a method of assessing GPCR pathway activity under test conditions by providing a test cell that expresses a GPCR, e.g., muscarinic, adrenergic, dopamine, angiotensin or endothelin, as a fusion protein to a mutant reporter enzyme and interacting a protein in the GPCR pathway, e.g., G-protein, arrestin or GRK, as a fusion protein with a complementing mutant reporter enzyme. When test cells are exposed to a known agonist to the target GPCR under test conditions, activation of the GPCR will be

monitored by complementation of the reporter enzyme. Increased reporter enzyme activity reflects interaction of the GPCR with its interacting protein partner.

A further aspect of the present invention is a method of assessing GPCR pathway activity in the presence of a test arrestin, e.g., β -arrestin.

5 A further aspect of the present invention is a method of assessing GPCR pathway activity in the presence of a test G-protein.

A further aspect of the present invention is a method of assessing GPCR pathway activity upon exposure of the test cell to a test ligand.

A further aspect of the present invention is a method of assessing GPCR
10 activity upon co-expression in the test cell of a second receptor. The second receptor could be the same GPCR or orphan receptor (i.e., homo-dimerization), a different GPCR or orphan receptor (i.e., hetero-dimerization) or could be a receptor of another type.

A further aspect of the present invention is a method for screening for a
15 ligand or agonist to an orphan GPCR. The ligand or agonist could be contained in natural or synthetic libraries or mixtures or could be a physical stimulus. A test cell is provided that expresses the orphan GPCR as a fusion protein with a mutant reporter enzyme, e.g., a β -galactosidase mutant, and, for example, an arrestin or mutant form of arrestin as a fusion protein with a complementing mutant reporter
20 enzyme, e.g., another β -galactosidase mutant. The interaction of the arrestin with the orphan GPCR upon receptor activation is measured by enzymatic activity of the complemented reporter enzyme. The test cell is exposed to a test compound, and an increase in reporter enzyme activity indicates the presence of a ligand or agonist.

A further aspect of the present invention is a method for screening a protein of interest, for example, an arrestin protein (or mutant form of the arrestin protein) for the ability to bind to a phosphorylated, or activated, GPCR. A test cell is provided that expresses a GPCR as a fusion protein with a mutant reporter enzyme, e.g., a β -galactosidase mutant, and contains arrestin (or a mutant form of arrestin) as a fusion protein with a complementing mutant reporter enzyme, e.g., another β -galactosidase mutant. The interaction of arrestin with the GPCR upon receptor activation is measured by enzymatic activity of the complemented reporter enzyme. The test cell is exposed to a known GPCR agonist and then reporter enzyme activity is detected. Increased reporter enzyme activity indicates that the β -arrestin molecule can bind to phosphorylated, or activated, GPCR in the test cell.

A further aspect of the present invention is a method to screen for an agonist to a specific GPCR. The agonist could be contained in natural or synthetic libraries or could be a physical stimulus. A test cell is provided that expresses a GPCR as a fusion protein with a mutant reporter enzyme, e.g., a β -galactosidase mutant, and, for example, an arrestin as a fusion protein with a complementing mutant reporter enzyme, e.g., another β -galactosidase mutant. The interaction of arrestin with the GPCR upon receptor activation is measured by enzymatic activity of the complemented reporter enzyme. The test cell is exposed to a test compound, and an increase in reporter enzyme activity indicates the presence of an agonist. The test cell may express a known GPCR or a variety of known GPCRs, or may express an unknown GPCR or a variety of unknown GPCRs. The GPCR may be, for example, an odorant GPCR or a β AR GPCR.

A further aspect of the present invention is a method for screening a test compound for GPCR antagonist activity. A test cell is provided that expresses a GPCR as a fusion protein with a mutant reporter enzyme, e.g., a β -galactosidase mutant, and, for example, an arrestin as a fusion protein with a complementing mutant reporter enzyme, e.g., another β -galactosidase mutant. The interaction of arrestin with the GPCR upon receptor activation is measured by enzymatic activity of the complemented reporter enzyme. The test cell is exposed to a test compound, and an increase in reporter enzyme activity indicates the presence of an agonist. The cell is exposed to a test compound and to a GPCR agonist, and reporter enzyme activity is detected. When exposure to the agonist occurs at the same time as or subsequent to exposure to the test compound, a decrease in reporter enzyme activity after exposure to the test compound indicates that the test compound has antagonist activity to the GPCR.

A further aspect of the present invention is a method of screening a sample solution for the presence of an agonist, antagonist or ligand to a GPCR. A test cell is provided that expresses GPCR as a fusion protein with a mutant reporter enzyme, e.g., a β -galactosidase mutant, and contains, for example, a β -arrestin as a fusion protein with a complementing reporter, e.g., another β -galactosidase mutant. The test cell is exposed to a sample solution, and reporter enzyme activity is assessed. Changed reporter enzyme activity after exposure to the sample solution indicates the sample solution contains an agonist, antagonist or ligand for a GPCR expressed in the cell.

A further aspect of the present invention is a method of screening a cell for the presence of a GPCR. According to this aspect, an arrestin fusion protein with a mutant reporter enzyme and a GPCR downstream signaling fusion protein with a mutant reporter enzyme are employed to detect GPCR action. A modification of this aspect of the invention can be employed to provide a method of screening a plurality of cells for those cells which contain a GPCR. According to this aspect, a plurality of cells containing a conjugate comprising a β -arrestin protein as a fusion protein with a reporter enzyme are provided; the plurality of cells are exposed to a GPCR agonist; and activity of reporter enzyme activity is detected. An increase in reporter enzymatic activity after exposure to the GPCR agonist indicates β -arrestin protein binding to a GPCR, thereby indicating that the cell contains a GPCR responsive to the GPCR agonist.

A further aspect of the invention is a method for mapping GPCR-mediated signaling pathways. For instance, the system could be utilized to monitor interaction of c-src with β -arrestin-1 upon GPCR activation. Additionally, the system could be used to monitor protein/protein interactions involved in cross-talk between GPCR signaling pathways and other pathways such as that of the receptor tyrosine kinases or Ras/Raf. According to this aspect, a test cell is provided that expresses a GPCR or other related protein with a mutant reporter enzyme, e.g., a β -galactosidase mutant, and contains a protein from another pathway as a fusion protein with a complementing mutant reporter enzyme, e.g., another β -galactosidase mutant. Increased reporter enzymatic activity indicates protein/protein interaction.

A further aspect of the invention is a method for monitoring homo- or hetero- dimerization of GPCRs upon agonist or antagonist stimulation. Increasing evidence indicates that GPCR dimerization is important for biological activity (AbdAlla, et al., "AT1-receptor heterodimers show enhanced G-protein activation and altered receptor sequestration." Nature, 407:94-98 (2000); Bockaert, et al., "Molecular tinkering of G protein-coupled receptors: an evolutionary success." EMBO J. 18:1723-29 (1999)). Jordan, et al., "G-protein-coupled receptor heterodimerization modulates receptor function." Nature, 399:697-700 (1999), demonstrated that two non-functional opioid receptors, κ and δ , heterodimerize to form a functional receptor. Gordon et al., "Dopamine D2 receptor dimers and receptor blocking peptides." Bioch. Biophys. Res. Commun. 227:200-204 (1996), showed different pharmacological properties associated with the monomeric and dimeric forms of Dopamine receptor D2. The D2 receptors exist either as monomers that are selective targets for spiperone or as dimer forms that are targets for nemonapride. Herbert, et al., "A peptide derived from a β 2-adrenergic receptor transmembrane domain inhibits both receptor dimerization and activation." J.B.C. 271:16384-92 (1996), demonstrated that the agonist stimulation was found to stabilize the dimeric state of the receptor, whereas inverse agonists favored the monomeric form. Indeed, the same study showed that a peptide corresponding to the sixth transmembrane domain of the β 2-adrenergic receptor inhibited both receptor dimerization and activation. Further, Angers et al., Detection of beta-2-adrenergic receptor dimerization in living cells using bioluminescence resonance energy transfer, Proc. Natl. Acad. Sci. USA, 97(7):3684-3689, discloses the use of

β 2-adrenergic receptor fusion proteins (i.e., β 2-adrenergic receptor fused to luciferase and β 2-adrenergic receptor fused to an enhanced red-shifted green fluorescent protein) to study β 2-adrenergic receptor dimerization.

GPCR dimerization in the context of cellular physiology and pharmacology can be monitored in accordance with the invention. For example, β -galactosidase complementation can be measured in test cells that co-express GPCR fusion proteins of β -galactosidase mutant enzymes, e.g., GPCR₁ $\Delta\alpha$ and GPCR₂ $\Delta\omega$ (FIGURE 27). According to this aspect, the interconversion between monomeric to dimeric forms of the GPCRs or orphan receptors can be measured by mutant reporter enzyme complementation. FIGURE 27 illustrates a test cell co-expressing GPCR or an orphan receptor as a fusion protein with $\Delta\alpha$ form of β -galactosidase mutant (e.g., GPCR₁ $\Delta\alpha$), and the same GPCR or orphan receptor as a fusion protein with $\Delta\omega$ form of β -galactosidase mutant (e.g., GPCR₁ $\Delta\omega$). Formation of the GPCR homodimer is reflected by formation of an active enzyme, which can be measured by enzyme activity assays, such as the Gal-Screen™ assay. Similarly, hetero-dimerization between two distinct GPCRs, or two distinct orphan receptors, or between one known GPCR and one orphan receptor can be analyzed in test cells co-expressing two fusion proteins, e.g., GPCR₁ $\Delta\alpha$ and GPCR₂ $\Delta\omega$. The increased β -galactosidase activity indicates that the two receptors can form a heterodimer.

A further aspect of the invention is a method of monitoring the interconversion between the monomeric and dimeric form of GPCRs under the influence of agonist or antagonist treatment. The test receptor(s) can be between the same GPCR or orphan receptor (homodimer), or between two distinct GPCRs

or orphan receptors (heterodimer). The increased β -galactosidase activity after treatment with a compound means that the compound binds to and/or stabilizes the dimeric form of the receptor. The decreased β -galactosidase activity after treatment with a compound means that the compound binds to and/or stabilizes the monomeric form of the receptor.

A further aspect of the invention is a method of screening a cell for the presence of a GPCR responsive to a GPCR agonist. A cell is provided that contains protein partners that interact downstream in the GPCR's pathway. The protein partners are expressed as fusion proteins to the mutant, complementing enzyme and are used to monitor activation of the GPCR. The cell is exposed to a GPCR agonist and then enzymatic activity of the reporter enzyme is detected. Increased reporter enzyme activity indicates that the cell contains a GPCR responsive to the agonist.

The present invention involves the use of a combination of proprietary technologies (including ICASTTM, Intercistronic Complementation Analysis Screening Technology, Gal-ScreenTM, etc.) to monitor protein/protein interactions in GPCR signaling. As disclosed in U.S. Application Serial No. 09/654,499, the method of the invention in part involves using ICASTTM, which in turn involves the use of two inactive β -galactosidase mutants, each of which is fused with one of two interacting target protein pairs, such as a GPCR and an arrestin. The formation of an active β -galactosidase complex is driven by interaction of the target proteins. In this system, β -galactosidase activity can be detected using, e.g., the Gal-ScreenTM assay system, wherein direct cell lysis is combined with rapid

ultrasensitive chemiluminescent detection of β -galactosidase reporter enzyme.

This system uses, e.g., a Galacton-*Star*® chemiluminescent substrate for measurement in a luminometer as a read out of GPCR activity.

FIGURE 23 is a schematic depicting the use of the complementation
5 technology in the method of the present invention. FIGURE 23 shows two inactive β -galactosidase mutants that become active when they are forced together by specific interactions between the fusion partners of an arrestin molecule and an activated GPCR or orphan receptor. This assay technology will be especially useful in high throughput screening assays for ligand fishing for orphan receptors, a
10 process called de-orphaning. As illustrated in FIGURE 28, a β -galactosidase fusion protein of an orphan receptor (e.g., GPCR_{orphan} $\Delta\alpha$) is co-expressed in the test cell with a fusion protein of β -arrestin (e.g., β -Arr $\Delta\omega$). When the test cell is subjected to compounds, which could be natural or synthetic, the increased β -galactosidase activity means the compound is either a natural or surrogate ligand
15 for this GPCR. The same assay system can be used to find drug leads for the new GPCRs. The increased β -galactosidase activity in the test cell after treatment indicates the agonist activity of the compound. The decreased β -galactosidase activity in the test cell indicates antagonist activity or inverse agonist activity of the compound. In addition, the method of the invention could be used to monitor
20 GPCR-mediated signaling pathways via other downstream signaling components such as G-proteins, GRKs or the proto-oncogene c-Src.

The invention is achieved in part by using ICAST™ protein/protein interaction screening to map signaling pathways. This technology is applicable to

a variety of known and unknown GPCRs with diverse functions. They include, but are not limited to, the following sub-families of GPCRs:

(a) receptors that bind to amine-like ligands-Acetylcholine muscarinic receptor (M1 to M5), alpha and beta Adrenoceptors, Dopamine receptors (D1, D2, D3 and D4), Histamine receptors (H1 and H2), Octopamine receptor and Serotonin receptors (5HT1, 5HT2, 5HT4, 5HT5, 5HT6, 5HT7);

(b) receptors that bind to a peptide ligand-Angiotensin receptor, Bombesin receptor, Bradykinin receptor, C-C chemokine receptors (CCR1 to CCR8, and CCR10), C-X-C type Chemokine receptors (CXC-R5), Cholecystokinin type A receptor, CCK type receptors, Endothelin receptor, Neurotesin receptor, FMLP-related receptors, Somatostatin receptors (type 1 to type 5) and Opioid receptors (type D, K, M, X);

(c) receptors that bind to hormone proteins-Follic stimulating hormone receptor, Thyrotrophin receptor and Lutropin-choriogonadotropic hormone receptor;

(d) receptors that bind to neurotransmitters-substance P receptor, Substance K receptor and neuropeptide Y receptor;

(e) Olfactory receptors-Olfactory type 1 to type 11, Gustatory and odorant receptors;

(f) Prostanoid receptors-Prostaglandin E2 (EP1 to EP4 subtypes), Prostacyclin and Thromboxane;

(g) receptors that bind to metabotropic substances-Metabotropic glutamate group I to group III receptors;

(h) receptors that respond to physical stimuli, such as light, or to chemical stimuli, such as taste and smell; and

(i) orphan GPCRs-the natural ligand to the receptor is undefined.

Use of the ICAST™ technology in combination with the invention provides many benefits to the GPCR screening process, including the ability to monitor protein interactions in any sub-cellular compartment-membrane, cytosol and nucleus; the ability to achieve a more physiologically relevant model without requiring protein overexpression; and the ability to achieve a functional assay for receptor binding allowing high information content.

10

BRIEF DESCRIPTION OF THE DRAWINGS

FIGURE 1. Cellular expression levels of $\beta 2$ adrenergic receptor ($\beta 2AR$) and β -arrestin-2 ($\beta Arr2$) in C2 clones. Quantification of β -galactosidase (β -gal) fusion protein was performed using antibodies against β -gal and purified β -gal protein in a titration curve by a standardized ELISA assay. Figure 1A shows expression levels of $\beta 2AR$ - $\beta gal\Delta\alpha$ clones (in expression vector pICAST ALC). Figure 1B shows expression levels of $\beta Arr2$ - $\beta gal\Delta\omega$ in expression vector pICAST OMC4 for clones 9-3, -7, -9, -10, -19 and -24, or in expression vector pICAST OMN4 for clones 12-4, -9, -16, -18, -22 and -24.

FIGURE 2. Receptor $\beta 2AR$ activation was measured by agonist-stimulated cAMP production. C2 cells expressing pICAST ALC $\beta 2AR$ (clone 5) or parental cells were treated with increasing concentrations of (-)-isoproterenol and 0.1mM

IBMX. The quantification of cAMP level was expressed as pmol/well.

FIGURE 3. Interaction of activated receptor $\beta 2AR$ and arrestin can be measured by β -galactosidase complementation. Figure 3A shows a time course of β -galactosidase activity in response to agonist (-)isoproterenol stimulation in C2
5 expressing $\beta 2AR$ - $\beta gal\Delta\alpha$ ($\beta 2AR$ alone, in expression vector pICAST ALC), or a pool of doubly transduced C2 co-expressing $\beta 2AR$ - $\beta gal\Delta\alpha$ and $\beta Arr2$ - $\beta gal\Delta\omega$ (in expression vectors pICAST ALC and pICAST OMC and clones isolated from the same pod (43-1, 43-2, 43-7 and 43-8)). Figure 3B shows a time course of β -galactosidase activity in response to agonist (-)isoproterenol stimulation in C2 cells
10 expressing $\beta 2AR$ - $\beta gal\Delta\alpha$ alone (in expression vector pICAST ALC) and C2 clones co-expressing $\beta 2AR$ - $\beta gal\Delta\alpha$ and $\beta Arr1$ - $\beta gal\Delta\omega$ (in expression vectors ICAST ALC and pICAST OMC).

FIGURE 4. Agonist dose response for interaction of $\beta 2AR$ and arrestin can be measured by β -galactosidase complementation. Figure 4A shows a dose
15 response to agonists (-)isoproterenol and procaterol in C2 cells co-expressing $\beta 2AR$ - $\beta gal\Delta\alpha$ and $\beta Arr2$ - $\beta gal\Delta\omega$ fusion constructs. Figure 4B shows a dose response to agonists (-)isoproterenol and procaterol in C2 cells co-expressing $\beta 2AR$ - $\beta gal\Delta\alpha$ and $\beta Arr1$ - $\beta gal\Delta\omega$ fusion constructs.

FIGURE 5. Antagonist mediated inhibition of receptor activity can be
20 measured by β -galactosidase complementation in cells co-expressing $\beta 2AR$ - $\beta gal\Delta\alpha$ and βArr - $\beta gal\Delta\omega$. Figure 5A shows specific inhibition with adrenergic

antagonists ICI-118,551 and propranolol of β -galactosidase activity in C2 clones co-expressing β 2AR- β gal $\Delta\alpha$ and β Arr2- β gal $\Delta\omega$ fusion constructs after incubation with agonist (-)isoproterenol. Figure 5B shows specific inhibition of β -galactosidase activity with adrenergic antagonists ICI-118,551 and propranolol in C2 clones co-expressing β 2AR- β gal $\Delta\alpha$ and β Arr1- β gal $\Delta\omega$ fusion constructs in the presence of agonist (-)isoproterenol.

FIGURE 6. C2 cells expressing adenosine receptor A2a show cAMP induction in response to agonist (CGS-21680) treatment. C2 parental cells and C2 cells co-expressing A2aR- β gal $\Delta\alpha$ and β Arr1- β gal $\Delta\omega$ as a pool or as selected clones (47-2 and 47-13) were measured for agonist-induced cAMP response (pmol/well).

FIGURE 7. Agonist stimulated cAMP response in C2 cells co-expressing Dopamine receptor D1 (D1- β gal $\Delta\alpha$) and β -arrestin-2 (β Arr2- β gal $\Delta\omega$). The clone expressing β Arr2- β gal $\Delta\omega$ (Arr2 alone) was used as a negative control in the assay. Cells expressing D1- β gal $\Delta\alpha$ in addition to β Arr2- β gal $\Delta\omega$ responded agonist treatment (3-hydroxytyramine hydrochloride at 3 μ M). D1(PIC2) or D1(PIC3) designate D1 in expression vector pICAST ALC2 or pICAST ALC4, respectively.

FIGURE 8. Variety of mammalian cell lines can be used to generate stable cells for monitoring GPCR and arrestin interactions. FIGURE 8A, FIGURE 8B and FIGURE 8C show the examples of HEK 293, CHO and CHW cell lines co-expressing adrenergic receptor β 2AR and arrestin fusion proteins of β -

galactosidase mutants. The β -galactosidase activity was used to monitor agonist-induced interaction of β 2AR and arrestin proteins.

FIGURE 9. Beta-gal complementation can be used to monitor β 2
adrenergic receptor homo-dimerization. FIGURE 9A shows β -galactosidase
5 activity in HEK 293 clones co-expressing β 2AR- β gal $\Delta\alpha$ and β 2AR- β gal $\Delta\omega$.
FIGURE 9B shows a cAMP response to agonist (-)-isoproterenol in HEK 293
clones co-expressing β 2AR- β gal $\Delta\alpha$ and β 2AR- β gal $\Delta\omega$. HEK293 parental cells
were included in the assays as negative controls.

FIGURE 10A. pICAST ALC: Vector for expression of β -gal $\Delta\alpha$ as a C-
10 terminal fusion to the target protein. This construct contains the following
features: MCS, multiple cloning site for cloning the target protein in frame with the
 β -gal $\Delta\alpha$; GS Linker, (GGGGS) $_n$; NeoR, neomycin resistance gene; IRES, internal
ribosome entry site; ColE1ori, origin of replication for growth in *E. coli*;
5'MoMuLV LTR and 3'MoMuLV LTR, viral promoter and polyadenylation
15 signals from the Moloney Murine leukemia virus.

FIGURE 10B. Nucleotide sequence for pICAST ALC.

FIGURE 11A. pICAST ALN: Vector for expression of β -gal $\Delta\alpha$ as an N-
terminal fusion to the target protein. This construct contains the following
features: MCS, multiple cloning site for cloning the target protein in frame with the
20 β -gal $\Delta\alpha$; GS Linker, (GGGGS) $_n$; NeoR, neomycin resistance gene; IRES, internal
ribosome entry site; ColE1ori, origin of replication for growth in *E. coli*;

5'MoMuLV LTR and 3'MoMuLV LTR, viral promoter and polyadenylation signals from the Moloney Murine leukemia virus.

FIGURE 11B. Nucleotide sequence for pICAST ALN.

FIGURE 12A. pICAST OMC: Vector for expression of β -gal $\Delta\omega$ as a C-terminal fusion to the target protein. This construct contains the following features: MCS, multiple cloning site for cloning the target protein in frame with the β -gal $\Delta\omega$; GS Linker, (GGGGS)_n; Hygro, hygromycin resistance gene; IRES, internal ribosome entry site; ColE1ori, origin of replication for growth in *E. coli*; 5'MoMuLV LTR and 3'MoMuLV LTR, viral promoter and polyadenylation signals from the Moloney Murine leukemia virus.

FIGURE 12B. Nucleotide sequence for pICAST OMC.

FIGURE 13A. pICAST OMN: Vector for expression of β -gal $\Delta\omega$ as an N-terminal fusion to the target protein. This construct contains the following features: MCS, multiple cloning site for cloning the target protein in frame with the β -gal $\Delta\omega$; GS Linker, (GGGGS)_n; Hygro, hygromycin resistance gene; IRES, internal ribosome entry site; ColE1ori, origin of replication for growth in *E. coli*; 5'MoMuLV LTR and 3'MoMuLV LTR, viral promoter and polyadenylation signals from the Moloney Murine leukemia virus.

FIGURE 13B. Nucleotide sequence for pICAST OMN.

FIGURE 14. pICAST ALC β Arr2: Vector for expression of β -gal $\Delta\alpha$ as a C-terminal fusion to β -arrestin-2. The coding sequence of human β -arrestin-2 (Genebank Accession Number: NM_004313) was cloned in frame to β -gal $\Delta\alpha$ in a

pICAST ALC vector.

FIGURE 15. pICAST OMC β Arr2: Vector for expression of β -gal $\Delta\omega$ as a C-terminal fusion to β -arrestin-2. The coding sequence of human β -arrestin-2 (Genebank Accession Number: NM_004313) was cloned in frame to β -gal $\Delta\omega$ in a pICAST OMC vector.

FIGURE 16. pICAST ALC β Arr1: Vector for expression of β -gal $\Delta\alpha$ as a C-terminal fusion to β -arrestin-1. The coding sequence of human β -arrestin-1 (Genebank Accession Number: NM_004041) was cloned in frame to β -gal $\Delta\alpha$ in a pICAST ALC vector.

FIGURE 17. pICAST OMC β Arr1: Vector for expression of β -gal $\Delta\omega$ as a C-terminal fusion to β -arrestin-1. The coding sequence of human β -arrestin-1 (Genebank Accession Number: NM_004041) was cloned in frame to β -gal $\Delta\omega$ in a pICAST OMC vector.

FIGURE 18. pICAST ALC β 2AR: Vector for expression of β -gal $\Delta\alpha$ as a C-terminal fusion to β 2 Adrenergic Receptor. The coding sequence of human β 2 Adrenergic Receptor (Genebank Accession Number: NM_000024) was cloned in frame to β -gal $\Delta\alpha$ in a pICAST ALC vector.

FIGURE 19. pICAST OMC β 2AR: Vector for expression of β -gal $\Delta\omega$ as a C-terminal fusion β 2 Adrenergic Receptor. The coding sequence of human β 2 Adrenergic Receptor (Genebank Accession Number: NM_000024) was cloned in frame to β -gal $\Delta\omega$ in a pICAST OMC vector.

FIGURE 20. pICAST ALC A2aR: Vector for expression of β -gal $\Delta\alpha$ as a C-terminal fusion to Adenosine 2a Receptor. The coding sequence of human Adenosine 2a Receptor (Genebank Accession Number: NM_000675) was cloned in frame to β -gal $\Delta\alpha$ in a pICAST ALC vector.

5 FIGURE 21. pICAST OMC A2aR: Vector for expression of β -gal $\Delta\omega$ as a C-terminal fusion to Adenosine 2a Receptor. The coding sequence of human Adenosine 2a Receptor (Genebank Accession Number: NM_000675) was cloned in frame to β -gal $\Delta\omega$ in a pICAST OMC vector.

10 FIGURE 22. pICAST ALC D1: Vector for expression of β -gal $\Delta\alpha$ as a C-terminal fusion to Dopamine D1 Receptor. The coding sequence of human Dopamine D1 Receptor (Genebank Accession Number: X58987) was cloned in frame to β -gal $\Delta\alpha$ in a pICAST ALC vector.

15 FIGURE 23. A schematic depicting use of the complementation technology in the method of the invention. FIGURE 23 shows two inactive mutant reporter enzymes that become active when the corresponding fusion partners, GPCR and β -arrestin interact.

20 FIGURE 24. Vector for expression of a GPCR with inserted seronine/threonine amino acid sequences as a fusion with β -gal $\Delta\alpha$. The open reading frame of a known or orphan GPCR is engineered to contain additional seronine/threonine sequences, such as SSS (seronine, seronine, seronine), within the C-terminal tail. The engineered GPCR is cloned in frame with β -gal $\Delta\alpha$ in a pICAST ALC vector. The pICAST ALC vector contains the following features:

MCS, multiple cloning site for cloning the target protein in frame with the β -gal $\Delta\alpha$;
GS Linker, (GGGGS) $_n$; NeoR, neomycin resistance gene; IRES, internal ribosome
entry site; ColE1ori, origin of replication for growth in *E. coli*; 5'MoMuLV LTR
and 3'MoMuLV LTR, viral promotor and polyadenylation signals from the
5 Moloney Murine leukemia virus.

FIGURE 25. Vector for expression of mutant (R170E) β -arrestin2 as a
fusion with β -gal $\Delta\omega$. The open reading frame of β -arrestin2 is engineered to
contain a point mutation that converts arginine 170 to a glutamate. The mutant β -
arrestin2 is cloned in frame with β -gal $\Delta\omega$ in a pICAST OMC vector. The pICAST
10 OMC vector contains the following features: MCS, multiple cloning site for
cloning the target protein in frame with the β -gal $\Delta\alpha$; GS Linker, (GGGGS) $_n$;
Hygro, hygromycin resistance gene; IRES, internal ribosome entry site; ColE1ori,
origin of replication for growth in *E. coli*; 5'MoMuLV LTR and 3'MoMuLV LTR,
viral promotor and polyadenylation signals from the Moloney Murine leukemia
15 virus.

FIGURE 26. Phosphorylation insensitive Mutant R170E β -Arrestin2 $\Delta\omega$
binds to β 2AR $\Delta\alpha$ in Response to Agonist Activation. A parental β 2AR $\Delta\alpha$ C2 cell
line was transduced with the Mutant R170E β -Arrestin2 $\Delta\omega$ construct. Clonal
populations co-expressing the two constructions were plated at 10,000 cells/well in
20 96 well plates and treated with 10 μ M (-)isoproterenol, 0.3mM ascorbic acid for the
indicated time period. β -galactosidase activity was measured by addition of Tropix
Gal-ScreenTM assay system substrate (Applied Biosystems) and luminescence was
measured using a Tropix TR717TM luminometer (Applied Biosystems). Treatments

were performed in triplicate. For comparison, a clonal cell line (43-8) co-expressing $\beta 2AR\Delta\alpha$ and wild-type β -Arrestin2 $\Delta\omega$ was also plated at 10,000 cells/well and given the same agonist treatment regimen. Minutes of (-)isoproterenol treatment is shown on the X-axis and β -galactosidase activity indicated by relative light units (RLU) is shown on the Y-axis.

FIGURE 27. GPCR dimerization measured by β -galactosidase complementation. A schematic depicting the utilization of the invention for monitoring GPCR homo- or hetero- dimerization. One GPCR is fused to one complement enzyme fragment, while the second GPCR is fused to the second complement enzyme fragment. Interaction of the two GPCRs is monitored by complementation of the enzyme fragments to produce an active enzyme complex (i.e., β -galactosidase activity). GPCR homo- or hetero- dimerization can be monitored in the absence or presence of ligand, agonists, inverse agonists or antagonists.

FIGURE 28. Ligand fishing for orphan receptors by β -galactosidase mutant complementation in ICASTTM system. A schematic depicting the utilization of the invention for ligand fishing and agonist/antagonist screening for orphan GPCRs. As an example, a test cell expressing two β -gal fusion proteins, GPCR_{orphan} $\Delta\alpha$ and Arrestin- $\Delta\omega$, is subjected to treatments with samples from natural or synthetic compound libraries, or from tissue extracts, or from conditioned media of cultured cells. An increased β -gal activity after treatment indicates the activation of the orphan receptor by a ligand in the testing sample. The readout of increased β -gal activity reflects the interaction of an activated

GPCR orphan receptor with a β -arrestin. Therefore, a cognate or a surrogate ligand for the testing receptor is identified.

DETAILED DESCRIPTION OF PREFERRED EMBODIMENTS

5 The present invention provides a method to interrogate GPCR function and pathways. The G-protein-coupled superfamily continues to expand rapidly as new receptors are discovered through automated sequencing of cDNA libraries or genomic DNA. It is estimated that several thousand GPCRs may exist in the human genome. Only a portion have been cloned and even fewer have been
10 associated with ligands. The means by which these, or newly discovered orphan receptors, will be associated with their cognate ligands and physiological functions represents a major challenge to biological and biomedical research. The identification of an orphan receptor generally requires an individualized assay and a guess as to its function. The present invention involves the interrogation of
15 GPCR function by monitoring the activation of the receptor using activation dependent protein-protein interactions between the test GPCR or orphan receptor and a β -arrestin. The specific protein-protein interactions are measured using the mutant enzyme complementation technology disclosed herein. This assay system eliminates the prerequisite guessing because it can be performed with and without
20 prior knowledge of other signaling events. It is sensitive, rapid and easily performed and is applicable to nearly all GPCRs because the majority of these receptors desensitize by a common mechanism.

The present invention provides a complete assay system for monitoring

protein-protein interactions in GPCR pathways. The invention employs the complementation technology, ICAST™ (Intercistronic Complementation Analysis Screening Technology as disclosed in pending U.S. patent application serial no. 053,614, filed April 1, 1998, the entire contents of which are incorporated herein
5 by reference). The ICAST™ technology involves the use of two mutant forms of a reporter enzyme fused to proteins of interest. When the proteins of interest do not interact, the reporter enzyme remains inactive. When the proteins of interest do interact, the reporter enzyme mutants come together and form an active enzyme. According to an embodiment of the invention, the activity of β -galactosidase may
10 be detected with the Gal-Screen™ assay system developed by Advanced Discovery Sciences™, which involves the use of Galacton-Star®, an ultrasensitive chemiluminescent substrate. The Gal-Screen™ assay system and the Galacton-Star® chemiluminescent substrate are disclosed in U.S. Patent Nos. 5,851,771; 5,538,847; 5,326,882; 5,145,772; 4,978,614; and 4,931,569, the contents of which
15 are incorporated herein by reference in their entirety. The invention provides an array of assays, including GPCR binding assays, that can be achieved directly within the cellular environment in a rapid, non-radioactive assay format. The methods of the invention are an advancement over the invention disclosed in U.S. Patent Nos. 5,891,646 and 6,110,693 and the method disclosed in Angers et al.,
20 supra., which rely on microscopic imaging or spectrometry of GPCR components as fusion with Green-fluorescent-protein. The imaging technique disclosed in U.S. Patent Nos. 5,891,646 and 6,110,693 and spectrometry-based technique in Angers et al. are limited by low-throughput and lack of thorough quantification.

The assay system of the invention combined with Advanced Discovery Sciences™ technologies provide highly sensitive cell-based methods for interrogating GPCR pathways which are amenable to high-throughput screening (HTS). Among some of the technologies developed by Advanced Discovery

5 Sciences™ that may be used with the present invention are the Gal-Screen™ assay system (discussed above) and the cAMP-Screen™ immunoassay system. The cAMP-Screen™ immunoassay system provides ultrasensitive determination of cAMP levels in cell lysates. The cAMP-Screen™ assay utilizes the high-sensitivity chemiluminescent alkaline phosphatase (AP) substrate CSPD® (disodium 3-(4-
10 methoxy Spiro {1,2-dioxetane-3,2'-(5'-chloro) tricyclo 3.3.1.1.^{3,7}} decan-4-yl phenyl phosphate) with Sapphire-II™ luminescence enhancer.

Unlike yeast-based-two-hybrid assays used to monitor protein/protein interactions in high-throughput assays, the present invention (1) is applicable to a variety of cells including mammalian cells, plant cells, protozoa cells such as E.
15 coli and cells of invertebrate origin such as yeast, slime mold (*Dictyostelium*) and insects; (2) detects interactions at the membrane at the site of the receptor target or in the cytosol at the site of downstream target proteins rather than a limited cellular localization, i.e., nucleus; and (3) does not rely on indirect read-outs such as transcriptional activation. The present invention thus provides assays with greater
20 physiological relevance and fewer false positives.

The present inventors have developed modifications to the embodiment disclosed in U.S. patent application serial no. 053,614 described above in order to enhance the sensitivity of the inventive GPCR assay. According to an

embodiment, the invention incorporates the use of serine/threonine clusters to enhance and prolong the interaction of GPCR with arrestin in order to make the detection more robust. The clusters can be utilized for orphan receptors or known GPCRs, which do not have this sequence motif. By adding this sequence to the C-terminal tail of the receptor, the activation of the receptor can be detected more readily by readouts of arrestin binding to GPCR, i.e., β -galactosidase complementation from fusion proteins of target proteins with β -galactosidase mutants.

According to another embodiment, the invention incorporates the use of arrestin point mutations to bypass the requirement of phosphorylation, by the action of specific GRK, on the C-terminal tail or intracellular loops of GPCR upon activation. The applications include i) wherein the cognate GRK for a particular GPCR or orphan receptor is unknown; and ii) wherein the specific GRK for the receptor of interest (or under test) may not be present or may have low activity in the host cell that is used for receptor activation assay.

According to another embodiment, the invention incorporates the use of a super arrestin to increase the binding efficiency of arrestin to an activated GPCR and to stabilize the GPCR/arrestin complex during GPCR desensitization. This application can be used to increase the robustness of ICAST/GPCR applications in cases where the GPCR is normally resensitized rapidly post desensitization.

Each of these methodologies is discussed below.

The invention will now be described in the following non-limiting examples.

EXAMPLE:

According to an embodiment of the invention, GPCR activation is measured through monitoring the binding of arrestin to ligand-activated GPCR. In this assay system, a GPCR, e.g., β -adrenergic receptor (β 2AR), and an arrestin, e.g., β -arrestin, are co-expressed in the same cell as fusion proteins with mutant forms of a reporter enzyme, e.g., β -galactosidase (β -gal). As illustrated in Figure 23, the β 2AR is expressed as a fusion protein with $\Delta\alpha$ form of β -gal mutant (β 2AR $\Delta\alpha$) and the β -arrestin as a fusion protein with the $\Delta\omega$ form of β -gal mutant (β -Arr $\Delta\omega$). The two fusion proteins, which at first exist in a resting (or unstimulated) cell in separate compartments, i.e., the membrane for GPCR and the cytosol for arrestin, cannot form an active β -galactosidase enzyme. When such a cell is treated with an agonist or a ligand, the ligand-occupied and activated receptor becomes a high affinity binding site for arrestin. The interaction between an activated GPCR, β 2AR $\Delta\alpha$, and arrestin, β -Arr $\Delta\omega$, drives the β -gal mutant complementation. The enzyme activity can be measured by using an enzyme substrate, which upon cleavage releases a product measurable by colorimetry, fluorescence, or chemiluminescence (e.g., the Gal-Screen™ assay system).

Experiment protocol-

1. In the first step, the expression vectors for β 2AR $\Delta\alpha$ and β Arr2 $\Delta\omega$ were engineered in selectable retroviral vectors pICAST ALC, as described in Figure 18 and pICAST OMC, as described in Figure 15.

2. In the second step, the two expression constructs were transduced into either C2C12 myoblast cells, or other mammalian cell lines, such as COS-7, CHO, A431, HEK 293, and CHW. Following selection with antibiotic drugs, stable clones expressing both fusion proteins at appropriate levels were selected.

5 3. In the last step, the cells expressing both $\beta 2AR\Delta\alpha$ and $\beta Arr2\Delta\omega$ were tested for response by agonist/ligand stimulated β -galactosidase activity. Triplicate samples of cells were plated at 10,000 cells in 100 microliter volume into a well of 96-well culture plate. Cells were cultured for 24 hours before assay. For agonist assay (Figures 3 and 4), cells were treated with variable concentrations of agonist, 10 for example, (-) isoproterenol, procaterol, dobutamine, terbutaline or L-L-phenylephrine for 60 min at 37° C. The induced β -galactosidase activity was measured by addition of Tropix Gal-Screen™ assay system substrate (Applied Biosystems) and luminescence measured in a Tropix TR717™ luminometer (Applied Biosystems). For antagonist assay (Figure 5), cells were pre-incubated for 15 10 min in fresh medium without serum in the presence of ICI-118,551 or propranolol followed by addition of 10 micro molar (-) isoproterenol.

Serine/Threonine Cluster Strategy

Background

20 Based on structure-function relationship studies on β -arrestins, a large region within the amino-terminal half of β -arrestins (termed the activation-recognition domain) recognizes the agonist-activated state of GPCRs. This region of β -arrestin also contains a small positively charged domain (approximately 20

amino acids with net charge +7) called the phosphorylation-recognition domain, which appears to interact with the GRK-phosphorylated carboxyl termini of GPCRs.

GPCRs can be divided into two classes based on their affinities for β -arrestins. Oakley et al., "Association of β -Arrestin with G Protein-Coupled Receptors During Clathrin-Mediated Endocytosis Dictates the Profile of Receptor Resensitization." J. Biol. Chem., 274(45):32248-32257 (1999). The molecular determinants underlying this classification appear to reside in specific serine or threonine residues located in the carboxyl-terminal tail of the receptor. The receptor class that contains serine/threonine clusters (defined as serine or threonine residues occupying three consecutive or three out of four positions) in the carboxyl-termini binds β -arrestin with high affinity upon activation and phosphorylation and remains bound with β -arrestin even after receptor internalization, whereas the receptor class that contains only scattered serine and threonine residues in the carboxy-terminal tail binds β -arrestins with less affinity and disassociates from the β -arrestin upon internalization. Several known GPCRs, such as vasopressin V2 receptor (Oakley, et al.), neurotensin receptor 1 and angiotensin II receptor type 1A (Zhang, et al., "Cellular Trafficking of G Protein-Coupled Receptor/ β -Arrestin Endocytic Complexes." J. Biol. Chem., 274(16):10999-11006 (1999)), which possess one or more of such serine/threonine clusters in their carboxyl-termini, were shown to bind β -arrestins with high affinity.

EXAMPLE

According to an embodiment of the invention, a serine/threonine cluster strategy is used to facilitate screening assays for orphan receptors that do not possess this structural motif of their own. The orphan receptors are easily classified by sequence alignment. Orphan receptors lacking the serine/threonine clusters are each cloned into an expression vector that is modified to introduce one or more serine/threonine cluster(s) to the carboxyl-terminal tail of the receptor (FIGURE 24). The serine/threonine clusters enhance the receptor activation dependent interaction between the activated and phosphorylated receptor (negative charges) and β -arrestin (positive charges in the phosphorylation-recognition domain) through strong ionic interactions, thus prolonging interaction between the receptor and arrestin. The modification of the orphan receptor tail thus makes detection of receptor activation more robust.

15 Experiment protocol -

1. In a first step, the open-reading-frame (ORF) of an orphan receptor, which lacks the serine/threonine clusters, is cloned into a modified expression vector such as pICAST ALC described in Figure 10A. The modified pICAST ALC includes coding sequences for one or more sets of serine/threonine clusters (for example, SSS or SST) located downstream from the insert of the ORF of an orphan receptor (FIGURE 24).

2. In a second step, chimeric orphan receptor, $\text{ORF}_{\text{orphan R}}\text{-(SSS)}_n\text{-}\Delta\alpha$, is co-

expressed in a mammalian cell with a β -arrestin chimera, such as β Arr2 $\Delta\omega$ described in Figure 15.

3. In a third step, the cell is treated with an agonist or a ligand and the activated receptor with phosphorylated serine cluster(s) binds the β -arrestin with high affinity producing strong signals in readouts of β -gal complementation.

This assay, which provides a means for sensitive measurement of functional activation of the orphan receptors, can be used to screen for natural or surrogate ligands for orphan receptors, a process called de-orphaning or target discovery for new GPCRs (FIGURE 28). Furthermore, this assay is also useful in screening for potential agonists and antagonists for lead discovery of GPCRs.

Enhanced Binding of Arrestin in the Presence and in the Absence of GPCR

Phosphorylation

Background

Six different classes of G-protein coupled receptor kinases (GRKs) have been identified and each of these has been reported to be expressed as multiple splice variants. Krupnick et al., "The role of receptor kinases and arrestins in G protein-coupled receptor regulation." Ann. Rev. Pharmacol. Toxicol., 38:289-319 (1998). Although many cell lines express a variety of GRKs, the specific GRK required for phosphorylation of a given GPCR may not always be present in the cell line used for recombinant GPCR and arrestin expression. This is particularly an issue for applications using orphan receptors, in which case the cognate GRK will likely be unknown. In other cases, the cell line used for recombinant

expression work may have the required GRK, but may express the GRK at low levels. In order to bypass such caveats, genetically modified arrestins that bind specifically to activated GPCRs, but without the requirement of GRK phosphorylation are employed.

5 Mutagenesis studies on arrestins demonstrate that point mutations in the phosphorylation-recognition domain, particularly mutations converting Arg175 (of visual arrestin) to an oppositely charged residue such as glutamate (R175E mutation), result in an arrestin which specifically binds to activated GPCRs, but does so without the requirement for phosphorylation.

10 Numerous observations have led to the hypothesis that arrestin exists in an inactive state that has a low affinity for GPCRs. Once a GPCR is both activated and phosphorylated, the phosphorylated region of the GPCR C-terminus interacts with the phosphorylation-recognition domain of arrestin causing the arrestin to change conformations allowing the activation-recognition region to be exposed for
15 binding to the activated/ phosphorylated receptor. Vishnivetskiy et al., "How does arrestin respond to the phosphorylated state of rhodopsin?" J. Biol. Chem., 274(17):11451-11454 (1999); Gurevich et al., "Arrestin interactions with G protein-coupled receptors. Direct binding studies of wild-type and mutant arrestins with rhodopsin, beta 2-adrenergic and m2 muscarinic cholinergic receptors." J.
20 Biol. Chem., 270(2):720-731, (1995); Gurevich et al., "Mechanism of phosphorylation-recognition by visual arrestin and the transition of arrestin into a high affinity binding site." Mol. Pharmacol., 51(1):161-169 (1997); Kovoor et al., "Targeted construction of phosphorylation-independent beta-arrestin mutants with

constitutive activity in cells." J. Biol. Chem., 274(11):6831-6834 (1999). In summary, binding studies of single mutation, double mutation, deletion, and chimerical arrestins with inactive, inactive and phosphorylated, activated but not phosphorylated, or activated and phosphorylated visual or non-visual GPCRs all support this model.

EXAMPLE

A phosphorylation insensitive mutant of arrestin fused to mutant reporter protein can be produced that will bind to activated GPCRs in a phosphorylation independent manner. As proof of concept, a point mutation for β -arrestin2, R170E β -arrestin2, has been produced and its interaction with β 2AR has been analyzed in accordance with the invention.

Experimental protocol:

- 1) In the first step, β -arrestin2 was mutated such that Arg170 was converted to Glu. This mutation is equivalent to the R175E mutation of visual arrestin. The mutant β -arrestin2 open reading frame was cloned in frame with $\Delta\omega$ - β -galactosidase in the pICAST OMC expression vector to produce a modified expression vector R170E β -arrestin2 (FIGURE 25).
- 2) In the second step, the R170E β -arrestin2 expression construct was transduced into a C2C12 myoblast cell line that had been engineered to express β 2AR as a fusion to $\Delta\alpha$ - β -galactosidase as described in Figure 18 of U.S. Application Serial No. 09/654,499. Following selection with antibiotic drugs, a

population of clones expressing both fusion proteins was obtained.

3) In the last step, this population of cells expressing both R170E β -arrestin2 $\Delta\omega$ and β 2AR $\Delta\alpha$ were tested for response by agonist/ligand stimulated β -galactosidase activity as demonstrated in FIGURE 26. The C2C12 clone 43-8 co-expressing β 2AR $\Delta\alpha$ and wild-type β -arrestin2 $\Delta\omega$ (FIGURE 26) was used as reference control. Triplicate samples of cells were plated at 10,000 cells in 100 microliter volume into wells of a 96-well culture plate. Cells were cultured for 24 hours before assay. For agonist assay as in FIGURE 26, cells were treated with 10 μ m (-)-isoproterenol stabilized with 0.3mM ascorbic acid 37° C for 0, 5, 10, 15, 30, 45 or 60 minutes. The induced β -galactosidase activity was measured by addition of Tropix Gal-Screen™ assay system substrate (Applied Biosystems) and luminescence measured in a Tropix TR717™ luminometer (Applied Biosystems). As shown in Figure 26, the mutant arrestin interacts with β 2AR in an agonist-dependent manner and was comparable with that of wild-type arrestin.

4) To expand the application of phosphorylation-insensitive arrestin, cell lines such as C2C12, CHO or HEK 293, are developed that express the R170E β -arrestin2 $\Delta\omega$ construction. These cell lines can be used to transduce orphan or known GPCRs as fusions with $\Delta\alpha$ - β -galactosidase in order to develop cell lines for agonist and antagonist screening and

Development of Super Arrestins:

Background

Attenuation of GPCR signaling by the arrestin pathway serves to ensure that a cell or organism does not over-react to a stimulus. At the same time, the arrestin pathway often serves to recycle the GPCR such that it can be temporarily inactivated but then quickly resensitized to allow for sensitivity to new stimuli. The down-regulation process involves phosphorylation of the receptor, binding to arrestin and endocytosis. Following endocytosis of the desensitized receptor, the receptor is either degraded in lysosomes or resensitized and sent back to the membrane. Resensitization involves release of arrestin from the receptor, dephosphorylation and cycling back to the membrane. The actual route a GPCR follows upon activation depends on its biological function and the needs of the organism. Because of these diverse pathways that may be required of the down-regulation pathway, arrestin affinities for activated GPCRs vary from receptor to receptor. It would thus be very advantageous to engineer super arrestins that have a higher affinity and avidity for activated GPCRs than what nature has provided.

Although mutational, deletion and chimerical studies of arrestins have focused on understanding regulatory switches in the molecule that respond to GPCR phosphorylation states, several of these altered recombinant forms of arrestin have resulted in molecules with enhanced binding to activated, phosphorylated GPCRs. Conversion of Arg175 to histidine, tyrosine, phenylalanine or threonine results in significantly higher amounts of binding to phosphorylated, activated rhodopsin than wild-type arrestin or R175E arrestin,

although these mutations result in less binding to activated, non-phosphorylated receptor. Gurevich et al. (1997). In addition, conversion of Valine 170 to alanine increased the constitutive affect of the R175E mutation, but also nearly doubled the amount of interaction of wild-type arrestin with activated, phosphorylated rhodopsin. Gurevich et al. (1997).

Truncation of β -arrestin1 at amino acid 382 has been reported to enhance binding of both R169E (equivalent to arrestin R175E) and wild-type β -arrestin1 to activated or activated and phosphorylated receptor, respectively. Kovoor et al. Chimerical arrestins in which functional regions of visual arrestin were swapped with those of β -arrestin1 have been reported to be altered in binding affinity to activated, phosphorylated GPCRs. Gurevich et al. (1995). Several of these chimeras, such as β -arrestin1 containing the visual arrestin extreme N-terminus, show increased specific binding to phosphorylated activated GPCRs compared to wild-type β -arrestin1 (Gurevich et al. (1995)). Modifications that enhance arrestin affinity for the activated GPCR such as described above, whether phosphorylated or non-phosphorylated, could also enhance signal to noise of β -galactosidase activity since the arrestin/GPCR complex is stabilized and/or more long-lived. The use of mutant arrestins with higher activated-GPCR affinity would improve the inventive technology for GPCR targets, without compromising receptor/ligand biology.

In addition, this "super arrestin" approach can be combined with the use of arrestin point mutations to provide a stronger signal to noise with or without GRK requirements.

EXAMPLE

An arrestin mutant fused to mutant reporter protein can be produced to enhance binding of the arrestin to an activated GPCR to enhance sensitivity of detection.

5 Experiment protocol -

- 1) In the first step, mutant β -arrestin2 constructions will be generated which include R170E/T/Y/or H, V165A, substitution of a.a. 1-43 with a.a. 1-47 of visual arrestin, or deletion of the C-terminal and combinations of these alterations. The mutant β -arrestin2 open reading frames will be cloned in frame with $\Delta\omega$ - β -galactosidase in the pICAST OMC expression vector similar to cloning of the R170E β -arrestin2 mutation shown in FIGURE 25.
- 10 2) In the second step, mutant expression constructs will be transduced into a C2C12 myoblast cell line that has been engineered to express β 2AR as a fusion to $\Delta\alpha$ - β -galactosidase. Following selection with antibiotic drugs, a population of clones expressing both fusion proteins will be obtained. Wild type and R170E β -arrestin2 constructions will be transduced to generate control, reference clonal populations.
- 15 3) In the third step, populations of cells expressing both β -arrestin2 $\Delta\omega$ (mutant or wild type) and β 2AR $\Delta\alpha$ will be tested for response by agonist/ligand stimulated β -galactosidase activity.
- 20 4) In the next step, mutant (super) β -arrestin2 $\Delta\omega$ constructions that show a significantly higher signal to noise ratio in the agonist assay compared with wild-type β -arrestin2 $\Delta\omega$ will be chosen. These constructions will be used to develop

stable cell lines expressing the "super" β -arrestin2 $\Delta\omega$ that can be used for transducing in known or orphan GPCRs. Use of a super β -arrestin2 $\Delta\omega$ could increase the signal to noise of ICAST/GPCR applications allowing improved screening capabilities for lead and ligand discovery.

5 Super Arrestin is used to increase the binding efficiency of arrestin to an activated GPCR and to stabilize the GPCR/arrestin complex during GPCR desensitization. This application can be used to increase the robustness of ICAST/GPCR applications in cases where the GPCR is normally resensitized rapidly post desensitization.

10 The assays of this invention, and their application and preparation have been described both generically, and by specific example. The examples are not intended as limiting. Other substituent identities, characteristics and assays will occur to those of ordinary skill in the art, without the exercise of inventive faculty. Such modifications remain within the scope of the invention, unless excluded by
15 the express recitation of the claims advanced below.

WHAT IS CLAIMED IS:

1. A method of assessing the effect of a test condition on G-protein-coupled receptor (GPCR) pathway activity, comprising:

5 a) providing a cell that expresses a GPCR as a fusion protein to one mutant form of reporter enzyme and an interacting protein partner as a fusion to another mutant form of enzyme,

wherein said cell also expresses an arrestin, wherein said arrestin is modified to enhance binding of said arrestin to said GPCR, wherein said enhanced binding between said arrestin and said GPCR increases sensitivity of detection of
10 said effect of said test condition;

b) exposing the cell to a ligand for said GPCR under said test condition; and

c) monitoring activation of said GPCR by complementation of said reporter enzyme;

wherein increased reporter enzyme activity in the cell compared to that
15 which occurs in the absence of said test condition indicates increased GPCR interaction with its interacting protein partner compared to that which occurs in the absence of said test condition, and decreased reporter enzyme activity in the cell compared to that which occurs in the absence of said test condition indicates decreased GPCR interaction with its interacting protein partner compared to that
20 which occurs in the absence of said test condition.

2. A method of assessing the effect of a test condition on G-protein-coupled receptor (GPCR) pathway activity, comprising:

a) providing a cell that expresses a GPCR as a fusion protein to one mutant

form of reporter enzyme and an interacting protein partner as a fusion to another mutant form of enzyme;

wherein said GPCR fusion protein is modified to include one or more sets of serine/threonine clusters, wherein said one or more sets of serine/threonine clusters enhance binding of said GPCR to arrestin, wherein said enhanced binding between said GPCR and said arrestin increases sensitivity of detection of said effect of said test condition;

b) exposing the cell to a ligand for said GPCR under said test condition; and

c) monitoring activation of said GPCR by complementation of said reporter enzyme;

wherein increased reporter enzyme activity in the cell compared to that which occurs in the absence of said test condition indicates increased GPCR interaction with said interacting protein partner compared to that which occurs in the absence of said test condition, and decreased reporter enzyme activity in the cell compared to that which occurs in the absence of said test condition indicates decreased GPCR interaction with interacting protein partner compared to that which occurs in the absence of said test condition.

3. A DNA molecule comprising a sequence encoding a biologically active hybrid GPCR, wherein said hybrid GPCR comprises a GPCR as a fusion protein to one mutant form of reporter enzyme and wherein said hybrid GPCR is modified to include one or more sets of serine/threonine clusters, wherein said one or more sets of serine/threonine clusters enhance binding of said hybrid GPCR to arrestin.

4. A DNA construct capable of directing the expression of a biologically

active hybrid GPCR in a cell, comprising the following operatively linked elements:

a promoter; and

a DNA molecule comprising a sequence encoding a biologically active hybrid GPCR, wherein said hybrid GPCR comprises a GPCR as a fusion protein to one mutant form of reporter enzyme and wherein said hybrid GPCR is modified to include one or more sets of serine/threonine clusters, wherein said one or more sets of serine/threonine clusters enhance binding of said hybrid GPCR to arrestin.

5. A cell transformed with a DNA construct capable of expressing a biologically active hybrid GPCR in a cell, comprising the following operatively linked elements:

a promoter; and

a DNA molecule comprising a sequence encoding a biologically active hybrid GPCR, wherein said hybrid GPCR comprises a GPCR as a fusion protein to one mutant form of reporter enzyme and wherein said hybrid GPCR is modified to include one or more sets of serine/threonine clusters, wherein said one or more sets of serine/threonine clusters enhance binding of said hybrid GPCR to arrestin.

6. A DNA molecule comprising a sequence encoding a biologically active hybrid arrestin, wherein said hybrid arrestin comprises an arrestin as a fusion to one mutant form of reporter enzyme and wherein said hybrid arrestin is modified to enhance binding of said arrestin to GPCR.

7. A DNA construct capable of directing the expression of a biologically active hybrid arrestin in a cell, comprising the following operatively linked

elements:

a promoter; and

a DNA molecule comprising a sequence encoding a biologically active hybrid arrestin, wherein said hybrid arrestin comprises an arrestin as a fusion to one mutant form of reporter enzyme and wherein said hybrid arrestin is modified to enhance binding of said arrestin to GPCR.

8. A cell transformed with a DNA construct capable of expressing a biologically active hybrid arrestin in a cell, comprising the following operatively linked elements:

a promoter; and

a DNA molecule comprising a sequence encoding a biologically active hybrid arrestin, wherein said hybrid arrestin comprises an arrestin as a fusion to one mutant form of reporter enzyme and wherein said hybrid arrestin is modified to enhance binding of said arrestin to GPCR.

9. A method of assessing the effect of a test condition on G-protein-coupled receptor (GPCR) pathway activity, comprising:

a) providing a cell that expresses a GPCR as a fusion protein to one mutant form of reporter enzyme and an interacting protein partner as a fusion to another mutant form of enzyme,

wherein said cell also expresses an arrestin, wherein said arrestin is modified by introducing a point mutation in a phosphorylation-recognition domain to remove a requirement for phosphorylation of said GPCR for arrestin binding to permit binding of said arrestin to said GPCR in said cell regardless of whether said

GPCR is phosphorylated,

b) exposing the cell to a ligand for said GPCR under said test condition; and

c) monitoring activation of said GPCR by complementation of said reporter enzyme;

5 wherein increased reporter enzyme activity in the cell compared to that which occurs in the absence of said test condition indicates increased GPCR interaction with its interacting protein partner compared to that which occurs in the absence of said test condition, and decreased reporter enzyme activity in the cell compared to that which occurs in the absence of said test condition indicates
10 decreased GPCR interaction with its interacting protein partner compared to that which occurs in the absence of said test condition.

10. The method of Claim 9, wherein said arrestin is mutated to increase a property selected from affinity and avidity for activated, non-phosphorylated GPCR.

15 11. The method of Claim 10, wherein said arrestin is β -arrestin2 and wherein said β -arrestin2 is mutated to convert Arg169 to an oppositely charged residue.

12. The method of Claim 11, wherein said oppositely charged residue is selected from the group consisting of histidine, tyrosine, phenylalanine and
20 threonine.

13. The method of Claim 9, wherein said arrestin is mutated to increase a property selected from affinity and avidity for activated and phosphorylated GPCR.

14. A method of assessing the effect of a test condition on G-protein-

coupled receptor (GPCR) pathway activity, comprising:

a) providing a cell that expresses a GPCR as a fusion protein to one mutant form of reporter enzyme and an interacting protein partner as a fusion to another mutant form of enzyme;

5 wherein said GPCR fusion protein is modified to include one or more sets of serine/threonine clusters, said one or more serine/threonine clusters defined as serine or threonine residues occupying three consecutive or three out of four positions in a carboxyl-termini of said GPCR, wherein said one or more sets of serine/threonine clusters enhance binding of said GPCR to arrestin, wherein said
10 enhanced binding between said GPCR and said arrestin increases sensitivity of detection of said effect of said test condition;

b) exposing the cell to a ligand for said GPCR under said test condition; and

c) monitoring activation of said GPCR by complementation of said reporter enzyme;

15 wherein increased reporter enzyme activity in the cell compared to that which occurs in the absence of said test condition indicates increased GPCR interaction with said interacting protein partner compared to that which occurs in the absence of said test condition, and decreased reporter enzyme activity in the cell compared to that which occurs in the absence of said test condition indicates
20 decreased GPCR interaction with interacting protein partner compared to that which occurs in the absence of said test condition.

15. The method of Claim 1, wherein said modified arrestin exhibits enhanced binding to activated, phosphorylated GPCR.

25. The method of Claim 14, wherein said modified arrestin comprises conversion of Arg170 to an amino acid selected from the group consisting of histidine, tyrosine, phenylalanine and threonine.

Cellular Expression of β_2 AR- β gal $\Delta\alpha$ Fusion Protein in C2 Clones
(measured by anti- β -gal ELISA)

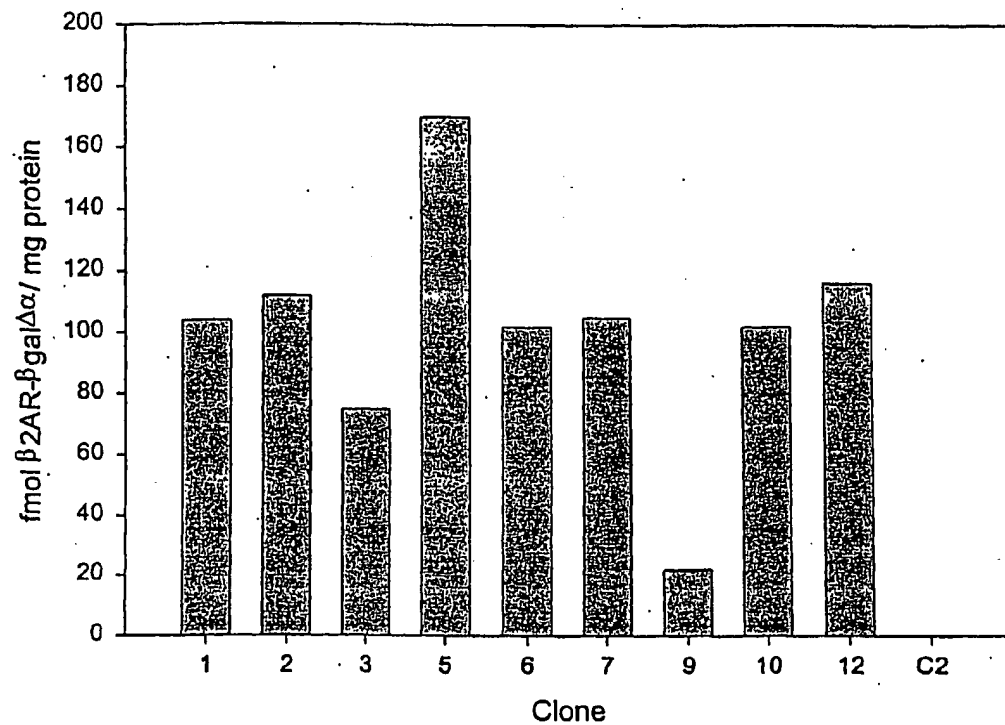


FIGURE 1A

Cellular expression of β Arr2- β gal $\Delta\omega$ fusion protein in C2 clones
(measured by anti- β gal ELISA)

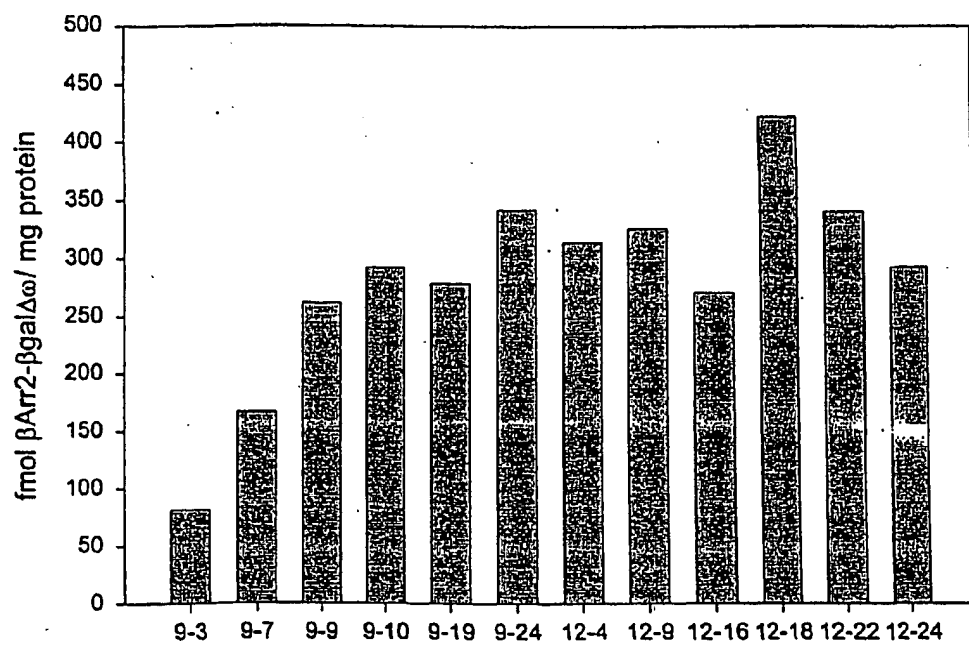


FIGURE 1B

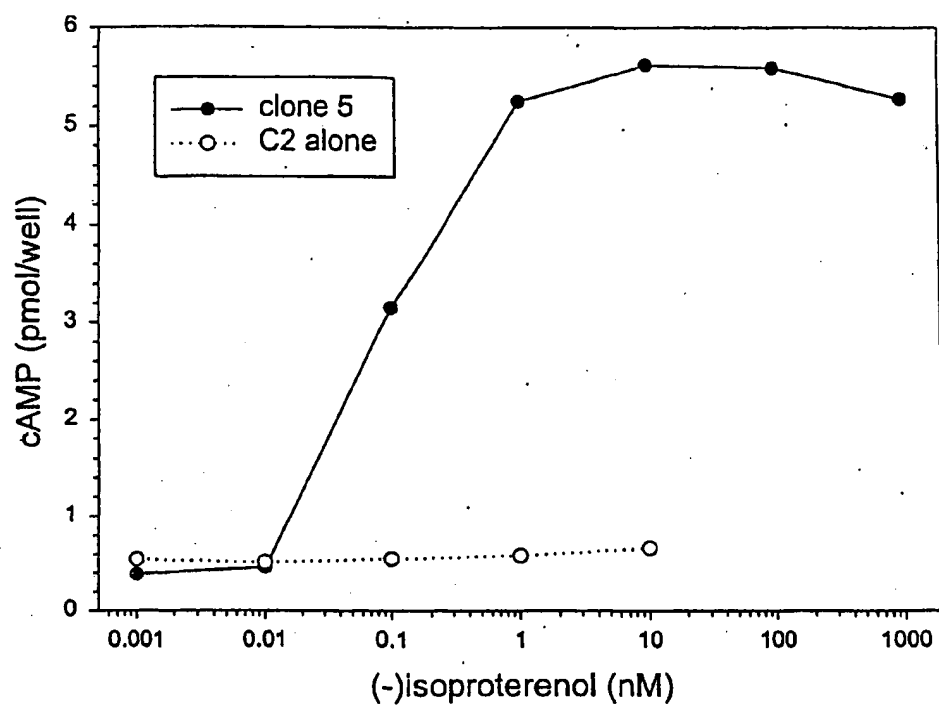
Agonist Stimulated cAMP Response in C2 Cells Expressing $\beta 2AR$ - $\beta gal\Delta\alpha$ 

FIGURE 2

β -galactosidase Complementation as a Measurement for β 2AR- β gal $\Delta\alpha$ interacting with β Arrestin2- β gal $\Delta\omega$ upon agonist Stimulation

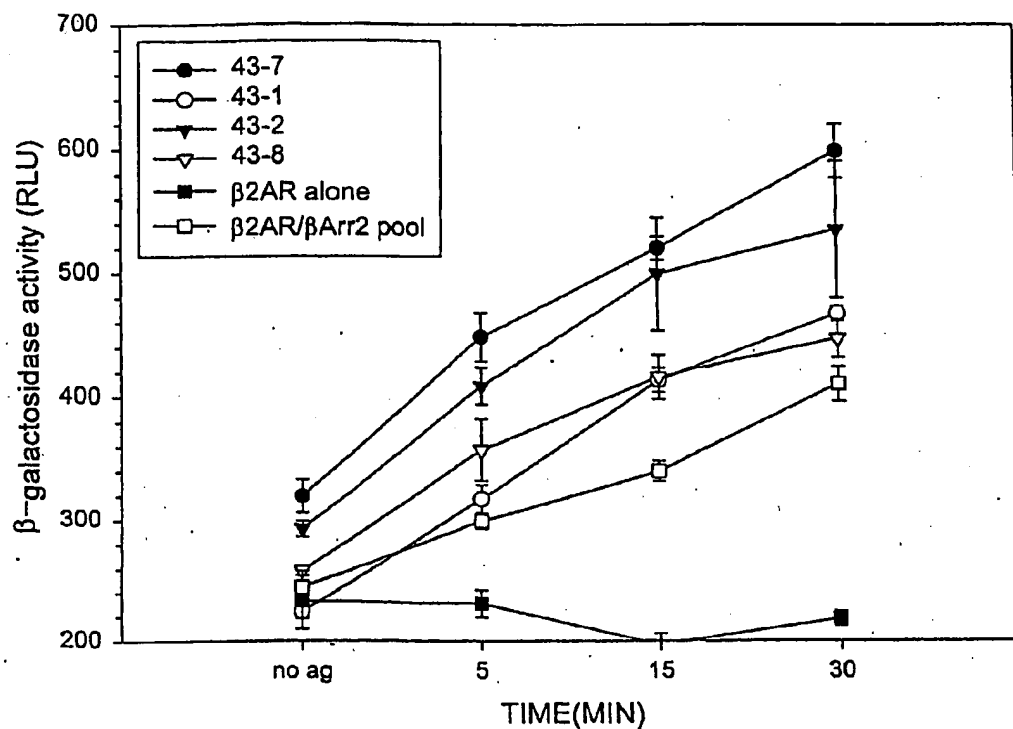


FIGURE 3A

β -galactosidase Complementation as a Measurement for β 2AR- β gal $\Delta\alpha$ Interaction with β Arrestin1- β gal $\Delta\omega$ upon Agonist Stimulation

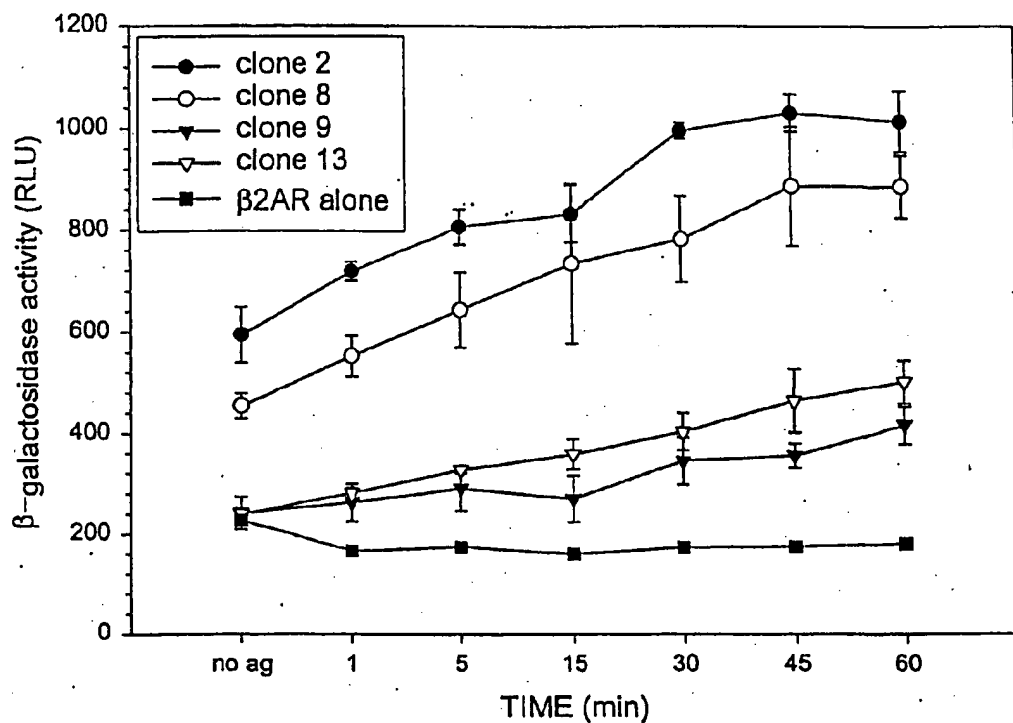


FIGURE 3B

β -galactosidase Activity in Response to Agonist in C2 Cells
Coexpressing β 2AR- β gal $\Delta\alpha$ and β Arrestin2- β gal $\Delta\omega$ Fusion Proteins

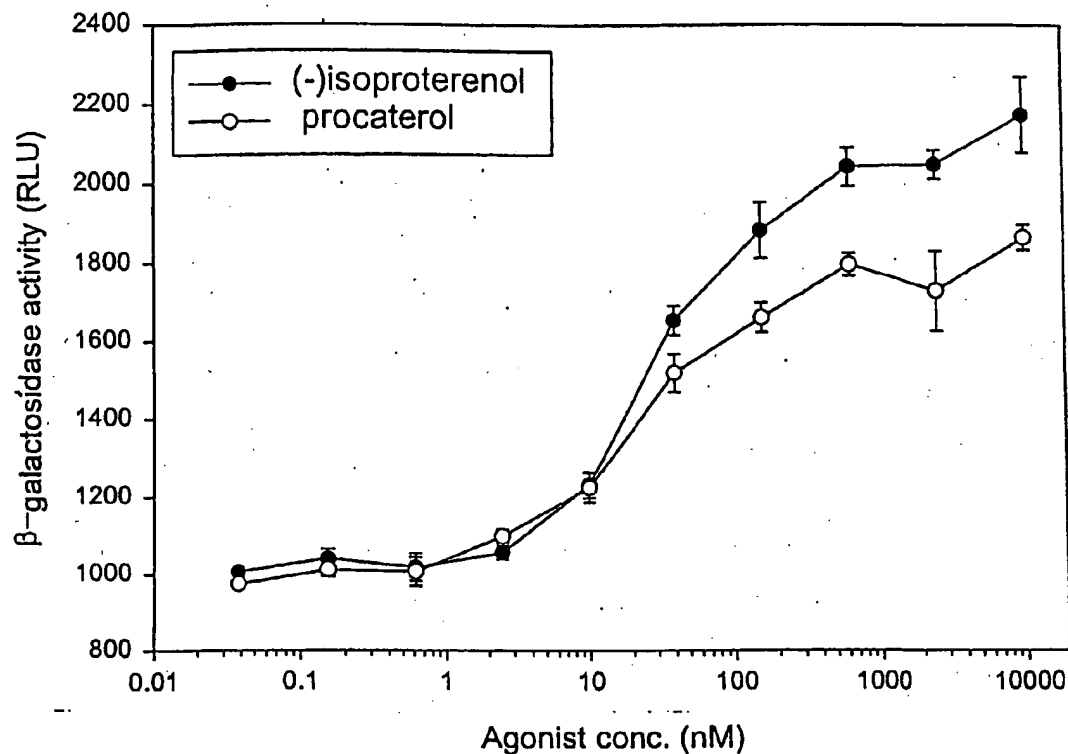


FIGURE 4A

β -galactosidase Activity in Response to Agonist in C2 Cells
Coexpressing β 2AR- β gal $\Delta\alpha$ and β Arrestin1- β gal $\Delta\omega$ Fusion Proteins

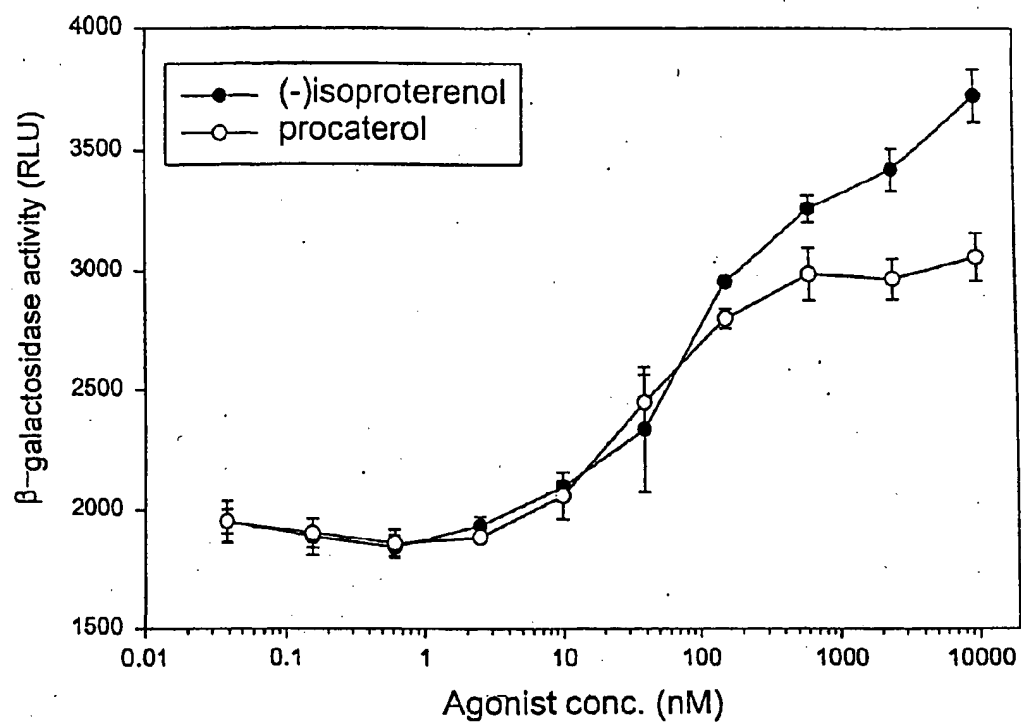


FIGURE 4B

Inhibition of β -galactosidase activity in C2 Cells Coexpressing β 2AR- β gal $\Delta\alpha$ and β Arrestin2- β gal $\Delta\omega$ Fusion Proteins

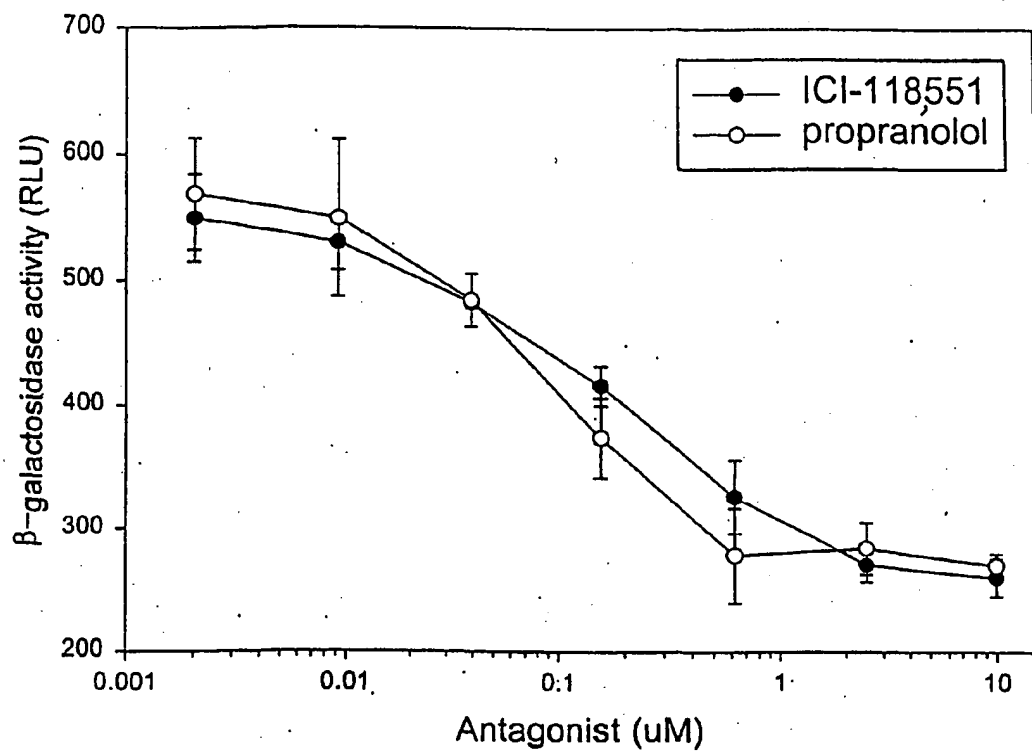


FIGURE 5A

Antagonist Inhibition of β -galactosidase Activity in C2 Cells
Coexpressing β 2AR- β gal $\Delta\alpha$ and β Arrestin1- β gal $\Delta\omega$ Fusion Proteins

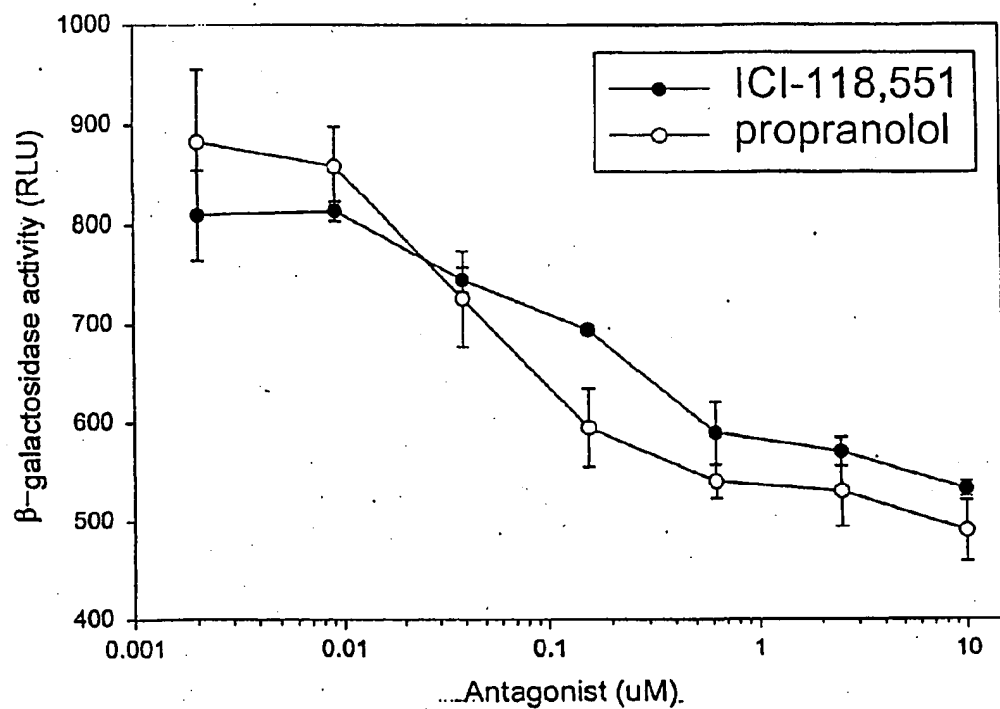


Figure 5B

Agonist Stimulated cAMP Response in Clones or Pools of C2 Cells
Coexpressing A2aR- β gal $\Delta\alpha$ and β Arrestin1- β gal $\Delta\omega$ Fusion Proteins

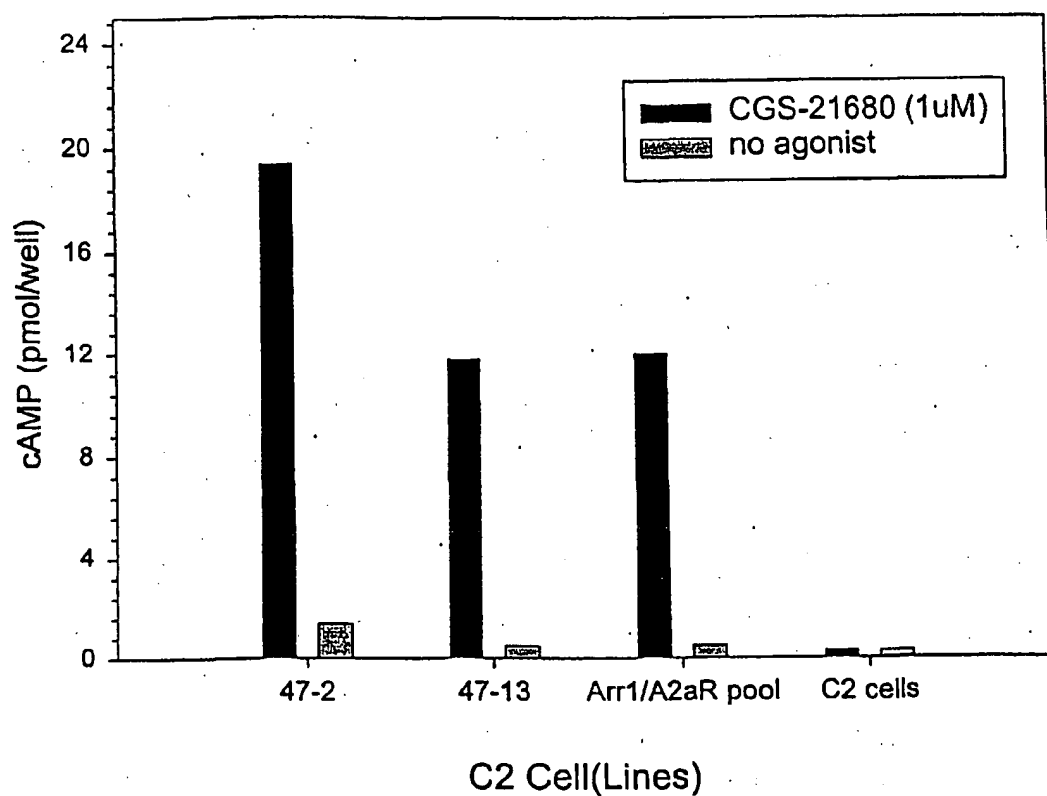


FIGURE 6

Agonist Stimulated cAMP Response in Clones or Pools of C2 Cells
Expressing D1- β gal $\Delta\alpha$ and β Arrestin2- β gal $\Delta\omega$ Fusion Proteins

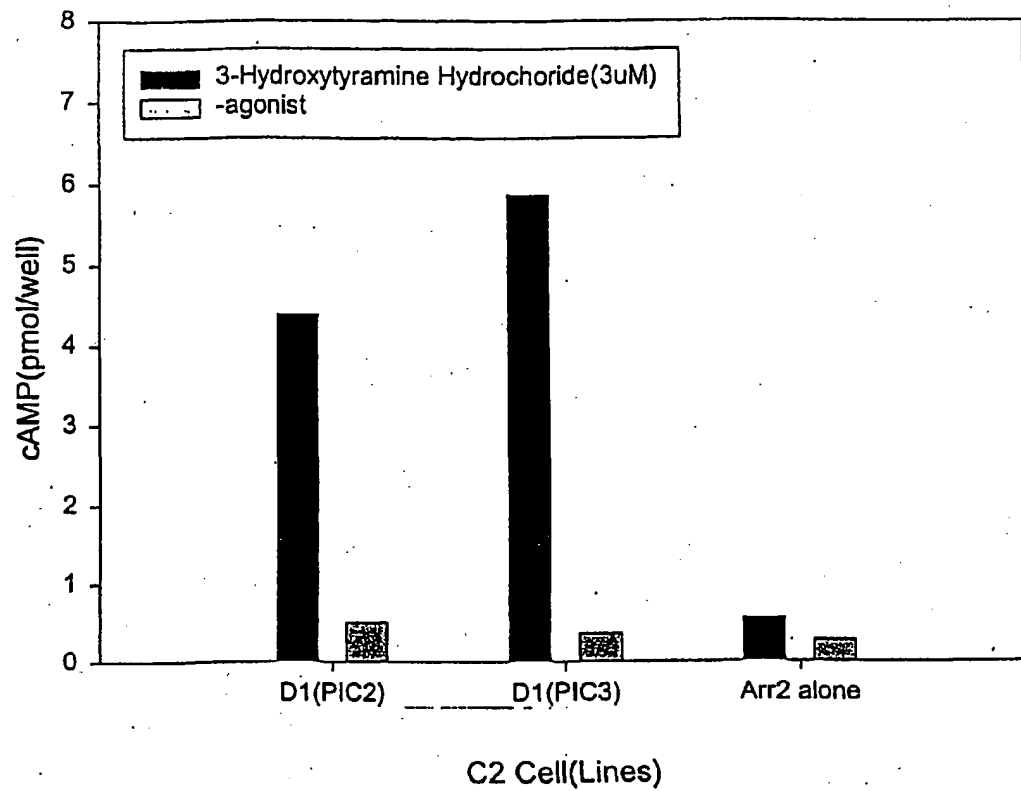


FIGURE 7

β_2 AR- β gal $\Delta\omega$ and β arr2- β gal $\Delta\alpha$ Interaction in HEK293
Clones in Response to Isoproterenol Treatment (1 μ M)

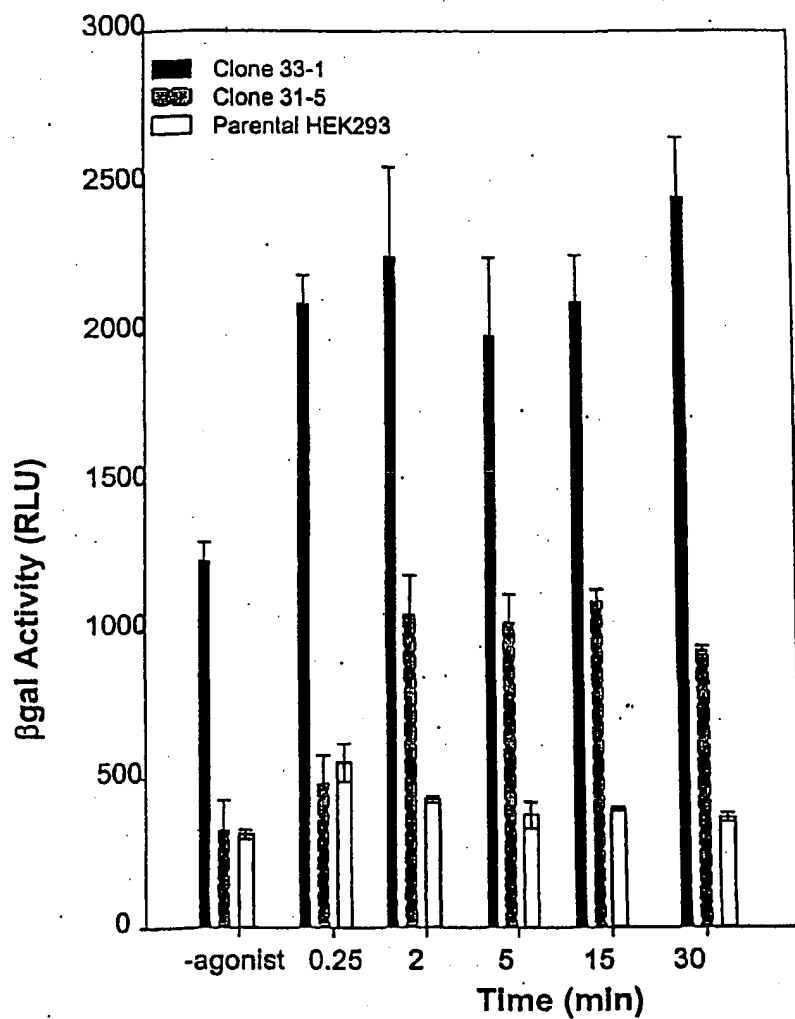


FIGURE 8A

β 2AR- β gal $\Delta\alpha$ and β Arr1- β gal $\Delta\omega$ Interaction in a CHO Pool
in Response to Isoproterenol Treatment(10uM)

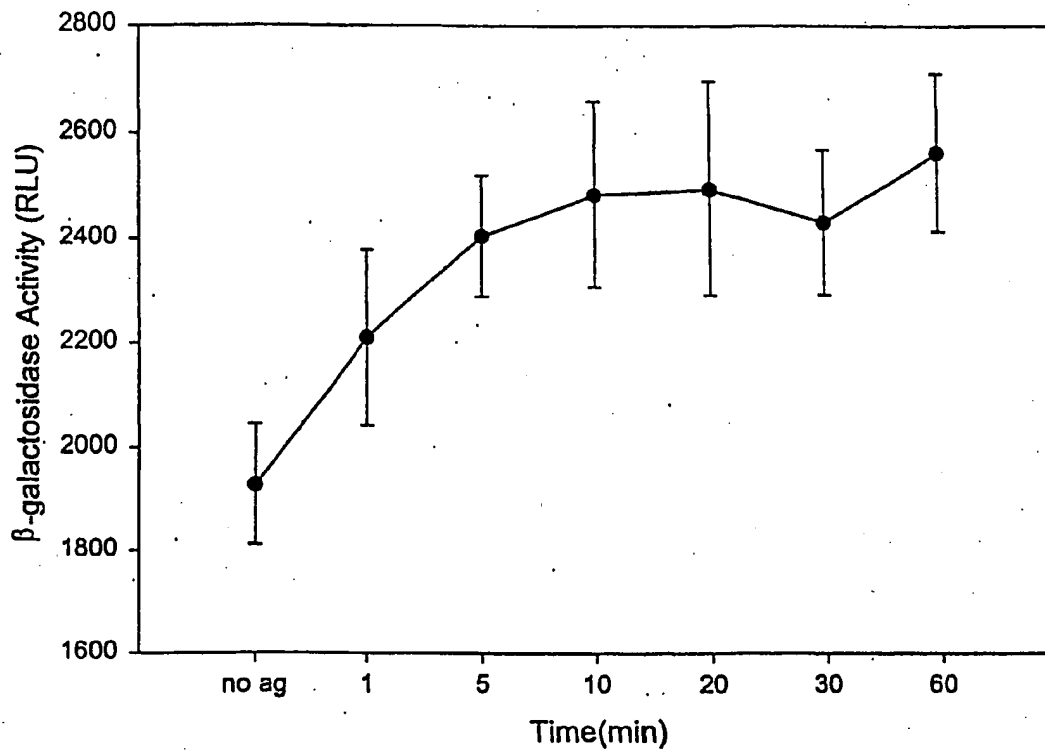


FIGURE 8B

β 2AR- β gal $\Delta\alpha$ and β Arr2- β gal $\Delta\omega$ Interaction in CHW Clone
in Response to Isoproterenol Treatment (10 μ M)

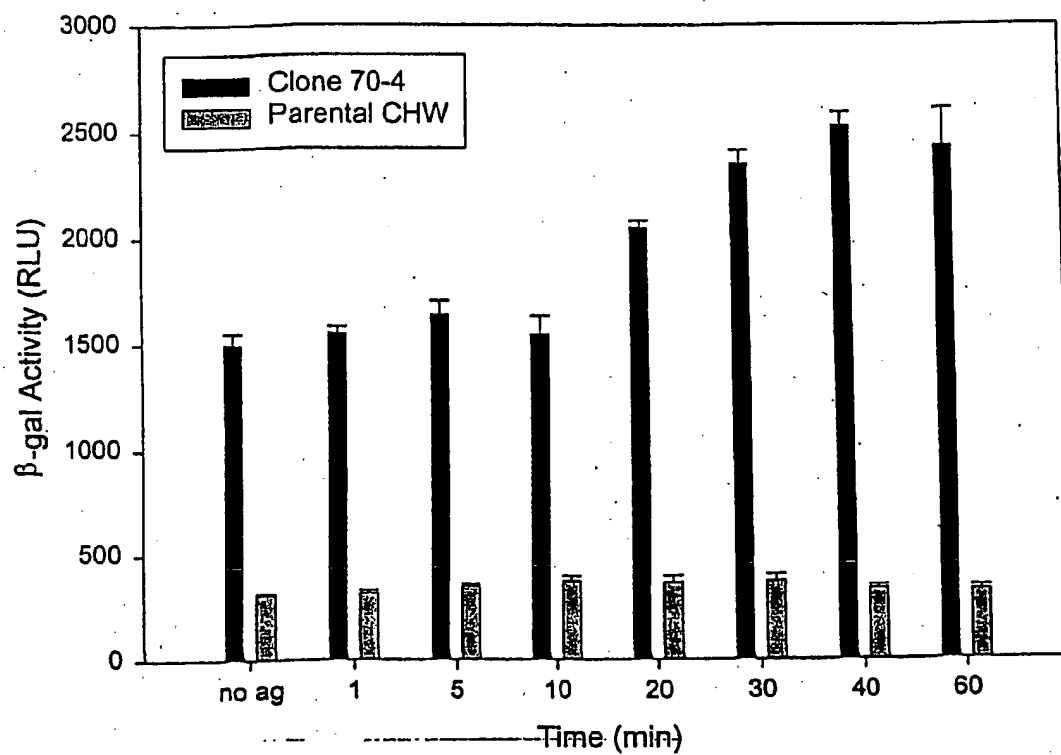


FIGURE 8C

β -galactosidase Complementation as a Measurement for
Adrenergic Receptor Homodimerization in HEK 293 Cells
Coexpressing β 2AR- β gal $\Delta\alpha$ and β 2AR- β gal $\Delta\omega$.

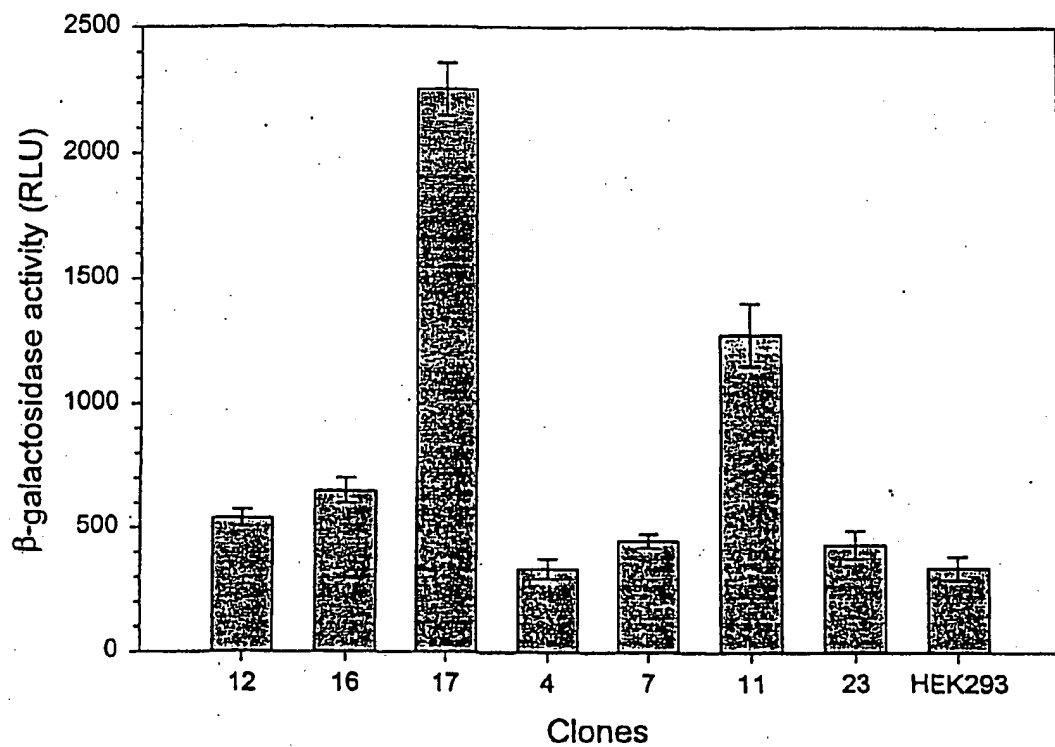


FIGURE 9A

Agonist Stimulated cAMP Response in HEK 293 Cells
Coexpressing $\beta 2AR$ - $\beta gal\Delta\alpha$ and $\beta 2AR$ - $\beta gal\Delta\omega$

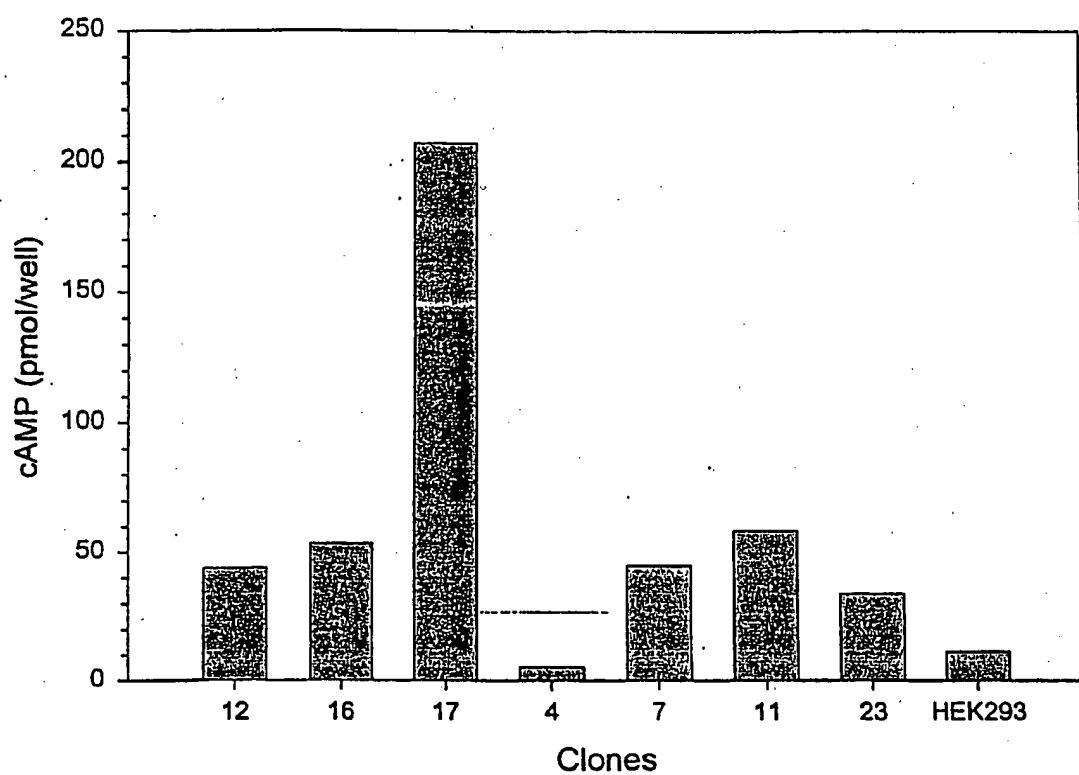


FIGURE 9B

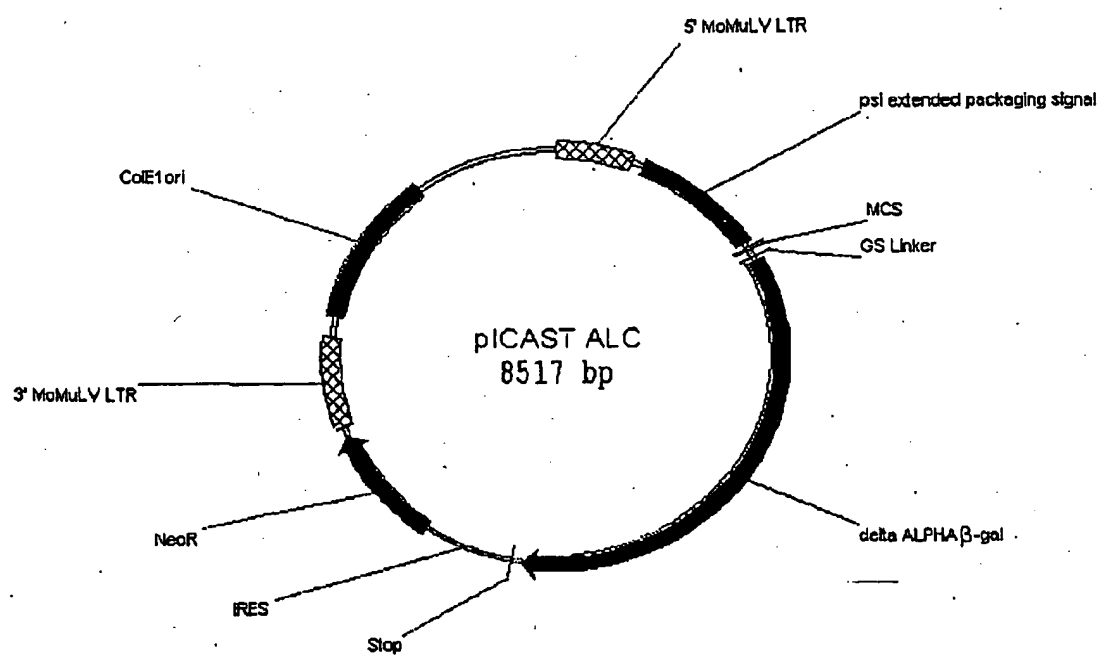


Figure 10A

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1  CTGCAGCCTG AATATGGGCC AAACAGGATA TCTGTGGTAA GCAGTTCCTG
   GACGTCCGGAC TTATACCCGG TTGTCTCTAT AGACACCATT CGTCAAGGAC
-----
51  CCCC GGCTCA GGGCCAAGAA CAGATGGAAC AGCTGAATAT GGGCCAAACA
   GGGGCCGAGT CCCGGTCTT GTCTACCTTG TCGACTTATA CCGGTTTGT
-----
101 GGATATCTGT GGTAAAGCAGT TCCTGCCCCG GCTCAGGGCC AAGAACAGAT
   CCTATAGACA CCATTCTGTCA AGGACGGGGC CGAGTCCCGG TTCTTGCTTA
-----
151 GGTCCCAGAG TGCGGTCCAG CCCTCAGCAG TTTCTAGAGA ACCATCAGAT
   CCAGGGGTCT ACGCCAGGTC GGGAGTCGTC AAAGATCTCT TGGTAGTCTA
-----
201 GTTCCAGGG TGCCCCAAGG ACCTGAAATG ACCCTGTGCC TTATTTGAAC
   CAAAGGTCCC ACGGGGTTC TGGACTTTAC TGGGACACGG AATAAACTTG
-----
251 TAACCAATCA GTTCGCTTCT CGCTTCTGTT CGCGCGCTTC TGCTCCCGA
   ATTGGTTAGT CAAGCGAAGA GCGAAGACAA GCGCGCGAAG ACGAGGGGCT
-----
301 GCTCAATAAA AGAGCCACACA ACCCTCACT CGGGGCGCCA GTCCCTCCGAT
   CGAGTTATTT TCTCGGGTGT TGGGGAGTGA GCCCGCGGT CAGGAGGCTA
-----
351 TGA CTGAGTC GCCCGGGTAC CCGTGTATCC AATAAACCCCT CTG CAGTTG
   ACTGACTCAG CGGGCCCATG GGCACATAGG TTATTTGGGA GAACGTCAAC
-----
401 CATCCGACTT GTGGTCTCGC TGTTCTTGG GAGGGTCTCC TCTGAGTGAT
   GTAGGCTGAA CACCAGAGCG ACAAGGAACC CTCCCAGAGG AGACTCTACTA
-----
451 TGA CTACCCG TCAGCGGGGG TCTTTCATTT GGGGGCTCGT CCGGGATCGG
   ACTGATGGGC AGTCGCCCCC AGAAAGTAAA CCCCCGAGCA GGCCCTAGCC
-----
501 GAGACCCCTG CCCAGGGACC ACCGACCCAC CACCGGGAGG CAAGCTGGCC
   CTC TGGGGGAC GGGTCCCTGG TGGCTGGGTG GTGGCCCTCC GTTCGACCGG
-----
551 AGCAACTTAT CTGTGTCTGT CCGATTGTCT AGTGTCTATG ACTGATTTTA
   TCGTTGAATA GACACAGACA GGCTAACAGA TCACAGATAC TGACTAAAT
-----
601 TCGCGCTGCG TCGGTACTAG TTAGCTRACT AGCTCTGTAT CTGGCGGACC
   ACGCGGACGC AGCCATGATC ATCTGATTGA TCGAGACATA GACCGCCTGG
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651 CGTGGTGGAA CTGACGAGTT CTGAACACCC GGCGGCAACC CTGGGAGACG
   GCACCACCTT GACTGTCTAA GACTTGTGGG CCGGCGTGG GACCTCTGC
-----
701 TCCCAGGGAC TTTGGGGGCC GTTTTGTGG CCCGACCTGA GGAAGGGAGT
   AGGGTCCCTG AAACCCCGG CAAAAACACC GGGCTGGACT CCTTCCCTCA
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751 CGATGTGGAA TCCGACCCG TCAGGATATG TGGTCTGGT AGGAGACGAG
   GCTACACCTT AGGCTGGGGC AGTCCTATAC ACCAAGACCA TCCTCTGCTC
-----
801 AACCTAAAC AGTTCCCGCC TCCGTCTGAA TTTTGTCTT CGGTTTGGAA
   TTGGATTTTG TCAAGGGCGG AGGCAGACTT AAAAACGAAA GCCAAACCTT
-----
851 CCGAAGCCGC GCGTCTTGTG TGCTGCAGCA TCGTTCTGTG TTGTCTCTGT
   GGCTTCGGCG CGCAGAACAG ACGACGTCGT AGCAAGACAC AACAGAGACA
-----
901 CTGACTGTGT TTCTGTATTT GCTGAAAAT TAGGGCCAGA CTGTTACCAC
   GACTGACACA AAGACATAAA CAGACTTTTA ATCCCGGTCT GACAAATGGTG
-----

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FIGURE 10B

951 TCGCTTAAGT TTGACCTTA/ GTAACCTGAA AGATGTGGAG CCGCTTCCTC
AGGGAATTCA AACTGGAATC CATTGACCTT TCTACAGCTC GCCGAGCGAG

1001 ACAACCAGTC GGTGATGTC AAGAAGAGAC GTTGGGTAC CTCTGCTCT
TGTGTGTCAG CCATCTACAG TTCTTCTCTG CAACCCAATG GAAGACGAGA

1051 GCAGAATGGC CAACCTTTAA CGTCGGATGG CCGCGAGACG GCACCTTTAA
CGTCTTACCG GTTGGAATTT GCAGCCTACC GGCCTCTGCT CGTGGAATTT

1101 CCGAGACCTC ATCACCAGG TTAAGATCAA GGTCTTTTCA CTTGGCCCCG
GGCTCTGGAG TAGTGGGTCC AATTCTAGTT CCAGAAAAGT GGACCGGGCG

1151 ATGGACACCC AGACCAGGTC CCCTACATCG TGACCTGGGA AGCCTTGGCT
TACCTGTGGG TCTGGTCCAG GGGATGTAGC ACTGGACCCT TCGGAACCGA

1201 TTTGACCCCC CTCCCTGGGT CAAGCCCTTT GTACACCCTA AGCCTCCGCC
AAACTGGGGG GAGGGACCCA GTTCGGGAAA CATGTGGGAT TCGGAGGCGG

1251 TCCTCTTCTT CCATCCGCCC CGTCTCTCCC CTTTGAACCT CCTCGTTTCA
AGGAGRAGGA GGTAGGCGGG GCAGAGAGGG GGAACCTGGA GGAGCAAGCT

1301 CCCCCTCTCG ATCCTCCCTT TATCCAGCCC TCACTCCTTC TCTAGGCGCC
GGGCGGAGC TAGGAGGGAA ATAGGTCCGG AGTGAGGAAG AGATCCGCGG

1351 GGCGCTCTTA GCCCATTAAT ACGACTCACT ATAGGGCGAT TCGAATCAGG
CCGCGAGAT CGGTAATTA TGCTGAGTGA TATCCCGCTA AGCTTAGTCC

1401 CCTTGGCGCG CCGGATCCTT AATTAGCGC AATTGGGAGG TGGCGGTAGC
GGAACCGCGC GGCCTAGGAA TTAATTGCGG TTAACCTOC ACCGCCATCG

+2 M G V I T D S L A V V A R T D
1451 CTCGAGATGG GCGTGATTAC GGATTCAGTG GCCGTCTGCG CCCGCACCGA
GAGCTCTACC CGCACTAATG CCTAAGTGAC CGGCAGCACC GGGCGTGGCT

+2 R P S Q Q L R S L N G E W R F A
1501 TCGCCCTTCC CAACAGTTAC GCAGCCTGAA TGGCGAATGG CGCTTGCCT
AGCGGGAAGG GTTGTCAATG CGTCGGACTT ACCGCTTACC GCGAAACGGA

+2 W F P A P E A V P E S W L E C D L
1551 GGTTCCTGGC ACCAGAAGCG GTGCCGGAAG GCTGGCTGGA GTGCGATCTT
CCAAAGGCCG TGGTCTTGGC CACGGCCTTT CGACCGACCT CACGCTAGAA

+2 P E A D T V V V P S N W Q M H G Y
1601 CCTGAGGCCG ATACTGTCTG CTCCCCTCA AACTGGCAGA TGCACGGTTA
GGACTCCGCG TATGACAGCA GCAGGGGAGT TTGACCGTCT ACGTGCCAT

+2 D A P I Y T N V T Y P I T V N P
1651 CGATGCGCCC ATCTACACCA ACGTGACCTA TCCCATIACG GTCAATCCGC
GCTACGCGGG TAGATGTGGT TGCACTGGAT AGGTAATGC CAGTTAGGCG

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+2 P F V P T E N P T G C Y S L T F N
1701 CGTTTGTTC CACGGAGAAT CCGACGGGTT GTTACTCGCT CACATTTAAT
GCAACAAGG GTGCCTCTTA GGCTGCCCAA CAATGAGCGA GTGTAAATTA
-----
+2 V D E S W L Q E G Q T R I I F D G
1751 GTTGATGAAA GCTGGCTACA GGAAGGCCAG ACGCGAATTA TTTTGATGG
CAACTACTTT CGACCGATGT CCTTCCGGTC TGGCTTAAT AAAAATACC
-----
+2 V N S A F H L W C N G R W V G Y
1801 CGTTAACTCG GCGTTTCATC TGTGGTGCAA CGGGCGCTGG GTCGGTTACG
GCAATTGAGC CGCAAAGTAG ACACACGTT GCCCGCGACC CAGCCAATGC
-----
+2 G Q D S R L P S E F D L S A F L R
1851 GCCAGGACAG TCGTTTGCCG TCTGAATTG ACCTGAGCGC ATTTTACGC
CGGTCTGTG AGCAAACGGC AGACTTAAAC TGGACTCGCG TAAAAATGCG
-----
+2 A G E N R L A V M V L R W S D G S
1901 GCCGGAGAAA ACCGCCTCGC GGTGATGGTG CTGCGCTGGA GTGACGGCAG
CGGCTCTTT TGGCGGAGCG CCACTACCAC GACGCGACCT CACTGCCGTC
-----
+2 Y L E D Q D M W R M S G I F R D
1951 TTATCTGGAA GATCAGGATA TGTGGCGGAT GAGCGGCATT TTCCGTGACG
AATAGACCTT CTAGTCTTAT ACACCGCCTA CTCGCCGTAA AAGGCACTGC
-----
+2 V S L L H K P T T Q I S D F H V A
2001 TCTCGTTGCT GCATAAACCG ACTACACAAA TCAGCGATTT CCATGTTGCC
AGAGCAACGA CGTATTGGC TGATGTGTTT AGTCGCTAAA GGTACAACGG
-----
+2 T R F N D D F S R A V L E A E V Q
2051 ACTCGCTTTA ATGATGATTT CAGCCGCGCT GTACTGGAGG CTGAAGTTCA
TGAGCGAAAT TACTACTAAA GTCGGCGCGA CATGACCTCC GACTTCAAGT
-----
+2 M C G E L R D Y L R V T V S L W
2101 GATGTGCGGC GAGTTGCGTG ACTACCTACG GGTAACAGTT TCTTTATGGC
CTACACGCCG CTCACGCAC TGATGGATGC CCATTGTCAA AGAAATACCG
-----
+2 Q G E T Q V A S G T A P F G G E I
2151 AGGGTGAAAC GCAGGTGCGC AGCGGCACCG CGCCTTTCGG CGGTGAAATT
TCCCACCTTG CGTCCAGCGG TCGCCGTGGC GCGGAAGCC GCCACTTTAA
-----
+2 I D E R G G Y A D R V T L R L N V
2201 ATCGATGAGC GTGGTGTTA TGCCGATCGC GTCACACTAC GTCTGAACGT
TAGCTACTCG CACCACCAAT ACGGCTAGCG CAGTGTGATG CAGACTTGCA
-----
+2 E N P K L W S A E I P N L Y R A
2251 CGAAAACCGG AACTGTGGA GCGCCGAAAT CCCGAATCTC TATCGTGCGG
GCTTTTGGGC TTTGACACCT CGCGGCTTTA GGGCTTAGAG ATAGCAGCGC

```

+2 V V E L H T A D G T L I E A E A C

2301 TGGTTGAAC GCACACCGCC GACGGCACGC TGATTGAAGC AGAAGCCTGC
ACCAACTTGA CGTGTGGCGG CTGCCGTGCG ACTAACTTCG TCTTCGGACG

+2 E V G F R E V R I E N G L L L L N

2351 GATGTCGGTT TCCGCGAGGT GCGGATTGAA AATGGTCTGC TGCTGCTGAA
CTACAGCCAA AGGCGCTCCA CGCCTAACTT TTACCAGACG ACGACGACTT

+2 G K P L L I R G V N R H E H H P

2401 CGGCAAGCCG TTGCTGATTC GAGGCGTTAA CCGTCACGAG CATCATCCTC
GCCGTTCGGC AACGACTAAG CTCCGCAATT GGCAGTGCTC GTAGTAGGAG

+2 L H G Q V M D E Q T M V Q D I L L

2451 TGCAATGGTCA GGTCAATGGAT GAGCAGACGA TGGTGCAGGA TATCCTGCTG
ACGTACCACT CCAGTACCTA CTCGTCTGCT ACCACGTCCT ATAGGACGAC

+2 M K Q N N F N A V R C S H Y P N H

2501 ATGAAGCAGA ACAACTTTAA CGCCGTGCGC TGTTCCGATT ATCCGAACCA
TACTTCGTCT TGTGAAATTT GCGGCACGCG ACAAGCGTAA TAGGCTTGGT

+2 P L W Y T L C D R Y G L Y V V D

2551 TCCGCTGTGG TACACGCTGT GCGACCGCTA CGGCCTGTAT GTGGTGGATG
AGGCGACACC ATGTGCGACA CGCTGGCGAT GCCGGACATA CACCACCTAC

+2 E A N I E T H G M V P M N R L T D

2601 AAGCCAATAT TGAACCCAC GGCATGGTGC CAATGAATCG TCTGACCGAT
TTGGTTATA ACTTTGGGTG CCGTACCACG GTTACTTAGC AGACTGGCTA

+2 D P R W L P A M S E R V T R M V Q

2651 GATCCGCGCT GGCTACCGGC GATGAGCGAA CGCGTAACGC GAATGGTGCA
CTAGGCGCGA CCGATGGCCG CTACTCGCTT GCGCATTGCG CTTACCACGT

+2 R D R N H P S V I I W S L G N E

2701 GCGCGATCGT AATCACCAGA GTGTGATCAT CTGGTCGCTG GGGAAATGAAT
CGCGCTAGCA TTAGTGGGCT CACACTAGTA GACCAGCGAC CCCTTACTTA

+2 S G H G A N H D A L Y R W I K S V

2751 CAGGCCACGG CGCTAATCAC GACGCGCTGT ATCGCTGGAT CAAATCTGTC
GTCCGGTGCC GCGATTAGTG CTGCGCGACA TAGCGACCTA GTTTAGACAG

+2 D P S R P V Q Y E G G G A D T T A

2801 GATCCTTCCC GCCCGGTGCA GTATGAAGGC GCGGAGCCG ACACCACGGC
CTAGGAAGGG CGGGCCACGT CATACTTCG CCGCCTCGGC TGTGGTGCCG

+2 T D I I C P M Y A R V D E D Q P

2851 CACCGATATT ATTGCCCCA TGTACGCGCG CGTGGATGAA GACCAGCCCT
GTGGCTATAA TAAACGGCT ACATGCGCGC GCACCTACTT CTGGTCGGGA

+2 F P A V P K W S I K K W L S L P G
 2901 TCCCGGCTGT GCCGAAATGG TCCATCAAAA AATGGCTTTC GCTACCTGGA
 AGGGCCGACA CGGCTTTACC AGGTAGTTT TTACCGAAAG CGATGGACCT

+2 E T R P L I L C E Y A H A N G N S
 2951 GAGACGCGCC CGCTGATCCT TTGCGAATAC GCCCAGCGCA TGGGTAACAG
 CTCTGCGCGG GCGACTAGGA AACGCTTATG CGGGTGCGCT ACCCATTTGC

+2 L G G F A K Y N Q A F R C Y P R
 3001 TCTTGGCGGT TTCGCTAAAT ACTGGCAGGC GTTTCGTACG TATCCCGTT
 AGAACCGCCA AAGCGATTTA TGACCGTCCG CAAAGCAGTC ATAGGGGCAA

+2 L Q G G F V W D W V D Q S L I K Y
 3051 TACAGGGCGG CTTGCTCTGG GACTGGGTGG ATCAGTCGCT GATTAAATAT
 ATGTCCCGCC GAAGCAGACC CTGACCCACC TAGTCAGCGA CTAATTTATA

+2 D E N G N P W S A Y G G D F G D T
 3101 GATGAAAACG GCAACCCGTG GTCGGCTTAC GCGGTGATT TTGGCGATAC
 CTACTTTTGC CGTTGGGCAC CAGCCGAATG CCGCCACTRA AACCGCTATG

+2 P N D R Q F C M N G L V F A D R
 3151 GCCGAACGAT CGCCAGTTCT GTATGAACGG TCTGGTCTTT GCCGACCGCA
 CGGCTTGCTA GCGGTCAAGA CATACTTGCC AGACCAGAAA CGGCTGGCGT

+2 T P H P A L T E A K H Q Q Q F F Q
 3201 CGCCGCATCC AGCGCTGACG GAAGCAAAC ACCAGCAGCA GTTTTCCAG
 GCGGCGTAGG TCGCGACTGC CTTGCTTTG TGGTCGTCGT CAAAAAGGTC

+2 F R L S G Q T I E V T S E Y L F R
 3251 TTCCGTTTAT CCGGGCAAAC CATCGAAGTG ACCAGCGAAT ACCTGTTCCG
 AAGGCAAATA GGGCCGTTG GTAGCTTCAC TGGTCGCTTA TGGACAAAGG

+2 H S D N E L L E W M V A L D G K
 3301 TCATAGCGAT AACGAGCTCC TGCCTGGAT GGTGGCGCTG GATGGTAAGC
 AGTATCGCTA TTGCTCGAGG ACGTGACCTA CCACCGCGAC CTACCATTOG

+2 P L A S G E V P L D V A P Q G K Q
 3351 CGCTGGCAAG CGGTGAAGTG CCTCTGGATG TCGCTCCACA AGGTAAACAG
 GCGACCGTTC GCCACTTCAC GGAGACCTAC ACCGAGGTGT TCCATTTGTC

+2 L I E L P E L P Q P E S A G Q L W
 3401 TTGATTGAAC TGCCTGAACT ACCGCAGCCG GAGAGCGCCG GGCAACTCTG
 AACTAACTTG ACGGACTTGA TGGCGTCGGC CTCTCGCGGC CCGTTGAGAC

+2 L T V R V V Q P N A T A W S E A
 3451 GCTCACAGTA CGGTAGTGC AACCGAACGC GACCGCATGG TCAGAAGCCG
 CGAGTGTCTAT GCGCATCAGG TTGGCTTGGC CTGGCGTACC AGTCTTCGGC

+2 G H I S A W Q Q W R L A E N L S V
 3501 GGCACATCAG CGCCTGGCAG CAGTGGCGTC TGGCGGAAAA CCTCAGTGTG
 CCGTGTAGTC GCGGACCGTC GTCACCGCAG ACCGCCTTIT GGAGTCACAC

 +2 T L P A A S E A I P H L T T S E M
 3551 ACGCTCCCCG CCGCGTCCCA CGCCATCCCG CATCTGACCA CCAGCGAAAT
 TGCGAGGGGC GCGCGAGGGT GCGGTAGGGC GTAGACTGGT GGTGCGTTTA

 +2 D F C I E L G N K R W Q F N R Q
 3601 GGATTTTTCG ATCGAGCTGG GTAATAAGCG TTGGCAATTT AACCGCCAGT
 CCTAAAAACG TAGCTCGACC CATTATTGCG AACCGTTAAA TTGGCGGTCA

 +2 S G F L S Q M W I G D K K Q L L T
 3651 CAGGCTTTCT TTCACAGATG TGGATTGGCG ATAAAAACA ACTGCTGACG
 GTCCGAAAGA AAGTGTCTAC ACCTAACCGC TATTTTTTGT TGACGACTGC

 +2 P L R D Q F T R A P L D N D I G V
 3701 CCGCTGCGCG ATCAGTTCAC CCGTGCACCG CTGGATAACG ACATTGGCGT
 GCGGACGCGC TAGTCAAGTG GGCACGTGGC GACCTATTGC TGTAAACGCA

 +2 S E A T R I D P N A W V E R W K
 3751 AAGTGAAGCG ACCCGCATTC ACCCTAACGC CTGGGTGAA CGCTGGAGG
 TTCATTGCG TGGGCGTAAC TGGGATTGCG GACCCAGCTT GCGACCTTC

 +2 A A G H Y Q A E A A L L Q C T A D
 3801 CCGCGGGCCA TTACCAGGCC GAAGCAGCGT TGTTCAGTG CACGGCAGAT
 GCCGCCCGGT AATGGTCCGG CTTCGTCCGA ACAACGTCAC GTGCCGTCTA

 +2 T L A D A V L I T T A H A W Q H Q
 3851 ACACTTGCTG ATGCGGTGCT GATTACGACC GCTCACGGT GGCAGCATCA
 TGTGAACGAC TACGCCACGA CTAATGCTGG CGAGTGCACA CCGTCGTAGT

 +2 G K T L F I S R K T Y R I D G S
 3901 GGGGAAAACC TTATTATCA GCCGAAAAC CTACCGGATT GATGGTAGTG
 CCCCTTTTGG AATAAATAGT CGGCCTTTTG GATGGCTTAA CTACCATCAC

 +2 G Q M A I T V D V E V A S D T P H
 3951 GTCAATGGC GATTACCGTT GATGTTGAAG TGGCGAGCGA TACACCGCAT
 CAGTTTACCG CTAATGGCAA CTACAACTTC ACCGCTCGCT ATGTGGCGTA

 +2 P A R I G L N C Q L A Q V A E R V
 4001 CCGGCGCGGA TTGGCCTGAA CTGCCAGCTG GCGCAGGTAG CAGAGCGGGT
 GGCGCGCGCT AACCGGACTT GACGGTCGAC CGCGTCCATC GTCTCGCCCA

 +2 N W L G L G P Q E N Y P D R L T
 4051 AAACCTGGCTC GGATTAGGGC CGCAAGAAAA CTATCCCGAC CGCCTTACTG
 TTTACCGAG CTAATCCCG GCGTTCTTTT GATAGGGCTG GCGGAATGAC

+2 A A C F D R W D L P L S D M Y T P

4101 CCGCCTGTTT TGACCGCTGG GATCTGCCAT TGTCAGACAT GTATACCCCG
GGCGGACAAA ACTGGCGACC CTAGACGGTA ACAGTCTGTA CATATGGGGC

+2 Y V F P S E N G L R C G T R E L N

4151 TACGTCTTCC CGAGCGAAAA CGGTCTGCGC TCGGGGACGC GCGAATTGAA
ATGCAGAAGG GCTCGCTTTT GCCAGACGCG ACGCCCTGCG CGCTTAACCT

+2 Y G P H Q W R G D F Q F N I S R

4201 TTATGGCCCA CACCAGTGGC GCGGCGACTT CCAGTTCAAC ATCAGCCGCT
AATACCGGGT GTGGTCACCG CGCCGCTGAA GGTCAGTTG TAGTCGGCGA

+2 Y S Q Q Q L M E T S H R H L L H A

4251 ACAGTCAACA GCAACTGATG GAAACCAGCC ATCGCCATCT GCTGCACGCG
TGTCAGTTGT CGTTGACTAC CTTTGGTCCG TAGCGGTAGA CGACGTGCGC

+2 E E G T W L N I D G F H M G I G G

4301 GAAGAAGGCA CATGGCTGAA TATCGACGGT TTCCATATGG GGATTGGTGG
CTTCTTCCGT GTACCGACTT ATAGCTGQCA AAGGTATAAC CCTAACACC

+2 D D S W S P S V S A E F Q L S A

4351 CGACGACTCC TGGAGCCCGT CAGTATCGGC GGAATTCCAG CTGAGCGCCG
GCTGCTGAGG ACCTCGGGCA GTCATAGCCG CCTTAAGGTC GACTCGCGGC

+2 G R Y H Y Q L V W C Q K R S D Y K

4401 GTCGGTACCA TTACCAGTTG GTCTGGTGTC AAAAAAGATC TGACTATAAA
CAGCGATGGT AATGGTCAAC CAGACCACAG TTTTCTTAG ACTGATATT

+2 D E D L D H H H H H H R
----->
4451 GATGAGGACC TCGACCATCA TCATCATCAT CACCGGTAAT AATAGGTAGA
CTACTCCTGG AGCTGGTAGT AGTAGTAGTA GTGGCCATTA TTATCCATCT

4501 TAAGTGACTG ATTAGATGCA TTGATCCCTC GACCAATICC GGTATTATTC
ATTCACTGAC TAATCTACGT AACTAGGGAG CTGGTTAAGG CCAATAAAAG

4551 CACCATATTG CCGTCTTTTG GCAATGTGAG GGCCCGGAAA CCTGGCCCTG
GTGGTATAAC GGCAGAAAAC CGTTACACTC CCGGGCCTTT GGACCGGGAC

4601 TCTTCTTGAC GAGCATTCCT AGGGGTCTTT CCCCTCTCGC CAAAGGAATG
AGAAGAAGTG CTCGTAAGGA TCCCCAGAAA GGGGAGAGCG GTTTCCTTAC

4651 CAAGGTCTGT TGAATGTCGT GAAGGAAGCA GTTCTCTGAG AAGCTTCTTG
GTTCCAGACA ACTTACAGCA CTTCTTCGT CAAGGAGACC TTCGAGAAG

4701 AAGCAAAACA ACGTCTGTAG CGACCCCTTG CAGGCAGCGG AACCCCCRC
TTCGTGTTGT TGCAGACATC GCTGGGAAAC GTCCGTCGCC TTGGGGGGTG

4751 CTGGCGACAG GTGCCTCTGC GGCCAAAAGC CACGTGATA AGATACACCT
GACCGCTGTC CACGGAGACG CCGGTTTTCG GTGCACATAT TCTATGTGGA

4801 GCAAAGGCGG CACAACCCCA GTGCCACGTT GTGAGTTGGA TAGTTGTGGA
CGTTTCCGCC GTGTTGGGGT CACGGTGCAA CACTCAACCT ATCAACACCT

4851 AAGAGTCAAA TGGCTCTCCT CAAGCGTATT CAACAAGCGG CTGAAGGATG
TTCTCAGTTT ACCGAGAGGA GTTCGCATAA GTTGTTCOCG GACTTCCTAC

4901 CCCAGAAAGG ACCCCATTGT ATGGGATCTG ATCTGGGGCC TCGGTGCACA
GGGTCTTCCA TGGGGTAACA TACCCTAGAC TAGACCCCGG AGCCACGTGT

4951 TGCTTTACAT GTGTTTAGTC GAGGTTAAAA AACGTCTAGG CCCCCCGAAC
ACGAAATGTA CACAAATCAG CTCCAATTTT TTGCAGATCC GGGGGGCTTG

5001 CACGGGGACG TGGTTTTCTT TTGAAAAACA CGATGATAAT ACCATGATTG
GTGCCCCGCG ACCAAAAGGA AACTTTTGT GCTACTATTA TGGTACTAAC

5051 AACAGATGG ATTGCACGCA GGTCTCTCCG CCGCTTGGCT GGAGAGGCTA
TTGTTCTACC TAACGTGCGT CCAAGAGGCC GCGCAACCCA CCTCTCCGAT

5101 TTGGGCTATG ACTGGGCACA ACAGACAATC GGCTGCTCTG ATGCCGCCGT
AAGCCGATAC TGACCCGTGT TGTCTGTTAG CCGACGAGAC TACGGCGGCA

5151 GTTCCGGCTG TCAGCGCAGG GCGGCCCGGT TCTTTTGTG AAGACCGACC
CAAGGCCGAC AGTCGCGTCC CCGCGGGCCA AGAAAAACAG TTCTGGCTGG

5201 TGTCCGGTGC CCTGAATGAA CTGCAGGACG AGGCAGCGCG GCTATCGTGG
ACAGGCCAAG GGACTTACTT GACGTCTGTC TCCGTGCGCG CGATAGCACC

5251 CTGGCCACGA CGGGCGTTCC TTGCGCAGCT GTGCTCGACG TTGTCACTGA
GACCGGTGCT GCCCGCAAGG AACGCGTGA CACGAGCTGC AACAGTGAAT

5301 AGCGGGAAGG GACTGGCTGC TATTGGGCGA AGTGCCGGGG CAGGATCTCC
TCGCCCCTCC CTGACCGACG ATAACCGGCT TCACGGCCCC GTCTAGAGG

5351 TGTCATCTCA CCTTGCTCCT GCCGAGAAAG TATCCATCAT GGCTGATGCA
ACAGTAGAGT GGAACGAGGA CGGCTCTTTC ATAGGTAGTA CCGACTACGT

5401 ATGCGGCGGC TGCATACGCT TGATCCGGCT ACCTGCCCAT TCGACCACCA
TACGCCGCGG ACGTATGCGA ACTAGGCCGA TGGACGGGTA AGCTGGTGGT

5451 AGCGAAACAT CGCATCGAGC GAGCACGTAC TCGGATGGAA GCCGGTCTTG
TCGCTTTGTA GCGTAGCTCG CTCGTGCATG AGCCTACCTT CGGCCAGAAC

5501 TCGATCAGGA TGATCTGGAC GAAGAGGATC ACGGGCTCGC GCCAGCCGAA
AGCTAGTCCT ACTAGACCTG CTTCTCGTAG TCCCCGAGCG CGGTCCGCTT

5551 CTGTTCCGCA GGCTCAAGGC GCGCATGCCC GACGGCGAGG ATCTCGTCTG
GACAGCGGCT CCGAGTTCOG CCGGTACGGG CTGCCGCTCC TAGAGCAGCA

5601 GACCCATGGC GATGCTGCT TGCCGAATAT CATGGTGGAA AATGGCCGCT
CTGGGTACCG CTACGGACGA ACGGCTTATA GTACCACTT TTACCGGCGA

5651 TTTCTGGATT CATCGACTGT GGCCGGCTGG GTGTGGGCGA CCGCTATCAG
AAGACCTAA GTAGCTGACA CCGGCCGACC CACACCGCCT GCGATAGTC

5701 GACATAGCGT TGGCTACCCG TGATATTGCT GAAGAGCTTG GCGGCGAATG
CTGTATCGCA ACCGATGGGC ACTATAACGA CTTCTCGAAC CCGCGCTTAC

5751 GGCTGACCGC TTCCTCGTGC TTTACGGTAT CGCCGCTCCC GATTCCGAGC
CCGACTGGCG AAGGAGCACG AAATGCCATA GCGGCGAGGG CTAAGCGTCG

5801 GCATCGCCTT CTATCGCCTT CTGACGAGT TCTTCTGAGC GGGACTCTGG
CGTAGCGGAA GATAGCGGAA GAACTGCTCA AGAAGACTCG CCTTGAGACC

5851 GGTTGCGATC GATAAAATAA AAGATTTTAT TTAGTCTCCA GAAAAAGGGG
CCAAGCGTAG CTATTTTAT TTTAAATA AATCAGAGGT CTTTTTCCCC

5901 GGAATGAAAG ACCCCACCTG TAGGTTTGGC AAGCTAGCTT AAGTAACGCC
CCTTACTTTC TGGGGTGGAC ATCCAAACCG TTCGATCGAA TTCATTGCGG

5951 ATTTTGCAAG GCATGGAAAA ATACATAACT GAGAATAGAG AAGTTCAGAT
TAAACGTTTC CGTACCTTTT TATGTATTGA CTCTTATCTC TTCAAGTCTA

6001 CAAGGTCAAG AACAGATGGA ACAGCTGAAT ATGGGCCAAA CAGGATATCT
GTTCCAGTCC TTGTCTACCT TGTCGACTTA TACCCGGTTT GTCTATAGA

6051 GTGGTAAGCA GTTCTGCCC CGGCTCAGGG CCAAGAACAG ATGGAACAGC
CACCATTCTG CAAGGACGGG GCCGAGTCCC GGTCTTTGTC TACCTTGTCG

6101 TGAATATGGG CCAAACAGGA TATCTGTGGT AAGCAGTTCC TGCCCCGGCT
ACTTATACCC GGTGTGTCCT ATAGACACCA TTCGTCAAGG ACGGGGCGGA

6151 CAGGGCCAAG AACAGATGGT CCCAGATGC GGTCCAGCCC TCAGCAGTTT
GTCCCGGTTT TTGTCTACCA GGGGTCTACG CCAGGTGCGG AGTCGTCAA

6201 CTAGAGAACC ATCAGATGTT TCCAGGTGTC CCCAAGGACC TGAATGACC
GATCTCTGG TAGCTACAA AGGTCCACG GGGTCTCTGG ACTTTACTGG

6251 CTGTGCCTTA TTTGAACATA CCAATCAGTT CGTTCTCGC TTCTGTTGCG
GACACGGAAT AAATTTGATT GGTAGTCAA GCGAAGAGCG AAGACAAGCG

6301 GCGCTTCTGC TCCCGAGCT CAATAAAGA GCCACAACC CCTCACTCGG
CGCGAAGACG AGGGGCTCGA GTTATTTTCT CGGGTGTGG GGAGTGAGCC

6351 GGCGCCAGTC CTCCGATTGA CTGAGTCGCC CGGGTACCGG TGTATCCAAT
CCGCGGTCAG GAGGCTAACT GACTCAGCGG GCCCATGGGC ACATAGGTTA

6401 AAACCTCTT CGAGTTGCAT CCGACTTGTG GTCTCGCTGT TCCTTGGGAG
TTTGGGAGAA CGTCAACGTA GGCTGAACAC CAGAGCGACA AGGAACCTC

6451 GGTCTCCTCT GAGTGATTGA CTACCGTCA GCGGGGGTCT TTCATTCTG
CCAGAGGAGA CTCCTAACT GATGGGCACT CGCCCCAGA AAGTAAGTAC

6501 CAGCATGTAT CAAAATTAAT TTGGTTTTTT TTCTTAAGTA TTTACATTAA
GTCGTACATA GTTTAATTA AACCAAAAAA AAGAATTCAT AATGTAATT

6551 ATGGCCATAG TTGCATTAAAT GAATCGGCCA ACGCGCGGGG AGAGGCGGTT
TACCGGTATC AACGTAATTA CTAGCCGGT TCGCGGCCCC TCTCCGCCAA

6601 TGCGTATTGG CGCTTTCCG CTTCCTCGCT CACTGACTCG CTGCGCTCGG
ACGCATAACC GCGAGAAGGC GAAGGAGCGA GTGACTGAGC GACGCGAGCC

6651 TCGTTCCGGT CCGCGGAGCG GTATCAGCTC ACTCAAAGGC GGTAAATACGG
AGCAAGCCGA CGCCGCTCGC CATAGTCGAG TGAGTTTCCG CCATTATGCC

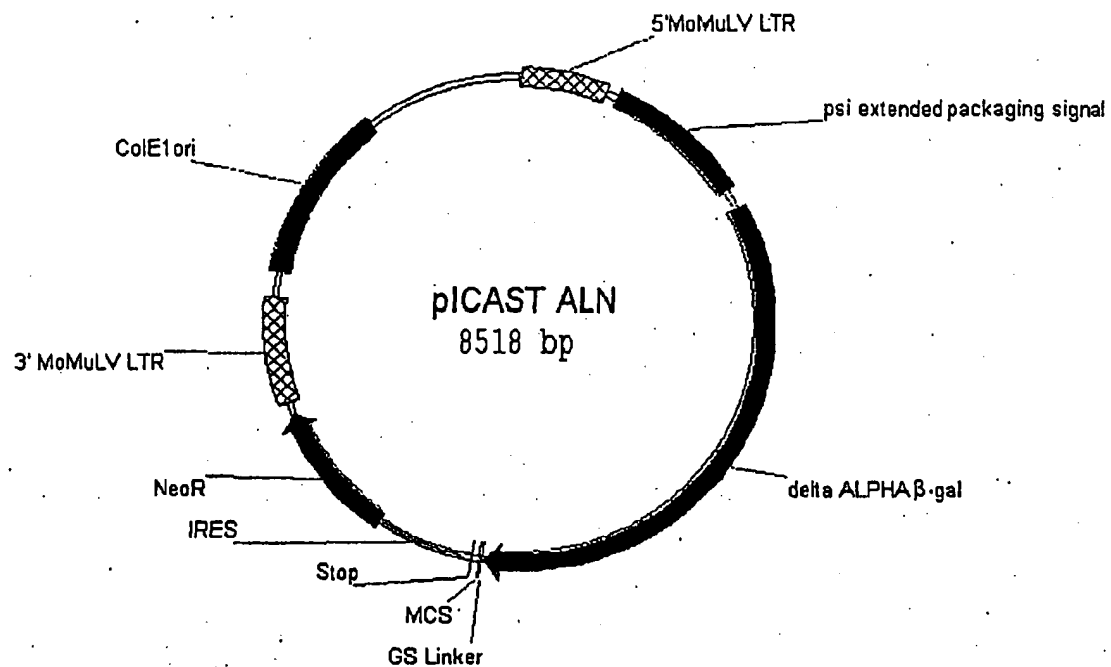


Figure 11A

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1  CTGCAGCCTG AATATGGGCC AAACAGGATA TCTGTGGTAA GCAGTTCTCTG
   GACGTCGGAC TTATACCCGG TTTGTCTTAT AGACACCATT CGTCAAGGAC
-----
51  CCCCCGCTCA GGGCCAAGAA CAGATGGAAC AGCTGAATAT GGGCCAAACA
   GGGGCCGAGT CCGGTTCTT GTCTACCTTG TCGACTTATA CCGGTTTGT
-----
101 GGATATCTGT GGTAAAGCAGT TCCTGCCCCG GCTCAGGGCC AAGAACAGAT
   CCTATAGACA CCATTCTGTC AGGACGGGGC CGAGTCCCGG TTCTTGCTA
-----
151 GGTCCCCAGA TGCGGTCCAG CCCTCAGCAG TTTCTAGAGA ACCATCAGAT
   CCAGGGGTCT ACGCCAGGTC GGGAGTCGTC AAAGATCTCT TGGTAGTCTA
-----
201 GTTCCAGGGG TGCCCCAAGG ACCTGAAATG ACCCTGTGCC TTATTTGAAC
   CAAAGGTCCC ACGGGGTTCC TGGACTTTAC TGGGACACGG AATAAACTTG
-----
251 TAACCAATCA GTTCGCTTCT CGCTTCTGTT CGCGCGCTTC TGCTCCCCGA
   ATTGGTTAGT CAAGCGAAGA GCGAAGACAA GCGCGGAAG ACGAGGGGCT
-----
301 GCTCAATAAA AGAGCCCAACA ACCCTCACT CGGGCGGCCA GTCTCTCGAT
   CGAGTTATTT TCTCGGGTGT TGGGGAGTGA GCGCCCGGT CAGGAGGCTA
-----
351 TGACTGAGTC GCGCGGTAC CCGTGATCC AATAAACCTT CTTGCAGTTG
   ACTGACTCAG CGGGCCCATG GGCACATAGG TTATTTGGGA GAACGTCAAC
-----
401 CATCCGACTT GTGGTCTCGC TGTTCCTTGG GAGGGTCTCC TCTGAGTGAT
   GTAGGCTGAA CACCAGAGCG ACAAGGAACC CTCCAGAGG AGACTCACTA
-----
451 TGACTACCCG TCAGCGGGGG TCTTTCATTT GGGGGCTCGT CCGGGATCGG
   ACTGATGGGC AGTCGCCCCC AGAAAGTAAA CCCCCAGCA GGCCTAGCC
-----
501 GAGACCCCTG CCCAGGGACC ACCGACCCAC CACCGGGAGG CAAGCTGGCC
   CTCTGGGGAC GGTCCCTGG TGGCTGGGTG GTGGCCCTCC GTTCGACCGG
-----
551 AGCAACTTAT CTGTGTCTGT CCGATTGTCT AGTGTCTATG ACTGATTTA
   TCGTTGAATA GACACAGACA GGCTAACAGA TCACAGATAC TGACTAAAAT
-----
601 TGCGCCTGCG TCGGTACTAG TTAGCTAACT AGCTCTGTAT CTGGCGGACC
   ACGCGGACGC AGCCATGATC AATCGATTGA TCGAGACATA GACCGCTTGG
-----
651 CGTGGTGGAA CTGACGAGTT CTGAACACCC GGCGCGCAACC CTGGGAGACG
   GCACCACTT GACTGCTCAA GACTTGTGGG CCGGCGTTGG GACCTCTGC
-----
701 TCCAGGGGAC TTTGGGGGCC GTTTTGTGG CCGCACTGA GGAAGGGAGT
   AGGGTCCCTG AAACCCCGG CAAAACACC GGGCTGGACT CCTTCCCTCA
-----
751 CGATGTGGAA TCCGACCCCG TCAGGATATG TGGTCTGGT AGGAGACGAG
   GCTACACCTT AGGCTGGGGC AGTCTTATAC ACCAAGACCA TCCTCTGCTC
-----
801 AACCTAAAC AGTTCCCGCC TCCGTCTGAA TTTTGTCTT CGGTTTGGAA
   TTGGATTTT TCAAGGGCGG AGGCAGACTT AAAACGAA GCGCAACCTT
-----
851 CCGAAGCCGC GCGTCTTGTG TGCTGCAGCA TCGTCTGTG TTGTCTCTGT
   GGCTTCGGCG CGCAGAACAG ACGACGTCGT AGCAAGACAC AACAGAGACA
-----
901 CTGACTGTGT TTCTGTATTT GTCTGAAAT TAGGGCCAGA CTGTTACCAC
   GACTGACACA AAGACATAAA CAGACTTTTA ATCCCGGTCT GACAATGGTG
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FIGURE 11B

951 TCCCTTAAGT TTGACCTTAG GTAACCTGGAA AGATGTCGAG CGGCTCGCTC
AGGGAATTCA AACTGGAATC CATTGACCTT TCTACAGCTC GCCGAGCGAG

1001 ACAACCAGTC GGTAGATGTC AAGAAGAGAC GTTGGGTTAC CTTCTGCTCT
TGTTGGTCTAG CCATCTACAG TTCTTCTCTG CAACCCAAATG GAAGACGAGA

1051 GCAGAATGGC CAACCTTTAA CGTCGGATGG CCGCGAGACG GCACCTTTAA
CGTCTTACCG GTTGGAAATT GCAGCCTACC GGCCTCTGCTC CGTGGAAATT

1101 CCGAGACCTC ATCACCAGG TTAAGATCAA GGTCTTTTCA CCTGGCCCCG
GGCTCTGGAG TAGTGGGTCC AATTCTAGTT CCAGAAAAGT GGACCGGGCG

1151 ATGGACACCC AGACCAGGTC CCCTACATCG TGACCTGGGA AGCCTTGGCT
TACCTGTGGG TCTGGTCCAG GGGATGTAGC ACTGGACCCT TCGGAACCGA

1201 TTGACCCCC CTCCCTGGGT CAGCCCTTT GTACACCTTA AGCCTCCGCC
AAACTGGGGG GAGGGACCCA GTTCGGGAAA CATGTGGGAT TCGGAGGCGG

1251 TCCTCTTCTT CCATCCGCCG CGTCTCTCCC CCTTGAACCT CCTCGTTTGA
AGGAGAAGGA GGTAGGCGGG GCAGAGAGGG GGAACCTTGA GGAGCAAGCT

1301 CCCCGGCTCG ATCCTCCCTT TATCCAGCCC TCACTCCTTC TCTAGGCGCG
GGGGCGGAGC TAGGAGGGAA ATAGGTCTGG AGTGAGGAAG AGATCCGCGG

1351 GGCGCTCTTA GCCCATTAAT ACGACTCACT ATAGGGCGAT TCGAACACCA
CGGCGAGAT CGGGTAATTA TGCTGAGTGA TATCCCGCTA AGCTTGTGGT

1401 TGCACCATCA TCATCATCAC GTCGACTATA AAGATGAGGA CCTCGAGATG
ACGTGGTAGT AGTAGTAGTG CAGCTGATAT TTCTACTCCT GGAGCTCTAC

1451 GGCGTGATTA CGGATTCCT GCGCGTCGTG GCCCGCACCG ATCGCCCTTC
CCGCACTAAT GCCTAAGTGA CCGGCAGCAC CGGCGGTGTC TAGCGGGAAG

1501 CCAACAGTTA CGCAGCCTGA ATGGCGAATG GCGCTTGGC TGGTTCCGG
GGTTGTCAAT GCGTCGGACT TACCGCTTAC CGCGAAACGG ACCAAAGGCC

1551 CACCAGAAAG GGTGCCGGAA AGCTG6CTGG AGTGCGATCT TCCTGAGGCC
GTGGTCTTOS CCACGGCCTT TCGACCGACC TCACGCTAGA AGGACTCCGG

1601 GATACTGTOS TCGTCCCCTC AACTGGCAG ATGCACGGTT ACGATGCGCC
CTATGACAGC AGCAGGGGAG TTTGACCGTC TACGTGCCAA TGCTACGCGG

1651 CATCTACACC AACGTGACCT ATCCCAATTAC GGTCAATCAG CCGTTTGTTC
GTAGATGTGG TTGCACTGGA TAGGGTAATG CCACTTAGGC GGCACCAAG

1701 CCACGGAGAA TCCGACGGGT TGTTACTCGC TCACATTTAA TGTGATGAA
GGTGCTCTT AGCCTGCCCA ACAATGAGCG AGTGTAATTT ACAACTACTT

1751 AGCTGGCTAC AGGAAGGCCA GACGCGAATT ATTTTGTATG GCGTTAATC
TCGACCGATG TCCTTCCGGT CTGCGCTTAA TAAAACTAC CGCAATTGAG

1801 GGCGTTTCAT CTGTGGTGCA ACGGGCGCTG GGTGGGTAC GGCAGGACA
CCGCAAGTA GACACCAGT TGCCCGGAC CCAGCCAATG CCGTCTCTGT

1851 GTCGTTTGCC GTCTGAATTT GACCTGAGCG CATTTTACG CGCCGAGAA
CAGCAACGG CAGACTTAAA CTGGACTCGC GTAAAAATTC GCGCCTCTT

1901 AACCGCCTCG CGGTGATGGT GCTGGGCTGG AGTGACGGCA GTTATCTGGA
TTGGCGGAGC GCCACTACCA CGATGAGACC TCACTGCCGT CAATAGACCT

1951 AGATCAGGAT ATGTGGCGGA TGAGCGGCAT TTTCCGTGAC GTCTCGTTGC
TCTAGTCCTA TACACCGCCT ACTCGCCGTA AAAGGCACTG CAGAGCAACG

2001 TGCATAAACC GACTACACAA ATCAGCGATT TCCATGTTGC CACTCGCTTT
ACGTATTGG CTGATGTGTT TAGTCGCTAA AGGTACACG GTGAGCGAAA

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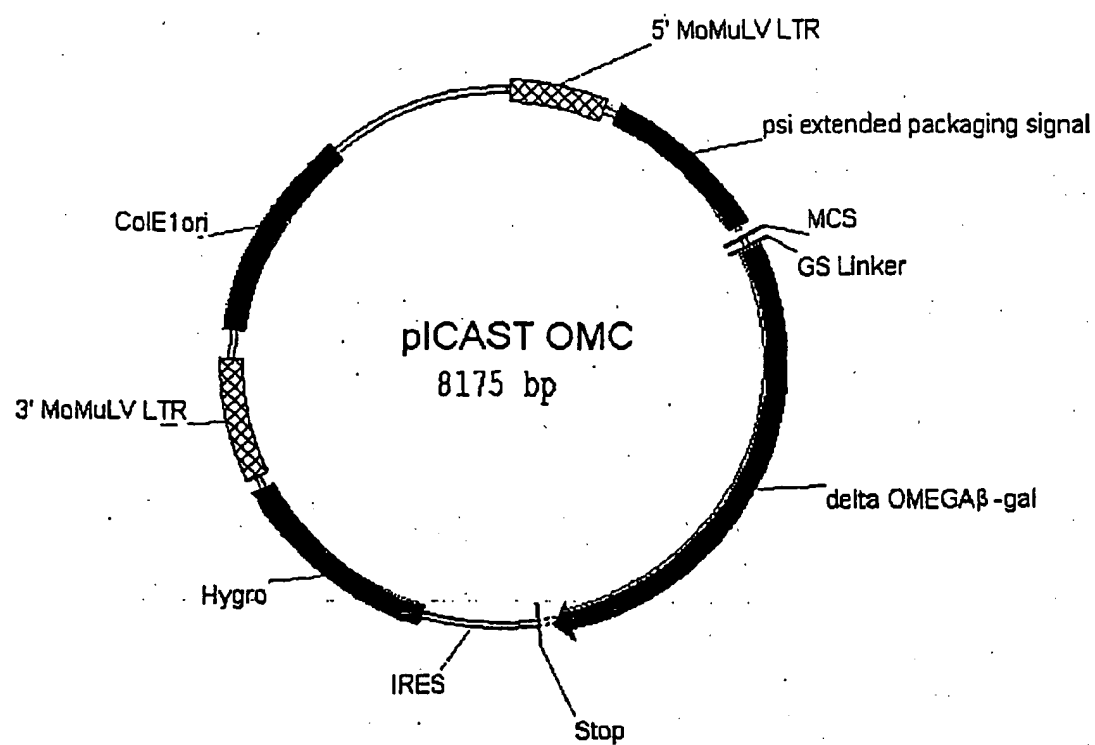


Figure 12A

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751 CGATGTGGAA TCCGACCCCG TCAGGATATG TGTTCTGCT AGGAGACGAG
   GCTACACCTT AGGCTGGGGC AGTCTTATAC ACCAAGACCA TCCTCTGCTC
-----
801 AACCTAAAC AGTCCCGGC TCCGTCTGAA TTTTGTCTT OGGTTGGAA
   TTGGATTTT TCAAGGGCGG AGGCGACTT AAAACGAAA GCCAAACCTT
-----
851 CCGAAGCGCG GCGTCTTGTG TGCTGCAGCA TCGTTCTGTG TTGTCTCTGT
   GGGTTCGGCG CGCAGAACAG ACGACGTCGT AGCAAGACAC AACAGAGCA
-----
901 CTGACTGTGT TTCTGTATTT GTCTGAAAT TAGGGCCAGA CTGTACCAC
   GACTGACACA AAGACATAAA CAGACTTTTA ATCCCGTCT GACAATGGTG
-----

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FIGURE 12B

951 TCCCTTAACT TTAGCCTTAG GTAACGGAA AGATGTCGAG CGGCTCGCTC
AGGGAATTCA AACTGGAATC CATTGACCTT TCTACAGCTC GCCGAGCGAG

1001 ACAACCACTC GGTAGATGTC AAGAAGAGAC GTTGGGTAC CTTCTGCTCT
TGTTGGTCAG CCATCTACAG TTCTTCTCTG CAACCCAATG GAAGACGAGA

1051 GCAGAATGGC CAACCTTTAA CGTCGGATGG CCGCGAGACG GCACCTTTAA
CGTCTTACCG GTTGGAAATT GCAGCCTACC GCGCTCTGCG CGTGGAAATT

1101 CCGAGACCTC ATCACCAGG TTAAGATCAA GGTCTTTTCA CCTGGCCCGC
GGCTCTGGAG TAGTGGGTCC AATTCTAGTT CCAGAAAAGT GGACCGGGCG

1151 ATGGACACCC AGACCAGGTC CCTTACATCG TGAACGGGA AGCCTTGGCT
TACCTGTGGG TCTGGTCCAG GGGATGTAGC ACTGGACCCT TCGGACCGA

1201 TTTGACCCCC CTCCTGGGT CAAGCCCTTT GTACCCCTA AGCCTCCGCC
AAACTGGGGG GAGGGACCCA GTTCGGGAAA CATGTGGGAT TCGGAGGCGG

1251 TCCTCTTCCT CCATCCGCCC CGTCTCTCCC CCTTGAACT CCTCGTTCGA
AGGAGAAGGA GGTAGGCGGG GCAGAGAGGG GGAACCTTGA GGAGCAAGCT

1301 CCCCCTCTCG ATCTCCCTT TATCCAGCCC TCACTCCTTC TCTAGGCGCC
GGGCGCGAGC TAGGAGGGA ATAGGTCGGG AGTGAGGAAG AGATCCGCGG

1351 GCGCGCTCTA GCCATTAAAT ACGACTCACT ATAGGGCGAT TCGAATCAGG
CCGGCGAGAT CCGGTAATTA TGCTGAGTGA TATCCGCTA AGCTTAGTCC

1401 CCTTGGCGCG CCGGATCCTT AATTAAGCGC AATTGGGAGG TGGCGGTAGC
GGAACCGCGC GGCCTAGGAA TTAATTGCGG TTAACCTCC ACCGCCATCG

1451 CTCGAGATGG GCGTGATTAC GGATTCACTG GCGCTCGTTT TACAACGTCG
GAGCTCTACC CGCACTAATG CCTAAGTGAC CGGCAGCAAA ATGTTCAGC

1501 TGACTGGGAA AACCTGGCG TTACCCAAT TAATCGCCTT GCAGCACATC
ACTGACCTT TTGGGACCGC AATGGGTGA ATTAGCGGAA CGTGTGTAG

1551 CCCCTTTCGC CAGCTGGCGT AATAGCGAAG AGGCCCGCAC CGATCGCCCT
GGGGAAGCG GTCGACCGCA TTATCGCTTC TCCGGGCGTG GCTAGCGGGA

1601 TCCCAACAGT TACGCAGCCT GAATGGCGAA TGGCGCTTTG CCTGGTTTCC
AGGGTTGTCA ATGCGTCGGA CTTACCGCTT ACCGCGAAAC GGACCAAGG

1651 GGCACCAGAA GCGGTGCCGG AAAGCTGGCT GGAGTGCAT CTTCTGAGG
CCGTGGTCTT CGGCACGGCC TTTCGACCGA CCTCAGCTA GAAGGACTCC

1701 CCGATACTGT CGTCGTCCCC TCRAACTGGC AGATGCACGG TTACGATCGG
GGCTATGACA GCAGCAGGGG AGTTTGACCG TCTACGTGCC AATGCTACGC

1751 OCCATCTACA CCAACGTGAC CTATCCCAT ACGGTCAATC CGCCGTTTGT
GGTAGATGT GGTGCACTG GATAGGGTAA TGCCAGTTAG GCGGCAACA

1801 TCCCACGGAG AATCCGACGG GTTGTACTC GCTCACATT AATGTTGATG
AGGGTGCCCT TTAGGCTGCC CAACAATGAG CGAGTGTAAT TTACAATAC

1851 AAAGCTGGCT ACAGGAAGGC CAGACGCGAA TTATTTTGA TGGCGTTAAC
TTTCGACCGA TGTCTTCCG GTCTGCGCTT AATAAAACT ACCGCAATG

1901 TCGGCGTTTC ATCTGTGGTG CAACGGGCGC TGGGTCGGTT ACGGCCAGGA
AGCCGCAAAAG TAGACACCAC GTTGCCCGCG ACCCAGCCAA TGCCGGTCCT

1951 CAGTCGTTTG CCGTCTGAAT TTGACCTGAG CGCATTTTTA CGCGCCGGAG
GTCAGCAAAC GGCAGACTTA AACTGGACTC GCGTAAAAAT GCGCGGCCTC

2001 AAAACCGCCT CCGGCTGATG GTGCTGCGCT GGAGTGACGG CAGTTATCTG
TTTTGGCGGA GCGCCACTAC CACGACGCGA CCTCACTGCC GTCATAGAC

2051 GAAGATCAGG ATATGTGGCG GATGAGCGGC ATTTCCGTG ACGTCTCGTT
CTTCTAGTCC TATACACCGC CTACTCGCCG TAAAGGCAC TGCAGAGCAA

2101 GCTGCATAAA CCGACTACAC AAATCAGCGA TTTCCATGTT GCCACTCGCT
CGACGTATTT GGCTGATGTG TTTAGTCGCT AAAGGTACAA CGGTGAGCGA

2151 TTAATGATGA TTTCAGCCGC GCTGTACTGG AGGCTGAAGT TCAGATGTGC
AATTACTACT AAAGTCGGCG CGACATGACC TCCGACTTCA AGTCTACAG

2201 GCGGAGTTGC GTGACTACCT ACGGGTAACA GTTCTTTTAT GGCAGGGTGA
CCGCTCAACG CACTGATGGA TGCCCATGT CAAAGAAATA CCGTCCCACT

2251 AACGCAGGTC GCCAGCGGCA CCGCGCCTTT CGGCGGTGAA ATTATCGATG
TTGGCTCCAG CGGTGCGCGT GCGCGGAAA GCCGCCACTT TAATAGCTAC

2301 AGCGTGGTGG TTATGCCGAT CGCGTCACAC TACGTCTGAA CGTCGAAAAC
TCGCACCACC AATACGGCTA GCGCAGTGTG ATGCAGACTT GCAGCTTTTG

2351 CCGAACTGT GGAGCGCCGA AATCCCGAAT CTCTATCGTG CGGTGGTTGA
GGCTTTGACA CCTCGCGGCT TTAGGGCTTA GAGATAGCAC GCCACCACT

2401 ACTGCACACC GCCGACGGCA CGCTGATTGA AGCAGAAGCC TCGATGTGCG
TGACGTGTGG CGGCTGCCGT GCGACTAACT TCGTCTTCGG ACGCTACAGC

2451 GTTTCGCGCA GGTGCGGATT GAAAAAGGTC TGCTGCTGCT GAACGGCAAG
CAAAGGCGCT CCACGCCTAA CTTTACCAG ACGACGACGA CTGCGCGTTC

2501 CCGTTGCTGA TTCGAGGCGT TAACCGTCAC GAGCATCATC CTCTGCATGG
GGCAACCACT AAGCTCCGCA ATTGGCAGTG CTCGTAGTAG GAGACGTACC

2551 TCAGGTCATG GATGAGCAGA CGATGGTGCA GGATATCCTG CTGATGAAGC
AGTCCAGTAC CTACTCGTCT GCTACCAGCT CCTATAGGAC GACTACTTCG

2601 AGAACAACTT TAACGCCGTG CGCTGTTGCG ATTATCCGAA CCATCCGCTG
TCTTGTGAA ATTGCGGCAC GCGACAAGCG TAATAGGCTT GGTAGGCGAC

2651 TGGTACACGC TGTGCGACCG CTACGGCCTG TATGTGGTGG ATGAAGCCAA
ACCATGTGCG ACACGCTGGC GATGCCGGAC ATACACCACC TACTTCGGTT

2701 TATTGAAACC CACGGCATGG TGCCAATGAA TCGTCTGACC GATGATCOGC
ATAACTTTGG GTGCCGTACC ACGGTTACTT AGCAGACTGG CTACTAGGCG

2751 GCTGGCTACC GCGGATGAGC GAACGCGTAA CGCGAATGGT GCAGCGCGAT
CGACCGATGG CCGCTACTCG CTGCGCATT GCGCTTACCA CGTCGCGCTA

2801 CGTAATCACC CGAGTGTGAT CATCTGGTGG CTGGGGAATG AATCAGGCCA
GCATTAGTGG GCTCACACTA GTAGACCAGC GACCCCTTAC TTAGTGGCGT

2851 CGGCGCTAAT CACGACGCGC TGTATCGCTG GATCAAATCT GTCGATCCTT
GCCGCGATTA GTGCTGCGCG ACATAGCGAC CTAGTTTASA CAGCTAGGAA

2901 CCCGCCCGGT GCAGTATGAA GCGCGCGGAG CCGACACCAC GGCACCGAT
GGGCGGGCCA CGTCATACCT CCGCCGCCCTC GGCTGTGGTG CCGGTGGCTA

2951 ATTATTGGCC CGATGTACGC GCGCGTGGAT GAAGACCAGC CCTTCCCGGC
TAATAAACGG GCTACATGCG CCGCACCTA CTCTGGTGG GGAAGGGCCG

3001 TGTGCCGAAA TGGTCCATCA AAAAATGGCT TTCGCTACCT GGAGAGACGC
ACACGGCTTT ACCAGGTAGT TTTTACC GAAGCATGGA CCTCTGCG

3051 GCCCGCTGAT CCTTTGCGAA TACGCCACG CGATGGGTAA CAGTCTTGGC
CGGCGGACTA GGAACGCTT ATGCGGGTGC GCTACCCATT GTCAGAACCG

3101 GGTTCGCTA AATACTGGCA GCGGTTTCGT CAGTATCCCC GTTACAGGG
CCAAAGCGAT TTATGACCGT CCGCAAAGCA GTCATAGGGG CAATGTCC

3151 CGGCTTCGTC TGGGACTGGG TGGATCAGTC GCTGATTAAA TATGATGAAA
GCCGAAGCAG ACCCTGACCC ACCTAGTCAG CGACTAATT ATACTACTTT

3201 ACGGCACCCC GTGGTCGGCT TACGGCGGTG ATTTGGGGA TACGCCGAAC
TGCCGTTGGG CACCAGCCGA ATGCCGCCAC TAAACCGCT ATGCGGCTTG

3251 GATCGCCAGT TCTGTATGAA CGGTCTGGTC TTGCGGACC GCACGCGCA
CTAGCGGTCA AGACATACCT GCCAGACCAG AACGGGCTGG CGTGGCGCT

3301 TCCAGCGCTG ACGGAAGCAA AACACCAGCA GCAGTTTTC CAGTTCGGTT
AGGTGCGGAC TGCCCTCGTT TTGTGGTCGT CGTCAAAAG GTCAGGCAA

3351 TATCCGGGCA AACCATCGAA GTGAOCAGCG AATACCTGTT CCGTCATAGC
ATAGGCCCGT TTGGTAGCTT CACTGGTCGC TTATGGACAA GGCAGTATCG

3401 GATAACGAGC TCCTGCACTG GATGGTGGCG CTGGATGGTA AGCCGCTGGC
CTATTGCTCG AGGACGTGAC CTACCACCG GACCTACCT TCGGCGACCG

3451 AAGCGGTGAA GTGCCCTCTGG ATGTGCTCC ACAAGGTAAA CAGTTGATTG
TTGCCCACTT CACGGAGACC TACAGCGAGG TGTTCCATT GTCAACTAAC

3501 AACTGCCTGA ACTACCGCAG CCGGAGAGCG CCGGGCAACT CTGGCTCACA
TTGACGGACT TGATGGCGTC GGCCTCTCG GGCCTGTA GACCGAGTGT

3551 GTACGCGTAG TGCAACCGAA CCGACCGCA TGGTCAGAG CCGGGCACAT
CATGCGCATC ACCTTGGCTT GCGCTGGCGT ACCAGTCTT GGCCTGTA

3601 CAGCGCCTGG CAGCAGTGGC GTCTGGCGGA AACCTCAGT GTGACGCTCC
GTGCGGACC GTCGTACCG CAGACCGCT TTGGAGTCA CACTGCGAGG

3651 CCGCCGCGTC CCACGCCATC CCGCATCTGA CCACCAGOGA AATGGATTTT
GGCGGCGCAG GGTGCGGTAG GCGTAGACT GGTGGTGGT TTACCTAAA

3701 TGCACTGAGC TGGTAATAA GCGTTGGCAA TTTAACCGCC AGTCAGGCTT
ACGTAGCTCG ACCCATATT CGCAACCGTT AATTGGGG TCACTCCGAA

3751 TCTTTCACAG ATGTGGATTG GCGATAAAAA ACAACTGCTG ACGCCGCTGC
AGAAAGTGC TACACCTAAC CGCTATTTT TGTGACGAC TCGGCGGACG

3801 GCGATCAGTT CACCCGTGTC GATAGATCTG AACAGAACT CATTTCCGAA
CGCTAGTCAA GTGGGCACAG CTATCTAGAC TTGTCTTTGA GTAAAGGCTT

3851 GAAGACCTAG TCGACCATCA TCATCATCAT CACCGGTAAT AATAGGTAGA
CTTCTGGATC AGCTGGTAGT AGTAGTAGTA GTGGOCATTA TTATCCATCT

3901 TAAGTGACTG ATTAGATGCA TTTCGACTAG ATCCCTCGAC CAATTCCGGT
ATTCACCTGAC TAATCTACGT AAAGCTGATC TAGGGAGCTG GTTAAGGCCA

3951 TATTTTCCAC CATATTGCCG TCTTTGGCA ATGTGAGGGC CCGGAAACCT
ATAAAAGGTG GTATAACGGC AGAAAACCGT TACACTCCCG GGCCTTTGGA

4001 GGCCTGTCT TCTTGACGAG CATTCTAGG GGTCTTTCCC CTCTCGCCAA
CCGGGACAGA AGAACTGCTC GTAAGGATCC CCAGAAAGGG GAGAGCGGT

4051 AGGAATGCAA GGTCTGTTGA ATGTCGTGAA GGAAGCAGTT CCTCTGGAAG
TCCTTACGTT CCAGACAAC TACAGCACTT CCTTCGTCAA GGAGACCTTC

4101 CTTCTTGAAG ACAACAACG TCTGTAGCGA CCCTTTGCAG GCAGCGGAAC
GAAGAACTTC TGTTGTTGC AGACATCGCT GGGAAACGTC CGTCGCCTTG

4151 CCCCCACCTG GCGACAGGTG CCTCTGCGGC CAAAAGCCAC GTGTATAAGA
GGGGGTGGAC CGCTGTCCAC GGAGACGCCG GTTTTCGGTG CACATATTCT

4201 TACACCTGCA AAGCGGCAC AACCCAGTG CCACGTTGTG AGTTGGATAG
ATGTGGACGT TTCCGCCGTG TTGGGGTCAC GGTGCAACAC TCAACCTATC

4251 TGTGGAAAG AGTCAAATGG CTCCTCTCAA GCGTATTCAA CAAGGGGCTG
AACACCTTTC TCAGTTTACC GAGAGGAGTT GGCATAAGTT GTTCCCCGAC

4301 AAGGATGCCC AGAAGGTACC CCATTGTATG GGATCTGATC TGGGGCCTCG
TTCTACGGG TCTTCCATGG GGTAACATAC CCTAGACTAG ACCCCGGAGC

4351 GTGCACATGC TTTACATGTG TTTAGTCGAG GTTAAAAAC GTCTAGGCCC
CAGCTGTACG AAATGTACAC AAATCAGCTC CAATTTTTTG CAGATCCGGG

4401 CCCGAACAC GGGGACGTGG TTTTCCTTTG AAAAACACGA TGATAATACC
GGGCTTGGTG CCCCTGCACC AAAAGGAAAC TTTTGTGCT ACTATTATGG

4451 ATGAAAAAGC CTGAACACAC CGCGACGTCT GTCGAGAAGT TTCTGATCGA
TACTTTTTTC GACTTGAGTG GCGCTGCAGA CAGCTCTTCA AAGACTAGCT

4501 AAAGTTCGAC AGCGTCTCCG ACCTGATGCA GCTCTCGAG GCGGAAGAAT
TTTCAAGCTG TCGCAGAGGC TGGACTACGT CGAGAGCCTC CCGCTTCTTA

4551 CTCGTGCTTT CAGCTTCGAT GTAGGAGGGC GTGGATATGT CCTGCGGGTA
GAGCACGAAA GTCGAAGCTA CATCCTCCCG CACCTATACA GGACGCCCAT

4601 AATAGCTGCG CCGATGGTTT CTACAAAGAT CGTTATGTTT ATCGGCACTT
TTATCGACGC GGCTACCAA GATGTTTCTA GCAATACAAA TAGCCGTGAA

4651 TGCATCGGCC GCGCTCCCGA TTCGGAAGT GCTTGACRTT GGGGAATTTA
ACGTAGCCGG CCGAGGGGCT AAGGCCCTCA CGAACTGTAA CCCCTTAAAT

4701 GCGAGAGCCT GACCTATTGC ATCTCCGCC GTGCACAGGG TGTCAGTTG
CGCTCTCGGA CTGGATAACG TAGAGGGCGG CAGGTGTCCT ACAGTGCAAC

4751 CAAGACCTGC CTGAAACCGA ACTGCCCCGCT GTTCTGCAGC CGGTGCGGGA
GTTCTGGACG GACTTTGGCT TGACGGGCGA CAAGACGTCG GCCAGCGCCT

4801 GGCCATGGAT GCGATCGCTG CGGCCGATCT TAGCCAGACG AGCGGGTTCG
CCGGTACCTA CGCTAGCGAC GCCGGCTAGA ATCGGTCTGC TCGCCCAAGC

4851 GCCCATTCGG ACCGCAAGGA ATCGGTCAAT ACACTACATG GCGTGATTTC
CGGGTAAGCC TGGCGTTCCT TAGCCAGTTA TGTGATGTAC CGCACTAAAG

4901 ATATGCGCGA TTGCTGATCC CCATGTGTAT CACTGGCAAA CTGTGATGGA
TATACGCGCT AACGACTAGG GGTACACATA GTGACCGTTT GACACTACCT

4951 CGACACCGTC AGTGCGTCCG TCGCGCAGGC TCTCGATGAG CTGATGCTTT
GCTGTGGCAG TCACGCAGGC AGCGCGTCCG AGAGCTACTC GACTACGAAA

5001 GGGCCGAGGA CTGCCCCGAA GTCCGGCACC TCGTGACGCG GGATTTCGGC
CCCGGCTCCT GACGGGGCTT CAGGCGGTGG AGCACGTGCG CCTAAAGCCG

5051 TCCAACAATG TCCTGACGGA CAATGGCCGC ATAACAGCGG TCATTGACTG
AGGTTGTTAC AGGACTGCCT GTTACCGGCG TATTGTGCGC AGTAACTGAC

5101 GAGCGAGGCG ATGTTGCGGG ATTCGCAATA CGAGGTCGCG AACATCTTCT
CTCGCTCCGC TACAAGCCCC TAAGGGTTAT GCTCCAGCGG TTGTAGAAGA

5151 TCTGGAGGCC GTGGTTGGCT TGTATGGAGC AGCAGACGCG CTACTTCGAG
AGACCTCCGG CACCAACCGA ACATACCTCG TCGTCTGCGC GATGAAGCTC

5201 CGGAGGCATC CGGAGCTTGC AGGATCGCCG CGGCTCCGGG CGTATATGCT
GCCTCCGTAG GCCTCGAAGC TCCTAGCGGC GCCGAGGCC GCATATACGA

5251 CCGCATTTGGT CTTGACCAAC TCTATCAGAG CTTGGTTGAC GGCAATTTGG
GGCGTAACCA GAATGCTTG AGATAGTCTC GAACCAACTG CCGTTAAAGC

5301 ATGATGCAGC TTGGGCGCAG GGTGATGCG ACGCAATCGT CCGATCCGGA
TACTACGTGC AACCCGCGTC CCAGCTACGC TCGGTTAGCA GGCTAGGCCT

5351 GCCGGGACTG TCGGGCGTAC ACAAATCGCC CGCAGAGCG CGGCCGTCTG
CGGCCCTGAC AGCCCGCATG TGTTTAGCGG GCGTCTTCGC GCCGGCAGAC

5401 GACCGATGGC TGTGTAGAAG TACTCGCCGA TAGTGGAAAC CGACGCCCA
CTGGCTACCG ACACATCTTC ATGAGCGGCT ATCACCTTTG GCTCGGGGCT

5451 GCACTCGTCC GAGGGCAAG GAATAGAGTA GATGCCGACC GGGATCTATC
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5501 GATAAAATAA AAGATTTTAT TTAGTCTCCA GAAAAAGGGG GGAATGAAG
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5551 ACCCCACCTG TAGGTTTGGC AAGCTAGCTT AAGTAACGCC ATTTTGCAAG
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5601 GCATGGAAAA ATACATAACT GAGATAGAG AAGTTCAGAT CAAGGTCAGG
CGTACCTTTT TATGTATTGA CTCTTATCTC TTCAAGTCTA GTTCCAGTCC

5651 AACAGATGGA ACAGCTGAAT ATGGGCCAAA CAGGATATCT GTGGTAAGCA
TTGTCTACCT TGTGACTTA TACCCGGTTT GTCCTATAGA CACCATTCGT

5701 GTTCCTGCCC CGGCTCAGGG CCAAGAACAG ATGGAACACC TGAATATGGG
CAAGGACGGG GCCGAGTCCC GGTCTTTGTC TACCTTGTCG ACTTATACCC

5751 CCAAACAGGA TATCTGTGGT AAGCAGTTCC TGCCCCGGCT CAGGGCCAAG
GGTTTGTCTT ATAGACACCA TTCGTCAAGG ACGGGGCCGA GTCCCGGTTT

5801 AACAGATGGT CCCCAGATGC GGTCCAGCCC TCAGCAGTTT CTAGAGAACC
TTGTCTACCA GGGGTCTACG CCAGGTCCGG AGTCGTCAA GATCTCTTGG

5851 ATCAGATGTT TCCAGGGTGC CCCAAGGACC TGAATGACC CTGTGCCTTA
TAGTCTACAA AGGTCCACG GGGTTCCTGG ACTTTACTGG GACACCGAAT

5901 TTTGAACTAA CCAATCAGTT CGCTTCTCGC TTCTGTTCGC GCGCTTCTGC
AAACTTGATT GGTTAGTCAA GCGAAGAGCG AAGACAAGCG CGCGAAGACG

5951 TCCCCGAGCT CAATAAAGA GCCCACAACC CCTCACTCGG GCGGCCAGTC
AGGGGCTCGA GTTATTTTCT CCGGTGTTGG GGAGTGAGCC CCGCGGTGAG

6001 CTCCGATTGA CTGAGTCGCC CGGGTACCCG TGTATCCAAT AAACCCCTTT
GAGGCTAACT GACTCAGCGG GCCCATGGGC ACATAGGTTA TTTGGGAGAA

6051 GCAGTTGCAT CCGACTTGTG GTCTCGCTGT TCCTTGGGAG GGTCTCCTCT
CGTCAACGTA GGCTGAACRC CAGAGCGACA AGGAACCCCTC CCAGAGGAGA

6101 GAGTGATTGA CTACCCGTCA GCGGGGGTCT TTCATTCTAG CAGCATGTAT
CTCACTAACT GATGGGCAGT CGCCCCAGA AAGTAAGTAC GTCGTACATA

6151 CAAAATTAA TTTGGTTTTT TTCCTAAGTA TTTACATTAA ATGGCCATAG
GTTTTAATTA AACCAAAAAA AAGAATTCAT AAATGTAATT TACCGGTATC

6201 TTGCATTAAAT GAATCGGCCA ACGCGCGGGG AGAGCGGCTT TGCCTATTGG
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6251 CGCTCTTCCG CTTCTCTGCT CACTGACTCG CTGCGCTCGG TCGTTCGGCT
GCGAGAAGGC GAAGGAGCGA GTGACTGAGC GACGCGAGCC AGCAAGCCGA

6301 GCGGCGAGCG GTATCAGCTC ACTCAAAGGC GGTAAATACG TTATCCACAG
CGCCGCTCGC CATAGTCGAG TGAGTTTCCG CCATTATGCC AATAGGTGTC

6351 AATCAGGGGA TAACGCAGGA AAGAACATGT GAGCAAAAGG CCAGCAAAAG
TTAGTCCCTT ATTGCGTCTT TTCTTGATCA CTCGTTTTCC GGTGTTTTTC

6401 GCCAGGAACC GTAAAAAGGC CGCGTTGCTG GCGTTTTTCC ATAGGCTCCG
CGGTCTTGG CATTTTTCCG GCGCAACGAC CGCAAAAAGG TATCCGAGGC

6451 CCCCCCTGAC GAGCATCACA AAAATCGACG CTCAAGTCAG AGGTGGCGAA
GGGGGGACTG CTCGTAGTGT TTTAGCTGC GAGTTCAGTC TCCACCGCTT

6501 ACCCGACAGS ACTATAAAGA TACCAGGCGT TTCCCCCTGG AAGCTCCCTC
TGGGCTGTCC TGATATTTCT ATGGTCCGCA AAGGGGGACC TTCGAGGGAG

6551 GTGCGCTCTC CTGTTCCGAC CTTGCCGCTT ACCGGATACC TGTCCGCTT
CACGCGAGAG GACAAGGCTG GAGCGGCGAA TGGCCTATGG ACAGGCGGAA

6601 TCTCCCTTCG GGAAGCGTGG CGCTTTCTCA TAGCTCACGC TGTAGGTATC
AGAGGGAAGC CCTTCGCACC GCGAAAGAGT ATCGAGTCGC ACATCCATAG

5651 TCAGTTCGGT GTAGGTCGTT CGCTCCAAGC TGGGCTGIGT GCACGAACCC
AGTCAAGCCA CATCCAGCAA GCGAGGTTCC ACCCGACACA CGTGCTTGGG

5701 CCCGTTTCAGC CCGACCGCTG CGCCTTATCC GGTAACTATC GTCTTGACTC
GGGCAAGTCG GGCTGGCGAC GCGGAATAGG CCATTGATAG CAGAACTCAG

5751 CAACCCGGTA AGACACGACT TATCGCCACT GGCAGCAGCC ACTGGTAACA
GTTGGGCCAT TCTGTGCTGA ATAGCGGTGA CCGTCGTCCG TGACCATGT

5801 GGATTAGCAG AGCGAGGTAT GTAGGCGGTG CTACAGAGTT CTTGAAGTGG
CCTAATCGTC TCGCTCCATA CATCCGCCAC GATGTCTCAA GAACTTCACC

5851 TGGCCTAACT ACGGCTACAC TAGAAGAACA GTATTTGGTA TCTGCGCTCT
ACCGGATTGA TGCCGATGTG ATCTTCTTGT CATAAACCAT AGACGCGAGA

5901 GCTGAAGCCA GTTACCTTCG GAAAAAGAGT TGGTAGCTCT TGATCCGGCA
CGACTTCGGT CAATGGAAGC CTTTTCTCA ACCATCGAGA ACTAGGCCGT

5951 AACAAACCAC CGCTGSTAGC GGTGGTTTTT TTGTTTGCAA GCACGAGATT
TTGTTTGGTG GCGACCATCG CCACCAAAAA AACAAACGTT CGTCGTCTAA

7001 ACGCGCAGAA AAAAAGGATC TCAAGAGAT CCTTGATCT TTTCTACGGG
TGCGCGTCTT TTTTCCTAG AGTTCCTCTA GGAACTAGA AAAGATGCCC

7051 GTCTGACGCT CAGTGAACG AAACTCAGC TTAAGGGATT TTGGTCATGA
CAGACTGCGA GTCACCTTGC TTTTGAGTGC AATTCCTAA AACCACTACT

7101 GATTATCAAA AAGGATCTTC ACCTAGATCC TTTTAAATTA AAAATGAAGT
CTAATAGTTT TTCCTAGAAG TGGATCTAGG AAAATTTAAT TTTTACTTCA

7151 TTGCGGCCGC AAATCAATCT AAAGTATATA TGAGTAACT TGGTCTGACA
AACGCCGCGC TTTAGTTAGA TTTATATAT ACTCATTGA ACCAGACTGT

7201 GTTACCAATG CTTAATCAGT GAGGCACCTA TCTCAGCGAT CTGTCTATTT
CAATGGTTAC GAATTAGTCA CTCCGTGGAT AGAGTCGCTA GACAGATAAA

7251 CGTTTCATCCA TAGTTGCCCTG ACTCCCGCTC GTGTAGATAA CTACGATACG
GCAAGTAGGT ATCAACGGAC TGAGGGGCGAG CACATCTATT GATGCTATGC

7301 GGAGGGCTTA CCATCTGGCC CCACTGCTGC AATGATACCG CGAGACCCAC
CCTCCCGAAT GGTAGACCGG GGTACGACG TTAATATGGC GCTCTGGGTG

7351 GCTCACCGGC TCCAGATTTA TCAGCAATAA ACCAGCCAGC CGGAAGGGCC
CGAGTGGCCG AGGTCTAAAT AGTCGTTATT TGGTCGGTCG GCCTTCCCGG

7401 GAGCGCAGAA GTGGTCCTGC AACTTTATCC GCGTCCATCC AGTCTATTAA
CTCGCGTCTT CACCAGGACG TTGAAATAGG CGGAGGTAGG TCAGATAATT

7451 TTGTTGCGCG GAGCTAGAG TAAGTAGTTC GCCAGTTAAT AGTTTGCGCA
AACACGGCC CTTGATCTC ATTCATCAAG CGGTCAATTA TCAAACGCGT

7501 ACGTTGTTGC CATTGCTACA GGCATCGTGG TGTACGCTC GTCGTTTGGT
TGCAACAACG GTAACGATGT CCGTAGCACC ACAGTCCGAG CAGCAAACCA

7551 ATGGCTTCAT TCAGCTCCGG TTCOCAACGA TCAAGGCGAG TTACATGATC
TACCGAAGTA AGTCGAGGCC AAGGTTGCT AGTTCGCTC AATGTACTAG

7601 CCCCATGTTG TGCAAAAAG CGGTTAGCTC CTTGGTCCT CCGATCGTTG
GGGGTACAAC ACGTTTTTTC GCCAATCGAG GAAGCCAGGA GGCTAGCAAC

7651 TCAGAAGTAA GTTGGCCGCA GTGTTATCAC TCATGGTTAT GGCAGCACTG
AGTCTTCATT CAACCGGCGT CACAATAGTG AGTACCAATA CCGTCGTGAC

7701 CATRAATCTC TTAAGTTCAT GCCATCCGTA AGATGCTTTT CTGTGACTGG
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7751 TGAGTACTCA ACCAAGTCAT TCTGAGAATA GTGTATGCCG CGACCGAGTT
ACTCATGAGT TGGTTCAGTA AGACTCTTAT CACATACGCC GCTGGCTCAA

7801 GCTCTTGCCC GCGGTCAATA CCGGATAATA CCGCGCCACA TAGCAGAACT
CGAGAACGGG CCGCAGTTAT GCCCTATTAT GCGCGGGTGT ATCGTCTTGA

7851 TTAAGAGTGC TCATCATTGG AAAACGTTCT TCGGGGCGAA AACTCTCAAG
AATTTTCACG AGTAGTAACC TTTTGCAAGA AGCCCCGCTT TTGAGAGTTC

7901 GATCTTACCG CTGTTGAGAT CCAGTTCGAT GTAACCCACT CGTGCACCCA
CTAGAATGGC GACAACCTA GGTCAAGCTA CATTGGGTGA GCACGTGGGT

7951 ACTGATCTTC AGCATCTTTT ACTTCAOCA GCGTTTCTGG GTGAGCAAAA
TGACTAGAAG TCGTAGAAAA TGAAAGTGGT CGCAAAGACC CACTCGTTTT

8001 ACAGGAAGGC AAAATGCCGC AAAAAGGGA ATAAGGGCGA CACGGAATG
TGTCTTCCG TTTTACGGCG TTTTTCCT TATTCCCGCT GTGCCTTAC

8051 TTGAATACTC ATACTCTTC TTTTCAATA TTATTGAAGC ATTTATCAGG
AACTTATGAG TATGAGAAGG AAAAGTTAT AATBACTTCG TAAATAGTCC

8101 GTTATTGTCT CATGAGCGGA TACATATTG AATGTATTTA GAAAAATAA
CAATAACAGA GTACTCGCCT ATGTATAAAC TTACATAAAI CTTTTATTT

8151 CAAATAGGGG TTCCGCGCAC ATTTT
GTTTATCCCC AAGGCGCGTG TAAAG

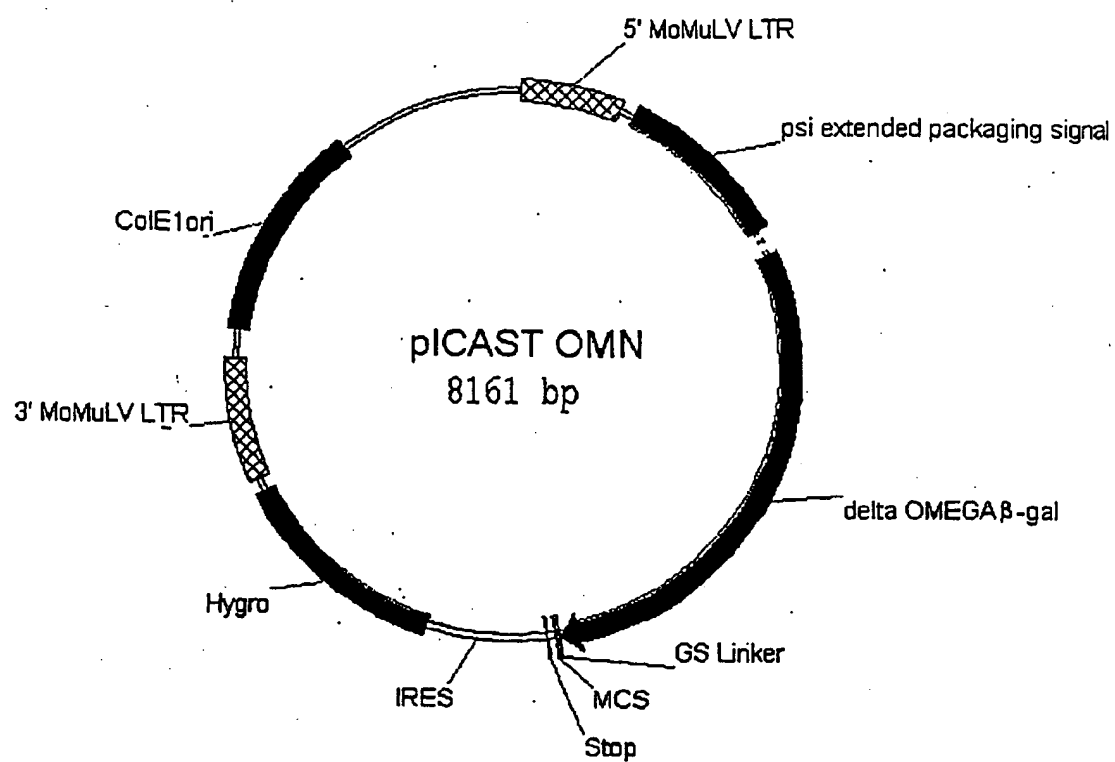


Figure 13A

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1  CTGCAGCCTG AATATGGGCC AAACAGGATA TCTGTGGTAA GCAGTTCCTG
   GACGTCGGAC TTATACCCGG TTTGTCTAT AGACACCATT CGTCAAGGAC
-----
51  CCCC GGCTCA GGGCCAAGAA CAGATGGAAC AGCTGAATAT GGGCCAAACA
   GGGGCCGAGT CCCGGTTCTT GTCTACCTTG TCGACTTATA CCCGGTTTGT
-----
101 GGATATCTGT GGTAAAGCAGT TCCTGCCCCG GCTCAGGGCC AAGAACAGAT
   CCTATAGACA CCATTCTGTA AGGACGGGGC CGAGTCCCGG TTCTTGTCTA
-----
151 GGTCCCCAGA TGC GGTCAG CCCTCAGCAG TTTCTAGAGA ACCATCAGAT
   CCAGGGGTCT ACGCCAGGTC GGGAGTCGTC AAAGATCTCT TGGTAGTCTA
-----
201 GTTTCCAGGG TGCCCAAGG ACCTGAAATG ACCCTGTGCC TTATTGAAC
   CAAAGGTCCC ACGGGGTTC TGGACTTTAC TGGGACACGG AATAAACTTG
-----
251 TAACCAATCA GTTCGCTTCT CGCTTCTGTT CGCGCGCTTC TGCTCCCGA
   ATTGGTTAGT CAAGCGAAGA GCGAAGACAA GCGCGCGAAG ACGAGGGGCT
-----
301 GCTCAATAAA AGAGCCCAACA ACCCTCACT CGGGGCGCCA GTCCTCCGAT
   CGAGTTATTT TCTCGGGTGT TGGGGAGTGA GCCCGCGGGT CAGGAGGCTA
-----
351 TGACTGAGTC GCCCGGGTAC CCGTGTATCC AATAAACCTT CTTCAGTTG
   ACTGACTCAG CCGGCCCATG GGCACATAGG TTATTTGGGA GAACGTCAAC
-----
401 CATCGACTT GTGGTCTCGC TGTTCTTGG GAGGGTCTCC TCTGAGTGAT
   GTAGGCTGAA CACCAGAGCG ACAAGGAACC CTCCAGAGG AGACTCACTA
-----
451 TGACTACCG TCAGCGGGGG TCTTTCATT GGGGGCTCGT CCGGGATCGG
   ACTGATGGGC AGTCGCCCC AGAAAGTAA CCCCCGAGCA GGCCCTAGCC
-----
501 GAGACCCCTG CCCAGGGACC ACCGACCAC CACCGGGAGG CAAGCTGGCC
   CTCTGGGGAC GGGTCCCTGG TGGCTGGGTG GTGGCCCTCC GTTCGACCGG
-----
551 AGCAACTTAT CTGTGTCTGT CCGATTGTCT AGTGTCTATG ACTGATTTTA
   TCGTTGAATA GACACAGACA GGCTAACAGA TCACAGATAC TGACTAAAAT
-----
601 TGCGCTGCG TC GGTA TAGCTA ACT AGCTCTGTAT CTGGCGGACC
   ACGCGGACGC AGCCATGATC AATCGATTGA TCGAGACATA GACCGCCTGG
-----
651 CGTGGTGGAA CTGACGAGTT CTGAACACCC GGCGCAACC CTGGGAGACG
   GCACCACTT GACTGCTCAA GACTTGTGGG CCGCGGTTGG GACCCTCTGC
-----
701 TCCAGGGAC TTTGGGGGCC GTTTTGTGG CCGACCTGA GGAAGGGAGT
   AGGGTCCCTG AAACCCCGG CAAAACACC GGGCTGGACT CCTTCCCTCA
-----
751 CGATGTGGAA TCCGACCCG TCAGGATATG TGGTCTGGT AGGAGACGAG
   GCTACACCTT AGGCTGGGGC AGTCCTATAC ACCAAGACCA TCCTCTGCTC
-----
801 AACCTAAAAC AGTCCCGCC TCCGTCTGAA TTTTGTCTT CGGTTTGGAA
   TTGGATTTT TCAAGGGCGG AGGCAGACTT AAAACGAAA GCCAAACCTT
-----
851 CCGAAGCCGC CGTCTTGTG TGCTGCAGCA TCGTTCTGTG TTGTCTCTGT
   GGCTTCGGCG CGCAGAACAG ACGACGTCGT AGCAAGACAC AACAGAGACA
-----
901 CTGACTGTGT TTCTGTATTT GTCTGAAAAT TAGGGCCAGA CTGTTACCAC
   GACTGACACA AAGACATAAA CAGACTTTTA ATCCCGGTCT GACAATGGTG
-----

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FIGURE 13B

951 TCCCTTAAGT TTGACCTTAG. GTAACCTGGAA AGATGTGAG CGGCTCGCTC
AGGGAATTCA AACTGGAATC CATTGACCTT TCTACAGCTC GCCGAGCGAG

1001 ACAACCAGTC GGTAGATGTC AAGAAGAGAC GTTGGGTTAC CTTCTGCTCT
TGTTGGTCAG CCATCTACAG TTCTTCTCTG CAACCCAATG GAAGACGAGA

1051 GCAGARTGGC CAACCTTTAA CGTCGGATGG CCGCGAGACG GCACCTTTAA
CGTCTTAGCG GTTGGAAATG GCAGCCTACC GCGGCTCTGC CGTGGAAAT

1101 CCGAGACCTC ATCACCAGG TTAAGATCAA GGTCTTTTCA CCTGGCCCGC
GGCTCTGGAG TAGTGGGTCC AATTCTAGTT CCAGAAAAGT GGACCGGGCG

1151 ATGGACACCC AGACCAGGTC CCCTACATCG TGACCTGGGA AGCCTTGGCT
TACCTGTGGG TCTGGTCCAG GGGATGTAGC ACTGGACCCT TCGGAACCGA

1201 TTTGACCCCC CTCCCTGGGT CAGGCCCTT GTACACCCTA AGCCTCCGCC
AAACTGGGGG GAGGGACCCA GTTCGGGAAA CATGTGGGAT TCGGAGGCGG

1251 TCCTCTTCCT CCATCCGCCG CGTCTCTCCC CCTTGAACCT CCTCGTTCCA
AGGAGAAGGA GGTAGGCGGG GCAGAGAGGG GGAACCTGGA GGAGCAAGCT

1301 CCCCCTCTCG ATCCTCCCTT TATCCAGCCC TCACTCCTTC TCTAGGCGCC
GGGCGGAGC TAGGAGGGAA ATAGGTCCGG AGTGAGGAAG AGATCCGCGG

1351 GGCCGCTCTA GCCCATTAAT ACGACTCACT ATAGGGCGAT TCGAACACCA
CCGCGGAGAT CCGGTAATTA TGCTGAGTGA TATCCCGCTA AGCTTGTGGT

1401 TGCACCATCA TCATCATCAC GTCGACGAAC AGAACTCAT TTCCGAAGAA
ACGTGGTAGT AGTAGTAGTG CAGCTGCTTG TCTTTGAGTA AAGGCTTCTT

1451 GACCTACTCG AGATGGGCGT GATTACGAT TCACTGGCCG TCGTTTACA
CTGGATGAGC TCTACCCGCA CTAATGCCTA AGTGACCGGC AGCAAAATGT

1501 ACGTCGTGAC TGGGAAAACC CTGGCGTTAC CCAACTTAAT CGCCTTGCAG
TGCAGCACTG ACCCTTTTGG GACCGCAATG GGTGAAATTA GCGGAACGTC

1551 CACATCCCCC TTTCGCCAGC TGGCGTAATA GCGAAGAGGC CCGCACCGAT
GTGTAGGGGG AAAGCGGTCC ACCGCATTAT CGCTTCTCCG GCGGTGGCTA

1601 CGCCCTTCCC AACAGTTACG CAGCCTGAAT GCGGAATGGC GCTTTGCCTG
GCGGGAAGGG TTGTCAATGC GTCGGACTTA CCGCTTACCG CGAAACGGAC

1651 GTTTCGGGCA CCAGAAGCGG TGCCGGAAAG CTGGCTGGAG TCGATCTTC
CAAAGGCCGT GGTCTTCGCC ACGGCCTTTC GACCGACCTC ACGCTAGAAG

1701 CTGAGGCCGA TACTGTCGTC GTCCCTCAA ACTGGCAGAT GCACGGTTAC
GACTCCGGCT ATGACAGCAG CAGGGGAGTT TGACCGTCTA CGTGCCAATG

1751 GATGCGCCCA TCTACACCAA CGTGACCTAT CCCATTACGG TCAATCCGCC
CTACGCGGGT AGATGTGGTT GCACTGGATA GGGTAATGCC AGTTAGGCGG

1801 GTTGTGTTCC ACGGAGAATC CGACGGGTG TTA CTGCTC ACATTTAATG
CAACAAGGG TGCTCTTAG GCTGCCAAC AATGAGCGAG TGTAATTAC

1851 TTGATGAAG CTGGCTACAG GAAGGCCAGA CGCGAATTAT TTTGATGGC
AACTACTTTC GACCGATGTC CTTCCGGTCT GCGCTTAATA AAACTACCG

1901 GTTAACTCGG CGTTTCATCT GTGGTGCAAC GGGCGCTGGG TCGGTTACGG
CAATTGAGCC GCAAAGTAGA CACCACGTTG CCCGCGACCC AGCCAATGCC

1951 CCAGGACAGT CGTTTGCCGT CTGAATTTGA CCTGAGCGCA TTTTACGGG
GGTCCTGTCA GCAAACGGCA GACTTAACT GGACTCGCGT AAAAATGCGC

2001 CCGGAGAAAA CCGCCTCGCG GTGATGGTGC TGGCTGGAG TGACGGCAGT
GGCCTCTTTT GCGGAGCGC CACTACCAGC ACGCGACCTC ACTGCCGTCA

2051 TATCTGGAAG ATCAGGATAT GTGGCGGATG AGCGGCATTT TCCGTGACGT
ATAGACCTTC TAGTCCTATA CACCGCCTAC TCGCCGTAAA AGGCACTGCA

2101 CTCGTTGCTG CATAAACCGA CTACACAAAT CAGCGATTTT CATGTTGCCA
GAGCAAGCAC GTATTGGCT GATGTGTTA GTCGCTAAAG GTACAACGGT

2151 CTCGCTTTAA TGATGATTTC AGCCGCGCTG TACTGGAGGC TGAAGTTCAG
GAGCGAAATT ACTACTAAG TCGGCGCGAC ATGACCTCG ACTTCAAGTC

2201 ATGTGCGCGG AGTTGCGTGA CTACCTACGG GTAACAGTTT CTTTATGGCA
TACACGCCGC TCAACGCACT GATGGATGCC CATTGTCAA GAAATACCGT

2251 GGGTGAACG CAGGTCGCCA GCGGCACCGC GCCTTTCGGC GGTGAAATTA
CCCACTTTGC GTCCAGCGGT CGCCGTGGCG CGGAAGCCG CCACTTTAAT

2301 TCGATGAGCG TGGTGGTTAT GCCGATCGCG TCACACTAAG TCTGAACGTC
AGCTACTCGC ACCACCAATA CGGCTAGCGC AGTGTGATGC AGACTTGCAG

2351 GAAACCCCGA AACTGTGGAG CGCCGAAATC CCGAATCTCT ATCGTGCGGT
CTTTGGGCT TTGACACCTC CGCGCTTTAG GGCTTAGAGA TAGCACGCCA

2401 GGTGAACTG CACACCGCGC ACGGCACGCT GATTGAAGCA GAAGCCTCGG
CCAACCTTAC GTGTGGCGGC TGGCGTGCGA CTAACCTCGT CTTGGGACGC

2451 ATGTCGGTTT CCGCGAGGTG CGGATTGAAA ATGGTCTGCT GCTGCTGAAC
TACAGCCAAA GCGGCTCCAC GCCTAAGTTT TACCRAGCGA CGACGACTTG

2501 GGCAAGCCGT TGCTGATTCG AGCGGTTAAC CGTCACGAGC ATCATCCTCT
CCGTTGCGCA ACGACTAAGC TCCGCAATTG GCAGTGCTCG TAGTAGGAGA

2551 GCATGGTCAG GTCATGGATG AGCAGACGAT GGTGCAGGAT ATCCTGCTGA
CGTACCAGTC CAGTACCTAC TCGTCTGCTA CCACGTCCTA TAGGACGACT

2601 TGAAGCAGAA CAACTTTAAC GCCGTGCGCT GTTCGCATTA TCCGAACCAT
ACTTCGTCTT GTTGAAATTG CGGCACGCGA CAAGCGTAAT AGGCTTGCTA

2651 CCGCTGTGGT ACACGCTGTG CGACCGCTAC GGCCTGTATG TGGTGGATGA
GGCGACACCA TGTGCGACAC GCTGGCGATG CCGGACATAC ACCACCTACT

2701 AGCCAATATT GAAACCCACG GCATGGTGCC AATGAATCGT CTGACCGATG
TCGTTTATAA CTTTGGGTGC CGTACCACGG TACTTAGCA GACTGGCTAC

2751 ATCCGCGCTG GCTACCGGCG ATGAGCGAAC GCGTAACGCG AATGGTGCAG
TAGGCGCGAC CGATGGCCGC TACTCGCTTG CGCATTGCGC TTACCACGTC

2801 CGCGATCGTA ATCACCAGAG TGTGATCATC TGGTGGCTGG GGAATGAATC
CGGCTAGCAT TAGTGGGCTC AACTAGTAG ACCAGCGACC CTTACTTAG

2851 AGGCCACGGC GCTAATCAGC ACGCGGTGTA TCGCTGGATC AAATCTGTCTG
TCCGGTGCCG CGATTAGTGC TCGCGGACAT AGCGACCTAG TTTAGACAGC

2901 ATCCTTCCCG CCCGGTGCAG TATGAAGGCG GCGGAGCCGA CACCACGGCC
TAGGAAGGGC GGGCCACGTC ATACTTCCGC CGCCTCGGCT GTGGTGCCGG

2951 ACCGATATTA TTTGCCCGAT GTACGCGCGC GTGGATGAAG ACCAGCCCTT
TGGCTATAAT AAACGGGCTA CATGCGCGCG CACCTACTTC TGGTCGGGAA

3001 CCCGGCTGTG CCGAAATGGT CCATCAAAA ATGGCTTTCG CTACCTGGAG
GGGCCGACAC GGCTTTACCA GGTAGTTTTT TACCGAAAGC GATGGACCTC

3051 AGACGCGCCC GCTGATCCTT TGCGAATACG CCCACGCGAT GGGTAACAGT
TCTGCGCGGG CACTAGGAA ACGCTTATGC GGGTGCGCTA CCCATTGTCA

3101 CTTGGCGGTT TCGCTAAATA CTGGCAGGCG TTTGCTCAGT ATCCCCGTTT
GAACCGCCAA AGCGATTAT GACCGTCCGC AAAGCAGTCA TAGGGGCAAA

3151 ACAGGGCGGC TTCGTCTGGG ACTGGGTGGA TCAGTCGCTG ATTAAATATG
TGTCGCGCGG AAGCAGACCC TGACCCACCT AGTCAGCGAC TAATTTATAC

3201 ATGAAAACGG CAACCCGTGG TCGGCTTACG GCGGTGATT TGGCGATACG
TACTTTTGCC GTTGGGCACC AGCCGAATGC CGCCACTPAA ACCGCTATGC

3251 CCGAACGATC GCCAGTTCTG TATGAACGGT CTGGTCTTTC CCGACCGCAC
GGCTTGCTAG CGGTCAAGAC ATACTTGCCA GACCAGAAAC GGCTGGCGTG

3301 GCCGCATCCA GCGCTGACGG AAGCAAAACA CCAGCAGCAG TTTTCCAGT
CGGCGTAGGT CCGGACTGCC TTCGTTTGT GGTGCTGCTC AAAAAGGTCA

3351 TCCGTTTTATC CGGGCAAACC ATCGAAGTGA CCAGCGAATA CCTGTTCCGT
AGGCAATAG GCCCGTTTG TAGCTTCACT GGTGCTTAT GGACAAGGCA

3401 CATAGCGATA ACGAGCTCCT GCACTGGATG GTGGCGCTGG ATGTAAGCC
GTATCGCTAT TGCTCGAGGA CGTGACCTAC CACCGCGACC TACCATTGG

3451 GCTGGCAAGC GGTGAAGTGC CTCTGGATGT CGCTCCACAA GGTAACAGT
CGACCGTTTC CCACTTCACG GAGACCTACA GCGAGGTCTT CCATTGTCA

3501 TGATTGAACT GCCTGAACTA CCGCAGCCGG AGAGCGCCGG GCAACTCTGG
ACTAACTGA CGGACTTGAT GCGCTCGGCC TCTCGCGGCC CGTTGAGACC

3551 CTCACAGTAC GCGTAGTGCA ACGGAACGCG ACCGCATGGT CAGAAGCCGG
GAGTGTATG CGCATCACGT TGGCTTGCGC TGGCGTACCA GTCTTCGGCC

3601 GCACATCAGC GCCTGGCAGC AGTGGCGTCT GCGGGAACAC CTCAGTGTGA
CGTGTAGTCG CGGACCGTCG TCACCGCAGA CCGCCTTTTG GAGTCACACT

3651 CGCTCCCCCG CGGTCACCAC GCCATCCCCG ATCTGAACCAC CAGCGAAATG
GCGAGGGGCG GCGCAGGGTG CGGTAGGGCG TAGACTGGTG GTCGCTTTAC

3701 GATTTTTGCA TCGAGCTGGG TAATAAGCGT TGGCAATTA ACCGCCAGTC
CTAAAAAGCT AGCTCGACCC ATTATTCGCA ACCGTAAAT TGGCGGTGAC

3751 AGGCTTTCTT TCACAGATGT GGATTGGCGA TAAAAACAA CTGCTGACGC
TCCGAAAGAA AGTGTCTACA CCTAACCGCT ATTTTTGTT GACGACTGCG

3801 CGCTGCGCGA TCAGTTCACC CGTGTGATA GATCTGGAGG TGGTGGCAGC
GCGACGCGCT AGTCAAGTGG GCACAGCTAT CTAGACCTCC ACCACCGTCG

3851 AGGCCTTGGC GCGCCGGATC CTTAATTAAC AATTGACCGG TAATAATAGG
TCCGGAACCG CGCGGCCTAG GAATTAATTG TTAAGTGGCC ATTATTATCC

3901 TAGATAAGTG ACTGATTAGA TGCATTTCGA CTAGATCCCT CGACCAATTC
ATCTATTAC TGACTAATCT ACGTAAAGCT GATCTAGGGA GCTGGTTAAG

3951 CGGTTATTTT CCACCATATT GCGTCTTTT GGCAATGTGA GGGCCCGGAA
GCCAATAAAA GGTGGTATAA CGGCAGAAAA CCGTTACACT CCGGGGCTT

4001 ACCTGGCCCT GTCTTCTTGA CGAGCATTC TAGGGGTCTT TCCCCTCTCG
TGGACCGGGA CAGAGAAGT GCTCGTAAGG ATCCCAGAA AGGGGAGAGC

4051 CCAAGGAAT GCAAGGTCTG TTGAATGTG TGAAGGAAG AGTTCCTCTG
GGTTCTCTTA CGTCCAGAC AACTTACAGC ACTTCTCTG TCAAGGAGC

4101 GAAGCTTCTT GAAGACAAAC AACGTCTGTA GCGACCTTT GCAGGCAGCG
CTTCGAAGAA CTTCTGTTT TGCAGACAT CGCTGGGAAA CGTCCGTGCG

4151 GAACCCCCCA CCTGGCGACA GTGCTCTG CGGCCAAAG CCACGTGTAT
CTTGGGGGT GGACCGCTGT CCACGGAGC GCGCGTTTC GGTGCACATA

4201 AAGATACACC TGCAAGGGG GCACAACCC AGTGCCACGT TGTGAGTTGG
TTCTATGTGG ACGTTTCGCG CGTGTGGGG TCACGGTGCA AACTCAACC

4251 ATAGTTGTGG AAAGAGTCAA ATGGCTCTCC TCAAGCGTAT TCAACAAGGG
TATCAACACC TTTCTCAGT TACCGAGAGG AGTTGCGATA AGTTGTTOCC

4301 GCTGAAGGAT GCCCAGAAGG TACCCATTG TATGGGATCT GATCTGGGGC
CGACTTCTTA CCGGTCTTCC ATGGGGTAAC ATACCTAGA CTAGACCCG

4351 CTCGGTGCAC ATGCTTTACA TGTGTTTGT CGAGGTTAAA AAACGTCTAG
GAGCCACGTG TACGAAATGT ACACAAATCA GCTCAATTT TTTGCAGATC

4401 GCGCCCGGAA CCACGGGGAC GTGGTTTCC TTTGAAAAAC ACGATGATAA
CGGGGGGCTT GGTGCCCTG CACCAAAGG AAACTTTTT TGCTACTATT

4451 TACCATGAAA AAGCCTGAAC TCACCGCGAC GTCTGTCGAG AAGTTTCTGA
ATGGTACTTT TTCGGAAGT AGTGGCGCTG CAGACAGCTC TTCAAAGACT

4501 TCGAAAAGTT CGACAGGTC TCCGACCTGA TGCAGCTCTC GGAGGGCGAA
AGCTTTTCRA GCTGTGCGAG AGGCTGGACT ACGTCGAGAG CCTCCGCTT

4551 GAATCTCGTG CTTTCAGCTT CGATGTAGGA GGGCGTGGAT ATGCTCTGCG
CTTAGAGCAC GAAGTGGAA GCTACATCCT CCGCACCTA TACAGGACGC

4601 GGTAAATAGC TCGCCGATG GTTCTACAA AGATCGTTAT GTTATCGGC
CCATTTATCG ACGCGGCTAC CAAAGATGTT TCTAGCAATA CAAATAGCCG

4651 ACTTTGCATC GCGCGGCTC CCGATTCCGG AAGTGCTTGA CATTGGGGAA
TGAAACGTAG CCGCGCGAG GGCTAAGGCC TTCACGAAGT GTAACCCCTT

4701 TTAGCGAGA GCCTGACCTA TTGCATCTCC CGCCGTGCAC AGGGTGTAC
AAATCGCTCT CGGACTGGAT AACGTAGAGG GCGGCACTG TCCACAGTG

4751 GTTGCAAGAC CTGCCTGAAA CCGAACTGCC CGCTGTTCTG CAGCCGGTCG
CAACGTTCTG GACGGACTTT GGCTTGACGG GCGACAAGAC GTCGGCCAGC

4801 CGGAGGCCAT GGATGCGATC GCTGCGGCCG ATCTTAGCCA GACGAGCGGG
GCCTCCGGTA CCTACGCTAG CGAGCGCGGC TAGAATCGGT CTGCTCGCCC

4851 TTCGGCCCAT TCGGACCGCA AGGAATCGGT CAATACACTA CATGGCGTGA
AAGCGGGTA AGCCTGGCGT TCCTTAGCCA GTTATGTGAT GTACCGCACT

4901 TTTCATATGC GCGATTGCTG ATCCCATGT GTATCACTGG CAACTGTGA
AAAGTATACG CGCTAACGAC TAGGGGTACA CATAGTGACC GTTTGACACT

4951 TGGACGACAC CGTCAGTGCG TCCGTCGCGC AGGCTCTCGA TGAGCTGATG
ACCTGCTGTG GCAGTCACGC AGGCAGCGCG TCCGAGAGCT ACTCGACTAC

5001 CTTTGGGCGG AGGACTGCCC CGAAGTCCGG CACCTCGTGC ACGCGGATT
GAAACCCGGC TCCTGACGGG GCTTCAGGCC GTGGAGCAGG TCGGCCTAAA

5051 CGGCTCCAAC AATGTCCTGA CCGACAATGG CCGCATAACA GCGGTCAATT
GCCGAGGTTG TTACAGGACT GCCTGTTACC GCGTATTGT CGCCAGTAAC

5101 ACTGGAGCGA GCGCATGTTT GGGGATTCCC AATACGAGGT CGCCAACATC
TGACCTCGCT CCGCTACAAG CCCCTAAGGG TTATGCTCCA GCGGTTGTAG

5151 TTCTTCTGGA GGCCGTGGTT GGCTGTATG GAGCAGCAGA CGCGCTACTT
AAGAAGACCT CCGGCACCAA CCGAACATAC CTCGTCTGCT GCGCGATGAA

5201 CGAGCGGAGG CATCCGAGC TTGCAGGATC GCCGCGGCTC CGGGCGTATA
GCTCGCCTCC GTAGGCCTCG AACGTCCTAG CGGCGCGAG GCCCGCATAT

5251 TGCTCCGCAT TGGCTCTGAC CAACCTATC AGAGCTTGGT TGACGGCAAT
ACGAGGCGTA ACCAGAACTG GTTGAGATAG TCTCGAACCA ACTGCCGTTA

5301 TTCGATGATG CAGCTTGGGC GCAGGTCGA TGGACGCAA TCGTCCGATC
AAGCTACTAC GTCGAACCCG CGTOCCAGCT ACGCTGCGTT AGCAGGCTAG

5351 CGGAGCCGGG ACTGTCGGGC GTACACAAAT CGCCCGCAGA AGCGCGGCCG
GCCTCGGCC TGACAGCCCG CATGTGTTA GCGGGCGTCT TCGCGCGGGC

5401 TCTGGACCGA TGGCTGTGTA GAAGTACTCG CCGATAGTGG AAACCGACGC
AGACCTGGCT ACCGACACAT CTTCTAGAGC GGCTATCACC TTTGGCTGCG

5451 CCCAGCACTC GTCCGAGGGC AAAGGAATAG AGTAGATGCC GACCGGGATC
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5501 TATCGATAAA ATAAAGATT TTATTTAGTC TCCAGAAAAA GGGGGGAATG
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5551 AAAGACCCCA CTGTAGGTT TGGCAAGCTA GCTTAAGTAA CGCCATTTTG
TTTCTGGGGT GGACATCCAA ACCGTTGAT CGAATTCATT GCGGTAAAC

5601 CAAGGCATGG AAAAATACAT AACTGAGAAT AGAGAAGTTC AGATCAAGGT
GTTCCGTACC TTTTATGTA TTGACTCTTA TCTCTCAAG TCTAGTTCCA

5651 CAGGAACAGA TGGAACAGCT GAATATGGGC CAAACAGGAT ATCTGTGGTA
GTCCTTGTCT ACCTTGTGTA CTTATACCG GTTTGTCTTA TAGACACCAT

5701 AGCAGTTCCT GCCCGGGCTC AGGGCCAAGA ACAGATGGAA CAGCTGAATA
 TCGTCAAGGA CGGGGCCGAG TCCCGGTTCT TGTCTACCTT GTCGACTTAT

 5751 TGGGCCAAAC AGGATATCTG TGGTAAGCAG TTCCTGCCCC GGCTCAGGGC
 ACCCGGTTTG TCCTATAGAC ACCATTCTGC AAGGACGGGG CCGAGTCCCC

 5801 CAAGAACAGA TGGTCCCCAG ATGCGGTCCA GCCCTCAGCA GTTCTAGAG
 GTTCTTGTCT ACCAGGGGTC TACGCCAGGT CGGGAGTCGT CAAAGATCTC

 5851 AACCATCAGA TGTTTCCAGG GTGCCCAAG GACCTGAAAT GACCCTGTGC
 TTGGTAGTCT ACAAAGGTCC CACGGGGTTC CTGGACTTTA CTGGGACACG

 5901 CTTATTTGAA CTAACCAATC AGTTCGCTTC TCGCTTCTGT TCGCGCGCTT
 GAATAAACTT GATTGGTTAG TCAAGCGAAG AGCGAAGACA AGCGCGCGAA

 5951 CTGCTCCCCG AGCTCAATAA AAGAGCCAC AACCCCTCAC TCGGGGCGCC
 GACGAGGGGC TCGAGTTATT TTCTCGGGTG TTGGGGAGTG AGCCCGCGG

 6001 AGTCCTCCGA TTGACTGAGT CGCCCGGTA CCCGTGTATC CAATAAACC
 TCAGGAGGCT AACTGACTCA GCGGGCCAT GGGCACATAG GTTATTGGG

 6051 TCTTGCACTT GCATCCGACT TGTGGTCTCG CTGTTCTTGG GGAGGGTCTC
 AGAACGTCAA CGTAGGCTGA ACACCAGAGC GACAAGGAAC CCTCCCAGAG

 6101 CTCTGAGTGA TTGACTACCC GTCAGCGGGG GTCTTTCATT CATGCAGCAT
 GAGACTCACT AACTGATGGG CAGTCGCCCC CAGAAAGTAA GTACGTCTGA

 6151 GTATCAAAAT TAATTGGTT TTTTCTTCA AGTATTTACA TTAAATGGCC
 CATAGTTTAA ATTAACCAA AAAAAAGAA TCATAAATGT AATTACCGG

 6201 ATAGTTGCAT TAATGAATCG GCCAACGCGC GGGGAGAGGC GGTTCGCTA
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 6251 TTGGCGCTCT TCCGCTTCCT CGCTCACTGA CTCGCTGCGC TCGGTGCTTC
 AACCGCGAGA AGGCGAAGGA GCGAGTGACT GAGCGACGCG AGCCAGCAAG

 6301 GGTGCGGCGC AGCGGTATCA GCTCACTCAA AGGCGGTAAT ACGGTTATCC
 CCGACGCGGC TCGCCATAGT CGAGTGAGTT TCCGCCATTA TGCCAATAGG

 6351 ACAGAATCAG GGGATAACGC AGGAAGAAGC ATGTGAGCAA AAGGCCAGCA
 TGTCTTAGTC CCTATTGCG TCCTTCTTG TACACTCGTT TTCCGGTCGT

 6401 AAAGGCCAGG AACCGTAAAA AGGCCGCGTT GCTGGCGTTT TTCCATAGGC
 TTTCGGGTCC TTGGCATTTT TCCGGCGCAA CGACCGCAA AAGGTATCCG

 6451 TCCGCCCCCC TGACGAGCAT CACAAAATC GACGCTCAAG TCAGAGGTGG
 AGGCGGGGGG ACTGCTCGTA GTGTTTTAG CTGCGAGTTC AGTCTCCACC

 6501 CGAAACCCGA CAGGACTATA AAGATACCAG GCGTTTCCCC CTGGAAGCTC
 GCTTTGGGCT GTCCTGATAT TTCTATGGTC CGCAAAGGGG GACCTTCGAG

 6551 CCTCGTGCGC TCTCTGTTT CGACCTGCC GCTTACCGGA TACCTGTCCG
 GGAGCACGCG AGAGGACAAG GCTGGGACGG CGAATGGCCT ATGGACAGGC

 6601 CCTTCTCCCT TTCGGGAAGC GTGGCGCTTT CTCATAGCTC ACGCTGTAGG
 GGAAAGAGGG AAGCCCTTCG CACCGCGAAA GAGTATCGAG TGCGACATCC

6651 TATCTCAGTT CGGTGTAGGT CGTTCGCTCC AAGCTGGGCT GTGTGCACGA
ATAGAGTCAA GCCACATCCA GCAAGCGAGG TTCGACCCGA CACACGTGCT

6701 ACCCCCCGTT CAGCCCCGACC GCTGCGCCTT ATCCGGTAAC TATCGTCTTG
TGGGGGGCAA GTCGGGCTGG CGACGCGGAA TAGGCCATTG ATAGCAGAAC

6751 AGTCCAAACC GGTAAGACAC GACTTATCGC CACTGGCAGC AGCCACTGGT
TCAGGTGGGG CCATTCTGTG CTGAATAGCG GTGACCGTCG TCGGTGACCA

6801 AACAGGATTA GCAGAGCGAG GTATGTAGGC GGTGCTACAG AGTTCTTGAA
TTGTCCTAAT CGTCTCGCTC CATACATCCG CCACGATGTC TCAAGAACTT

6851 GTGGTGGCCT AACTACGGCT ACACTAGAAG AACAGTATTT GGTATCTGCG
CACCACCGGA TTGATGCCGA TGTGATCTTC TTGTCATAAA CCATAGACGC

6901 CTCTGCTGAA GCCAGTTACC TTCGGAAAAA GAGTTGGTAG CTCTTGATCC
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6951 GGCAAAACAAA CCACCGCTGG TAGCGGTGGT TTTTGTGTT GCAAGCAGCA
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7001 GATTACGCGC AGAAAAAAG GATCTCAAGA AGATCCTTTG ATCTTTTCTA
CTAATGCGCG TCTTTTTC CTAGAGTTCT TCTAGGAAAC TAGAAAAGAT

7051 CGGGGTCTGA CGCTCAGTGG AACGAAAAC CACGTTAAGG GATTTTGGTC
GCCCCAGACT GCGAGTCACC TTGCTTTTGA GTGCAATTCC CTAAAACCAG

7101 ATGAGATTAT CAAAAGGAT CTTACCTAG ATCCTTTTGC GGCCGCAAT
TACTCTAATA GTTTTCTTA GAAGTGGATC TAGGAAACG CCGGCGTTTA

7151 CAATCTAAAG TATATATGAG TAACTTGGT CTGACAGTTA CCAATGCTTA
GTTAGATTIC ATATATACTC ATTTGAACCA GACTGTCAAT GGTACGAAT

7201 ATCAGTGAAG CACCTATCTC AGCGATCTGT CTATTTCGTT CATCCATAGT
TAGTCACTCC GTGGATAGAG TCGTAGACA GATAAAGCAA GTAGGTATCA

7251 TGCCTGACTC COCGTCGTGT AGATAACTAC GATACGGGAG GGCTTACCAT
ACGGACTGAG GGGCAGCACA TCTATTGATG CTATGCCCTC CCGAATGGTA

7301 CTGGCCCCAG TGCTGCAATG ATACCGCGAG ACCCAGCTC ACCGGCTCCA
GACCGGGGTC ACGACGTTAC TATGGCGCTC TGGGTGCGAG TGGCCGAGGT

7351 GATTTATCAG CAATAAACCA GCCAGCCGGA AGGGCCGAGC GCAGAAGTGG
CTAAATAGTC GTTATTGGT CGGTGCGCCT TCCCGGCTCG CGTCTTCACC

7401 TCCTGCAACT TTATCCGCCT CCATCCAGTC TATTAATTGT TGCCGGGAAG
AGGACGTTGA AATAGGCGGA GGTAGGTCAG ATAATTAACA ACGGCCCTTC

7451 CTAGAGTAAG TAGTTCGCCA GTTAATAGTT TGCGCAACGT TGTTGCCATT
GATCTCATT ATCAAGCGGT CAATTATCAA ACGCGTTGCA ACAACGGTAA

7501 GCTACAGGCA TCGTGGTGTG ACGCTCGTCG TTTGGTATGG CTTCATTCAG
CGATGTCCGT AGCACCACAG TCGGAGCAGC AAACCATACC GAAGTAAGTC

7551 CTCGGGTTCC CAACGATCAA GGCGAGTTAC ATGATCCCCC ATGTTGTGCA
GAGGCCAAGG GTTGCTAGTT CCGCTCAATG TACTAGGGGG TACAACACGT

7601 AAAAAGCGGT TAGCTCCTTC GGTCTCCGA TCGTTGTGAG AAGTAAGTTG
TTTTTCGCCA ATCGAGGAAG CCAGGAGGCT AGCAACAGTC TTCATTCAAC

7651 GCCGCAGTGT TATCACTCAT GGTATGGCA GCACTGCATA ATTCTCTTAC
CGGCGTCACA ATAGTGAGTA CCAATACCGT CGTGACGTAT TAAGAGAATG

7701 TGTGATGCCA TCCGTAAGAT GCTTTTCTGT GACTGGTGAG TACTCAACCA
ACAGTACGGT AGGCATTCTA CGAAAAGACA CTGACCACTC ATGAGTTGGT

7751 AGTCATTCTG AGAATAGTGT ATGCGGCGAC CGAGTTGCTC TTGCCCGGCG
TCAGTAAGAC TCTTATCACA TACGCCGCTG GCTCAACGAG AACGGGCGCG

7801 TCAATACGGG ATAATACCGC GCCACATAGC AGAACTTTAA AAGTGCTCAT
AGTTATGCCC TATTATGGCG CGGTGTATCG TCTTGAAATT TTCAAGAGTA

7851 CATTGGAAAA CGTTCTTCGG GCGGAAAACT CTCAGGATC TTACGCGTGT
GTAACCTTTT GCAAGAAGCC CCGCTTTTGA GAGTTCCCTAG AATGGCGACA

7901 TGAGATCCAG TTCGATGTAA CCCACTCGTG CACCCAACTG ATCTTCAGCA
ACTCTAGGTC AAGCTACATT GGGTGAGCAC GTGGGTTGAC TAGAAGTCGT

7951 TCTTTTACTT TCACCAGCGT TTCTGGGTGA GCAAAAACAG GAAGGCAAAA
AGAAAATGAA AGTGGTCGCA AAGACCCACT CGTTTTTGTC CTTCCGTTTT

8001 TGCCGCAAAA AAGGGAATAA GGGCGACACG GAAATGTTGA ATACTCATAC
ACGGCGTTTT TTCCCTTATT CCCGCTGTGC CTTTCAACT TATGAGTATG

8051 TCTTCCTTTT TCAATATTAT TGAAGCATT ATCAGGGTTA TTGTCTCATG
AGAAGGAAAA AGTTATAATA ACTTCGTAAA TAGTCCCAAT AACAGAGTAC

8101 AGCGGATACA TATTGAATG TATTTAGAAA AATAAACAAA TAGGGGTTCC
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8151 GCGCACATTT C
CGCGTGTAAG G

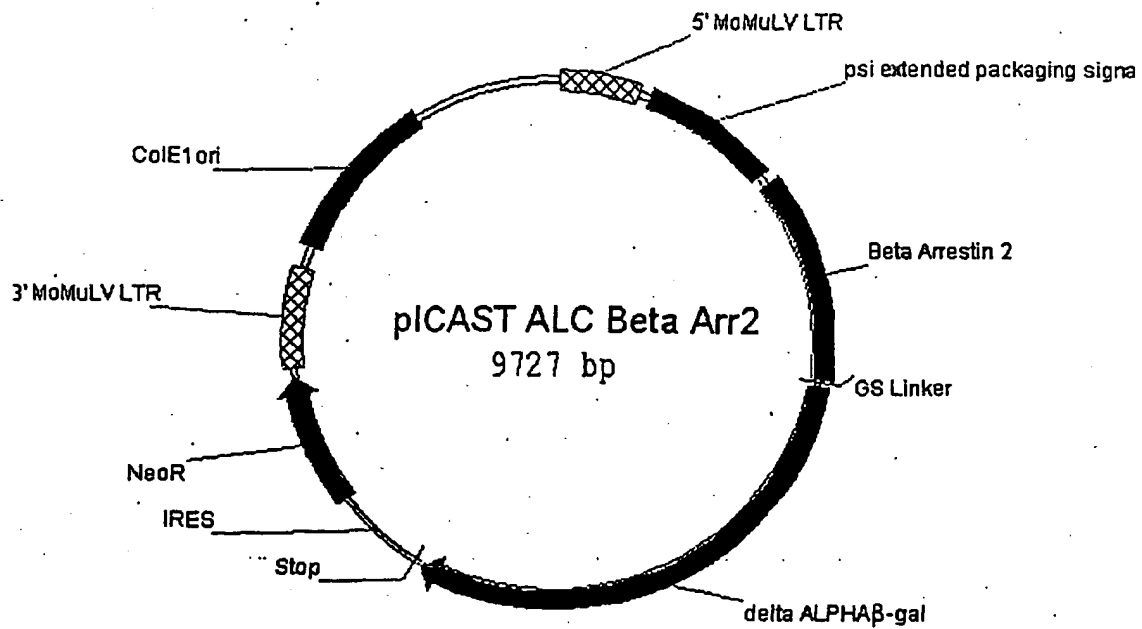


Figure 14

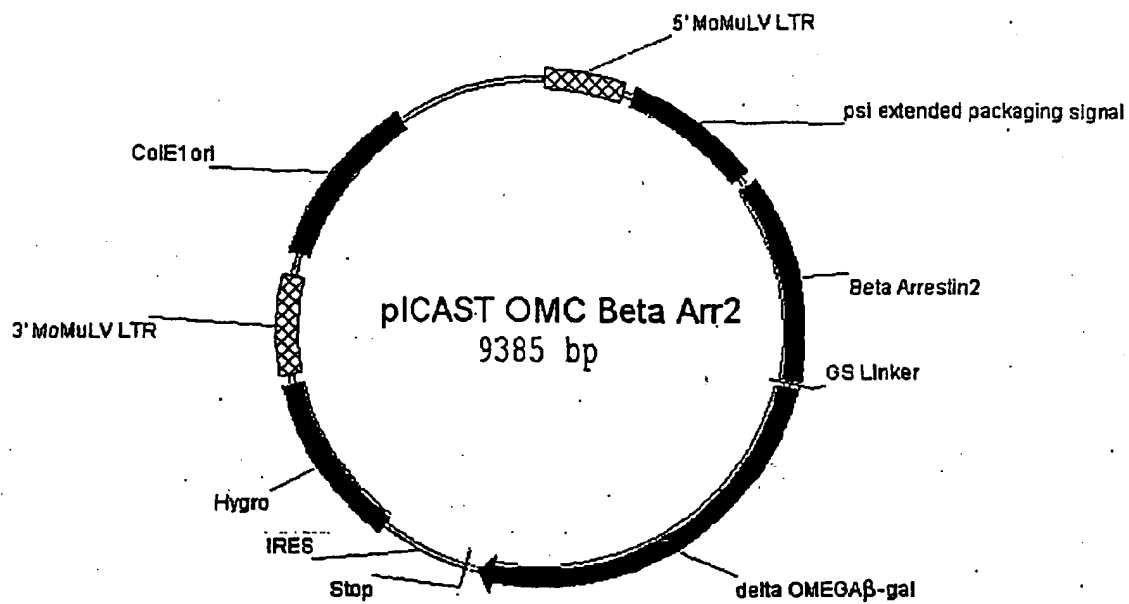


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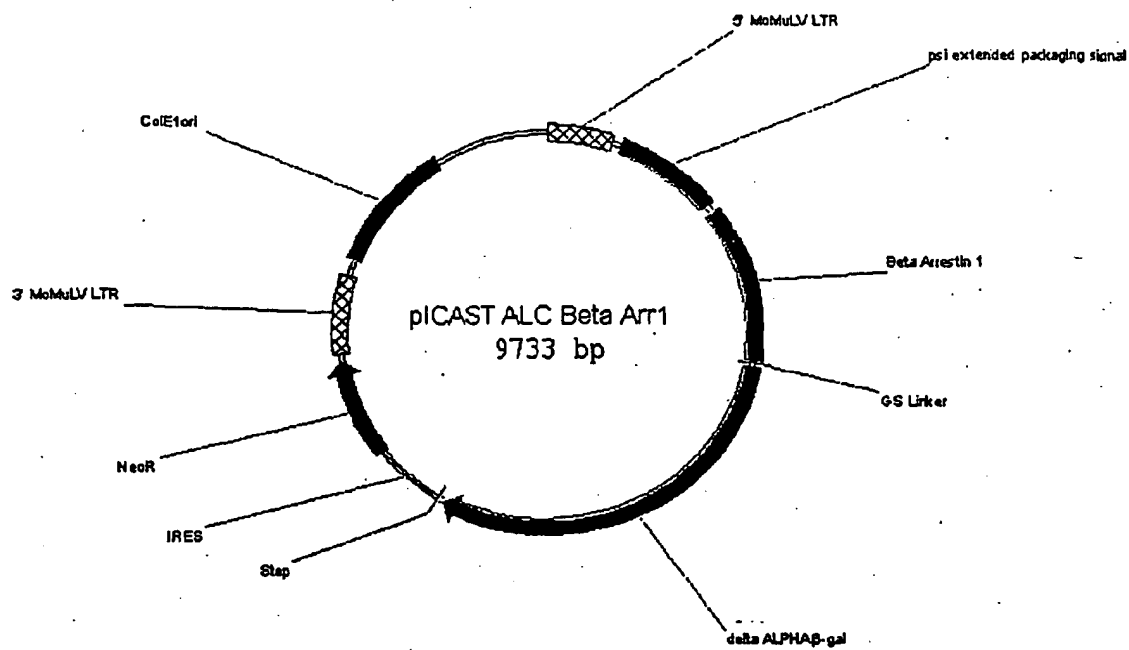


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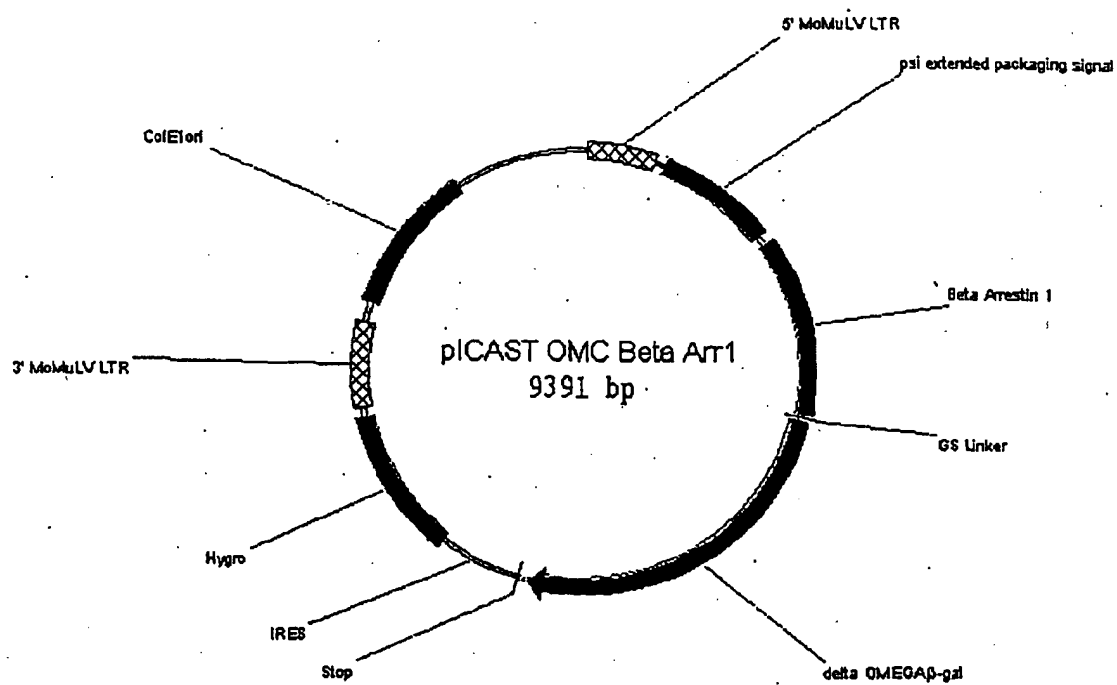


Figure 17.

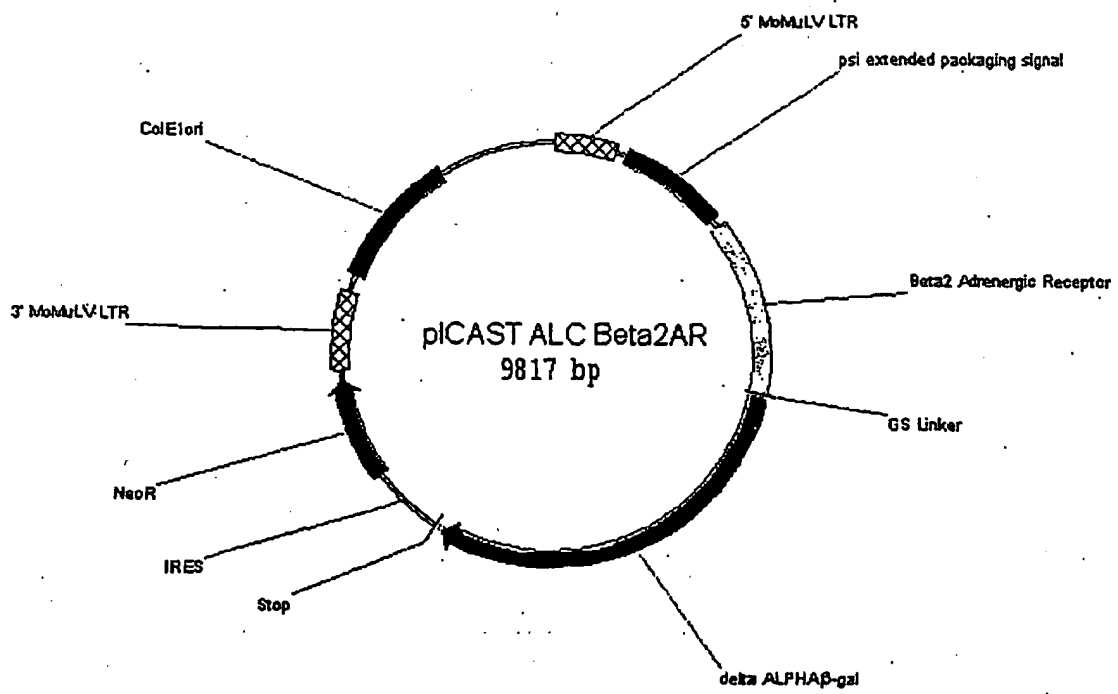


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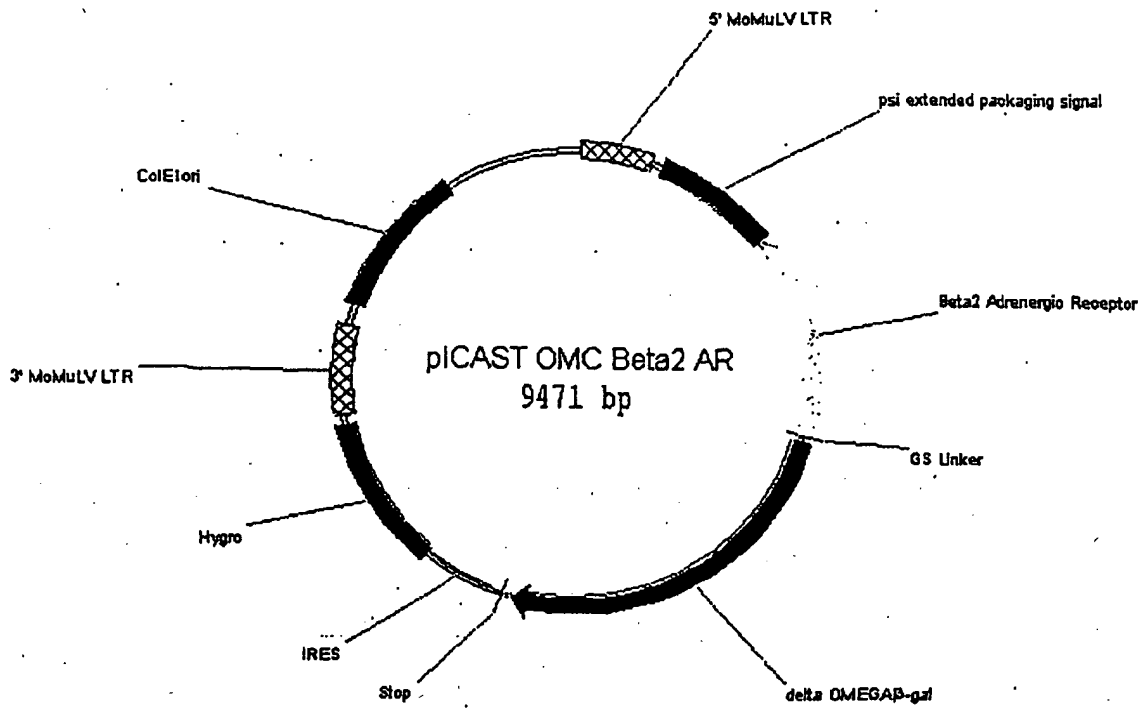


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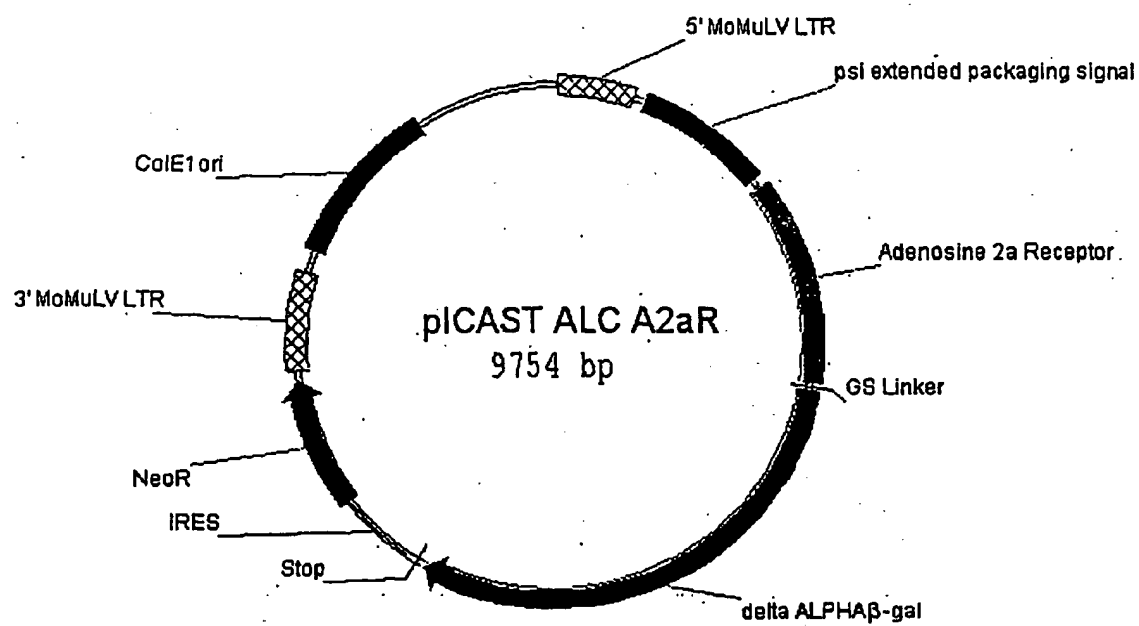


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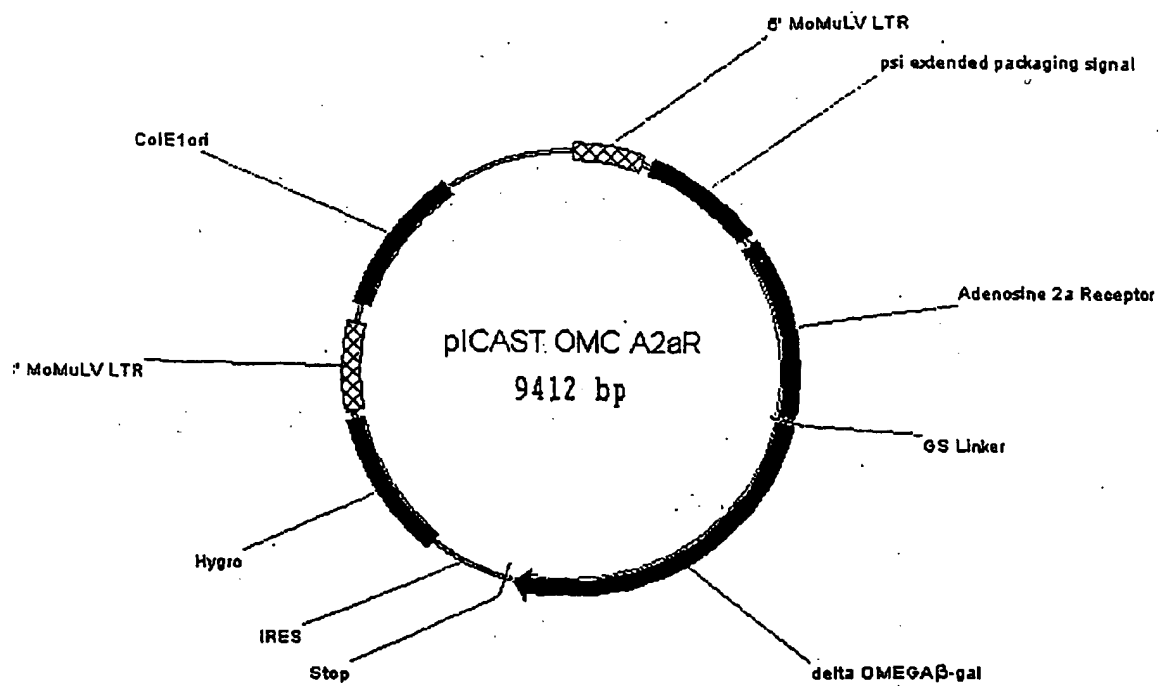


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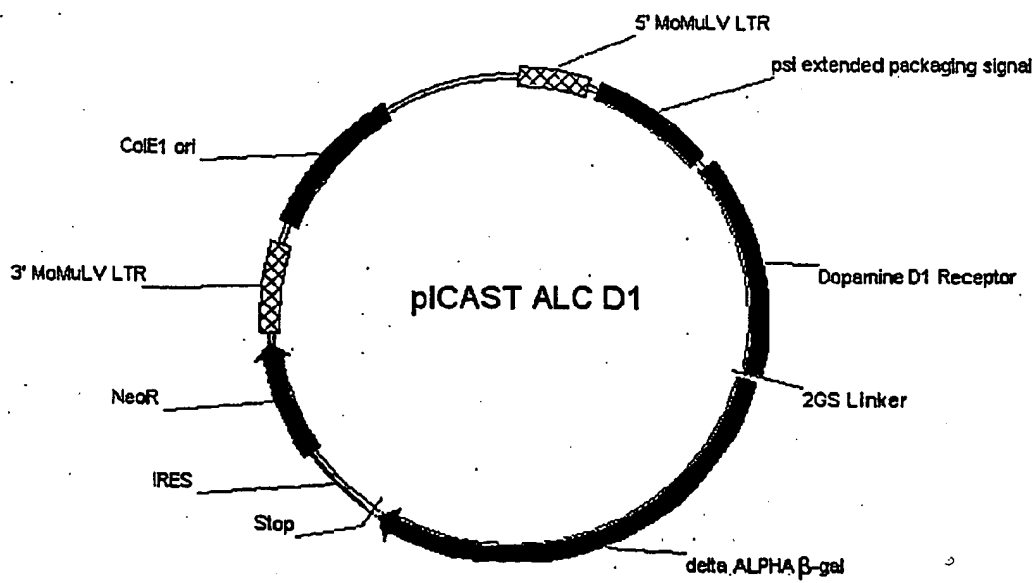


Figure 22

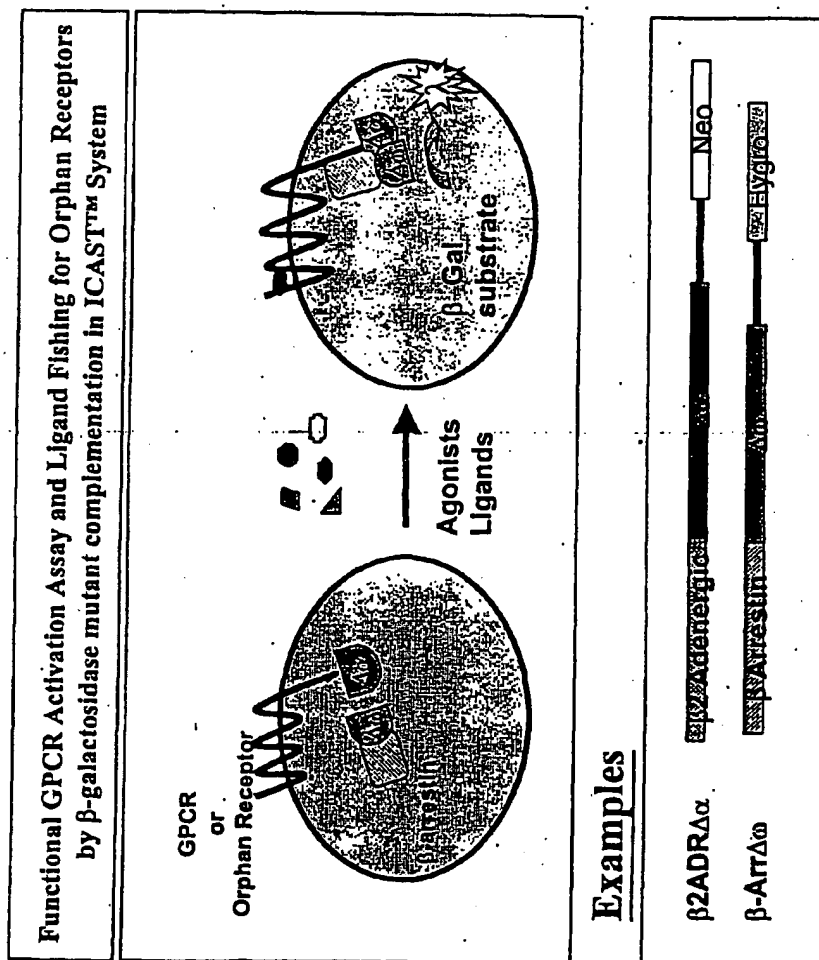
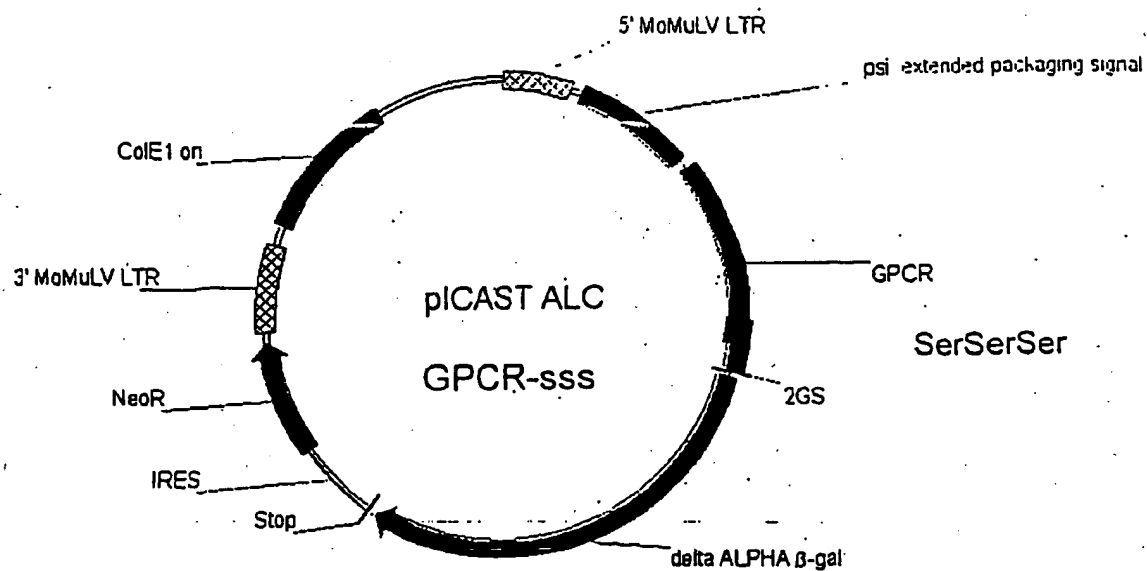
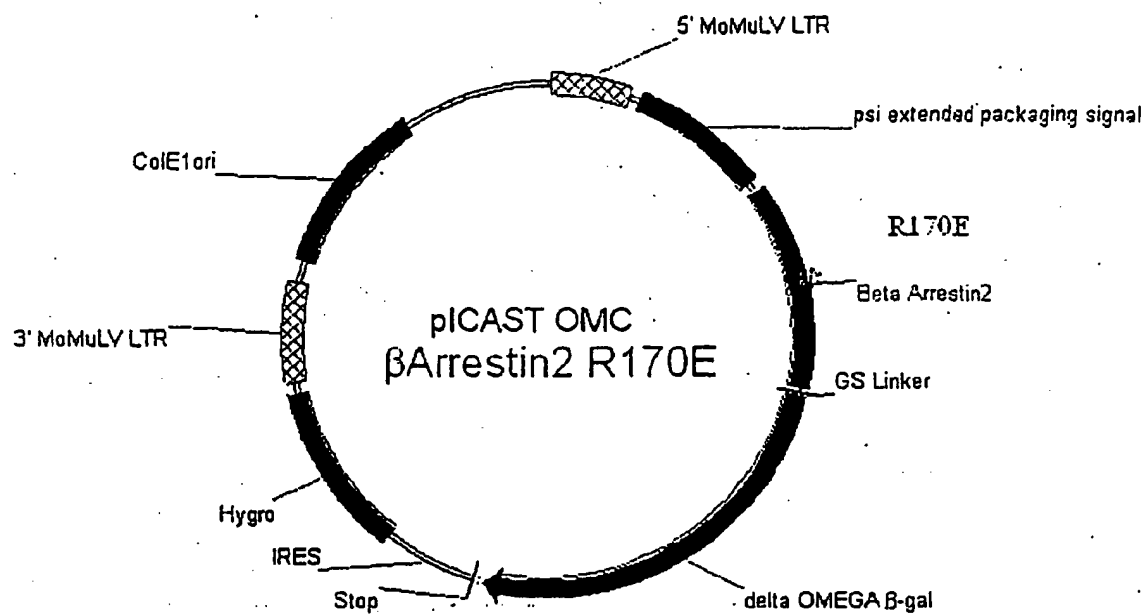


Figure 23



Vector for Expression of a GPCR with inserted
Serine/Threonine amino acid sequences as a fusion with β -gal $\Delta\alpha$.

FIGURE 24



Vector for Expression of mutant (R170E) β -arrestin2 as a fusion with β -gal $\Delta\omega$.

FIGURE 25

Phosphorylation Insensitive Mutant R170E β -Arrestin2 $\Delta\omega$
Binds to β_2 AR $\Delta\alpha$ in Response to Agonist Activation

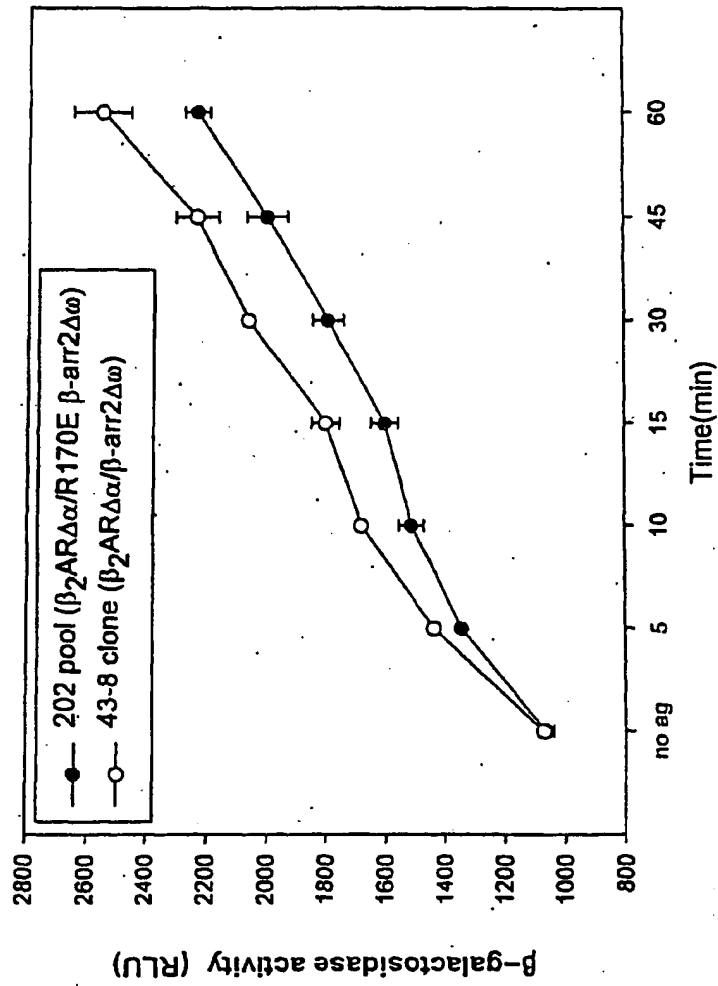
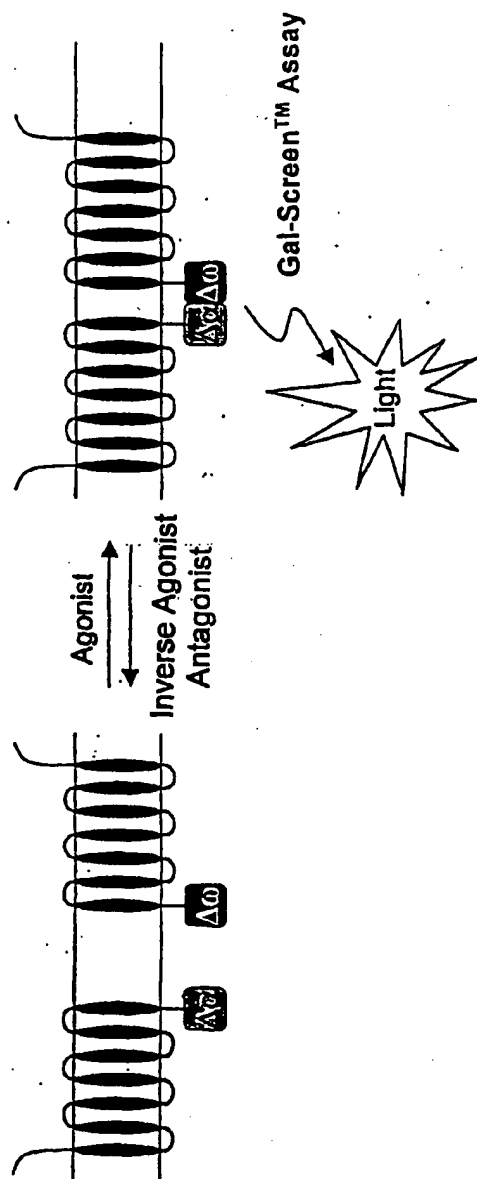


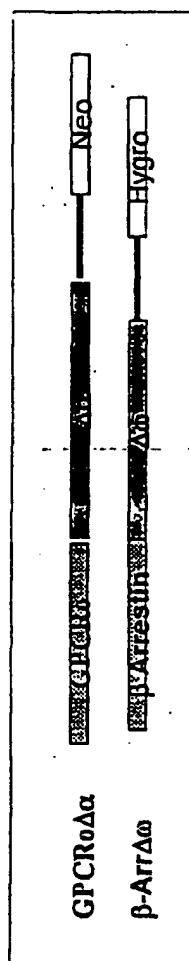
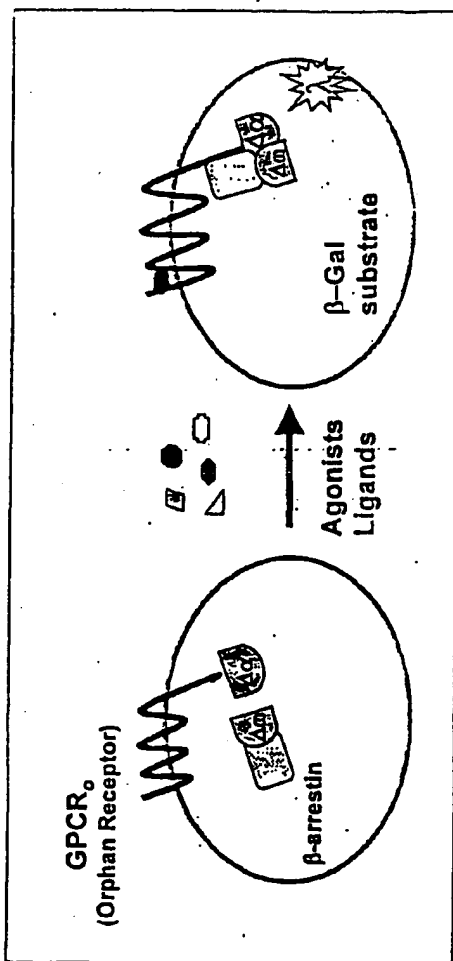
FIGURE 26



GPCR dimerization measured by β -gal complementation

FIGURE 27

Example-



Ligand Fishing for Orphan Receptors by β-galactosidase mutant complementation in ICASTM System

FIGURE 28